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Microfluidic chips for microbially induced calcium carbonate precipitation: Advantages, challenges, and insights

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Abstract: Microbially induced calcium carbonate precipitation (MICP) has garnered significant attention as a biomineralization process with diverse applications spanning from construction to environmental remediation. To propel MICP research and deepen our comprehension of MICP mechanisms, microfluidic chips have emerged as potent tools offering precise control over environmental parameters and real-time observations. Herein, we explore the benefits and challenges associated with employing microfluidic chips as a platform for investigating MICP. The advantages of microfluidic chips lie in their capacity to create controlled microenvironments conducive to emulating specific conditions crucial for MICP. The high-throughput nature of these devices accelerates experimentation by facilitating simultaneous testing of various microbial strains and nutrient compositions. Throughout the MICP process, observations were made on the behaviors of both bacterial cells and CaCO3 cementation. The inherent reduction in reagent consumption offered by microfluidics is both cost-effective and environmentally friendly. However, scaling up from microscale findings to practical applications necessitates careful consideration. Fully replicating the three-dimensional complexity and heterogeneous structures of the soil matrix, which influence microbial behavior, mineral distribution, and overall precipitation dynamics, using microfluidic chips remains challenging. Additionally, certain environmental complexities, including macroscopic soil components such as organic matter and various particle types, which significantly affect microbial activities and mineral precipitation patterns, may be difficult to replicate in microfluidic setups. However, microfluidic chips stand as invaluable tools for advancing MICP research. By addressing the advantages and disadvantages outlined here, researchers can harness the capabilities of microfluidic systems to unravel the intricacies of MICP, ultimately bridging the gap between fundamental understanding and real-world applications.

1.Introduction

Microbially Induced Calcium Carbonate Precipitation (MICP) stands out as a remarkable biotechnological process, emerging at the intersection of microbiology and geotechnical engineering. Understanding the primary mechanisms underlying MICP is pivotal for its effective application. This process relies on the metabolic activities of microorganisms, often specifically selected for their ability to produce enzymes such as urease [1]. These enzymes catalyze the hydrolysis of urea, releasing carbonate ions (CO_3^{2-}) that subsequently react with calcium ions (Ca^{2+}) in the surrounding environment. This chemical interplay ultimately leads to the precipitation of calcium carbonate, binding soil particles, and reinforcing geological structures [2].

$$CO(NH_2)_2 + 2H_2O \xrightarrow{\text{urease}} 2NH_3 + H_2CO_3$$
(1)

$$2NH_3 + 2H_2O \rightarrow 2NH_4^+ + 2OH^-$$
⁽²⁾

$$HCO_3^- + OH^- \rightarrow CO_3^{2-} + H_2O$$
 (3)

$$Ca^{2+} + CO_3^{2-} \to CaCO_3 \tag{4}$$

The significance of MICP spans a diverse spectrum of applications, making it a subject of paramount importance in various fields. In the realm of geotechnical engineering, MICP has found utility in slope stabilization, ground reinforcement, and mitigation of liquefaction in seismic-prone regions [3–8]. Its ability to enhance soil strength without compromising permeability renders MICP an appealing choice for soil improvement projects. Furthermore, it has proven instrumental in repairing soil cracks and restoring historical structures and monuments [9,10].

MICP is a multifaceted biogeochemical process driven by microbial metabolism, mineral precipitation, environmental factors, microbial interactions, transport phenomena, temporal dynamics, spatial heterogeneity, and practical applications, [11–15] offering valuable advantages for effectively studying this complexity. Traditional MICP experimental methods typically involve post-process testing of the final mechanical and permeability properties of samples to detect the MICP transport reaction, lacking real-time dynamic detection capability. Traditional methods also fall short in elucidating the microscale mechanisms of MICP, making it challenging to visualize processes such as bacterial migration, distribution, and proliferation, as well as the nucleation, growth, and crystalline changes of calcium carbonate. Microfluidic systems confine liquids within channels ranging from microns to millimeters, thereby facilitating operation, flow, visualization, and analysis at the micron scale. Consequently, leveraging microfluidic technology to investigate MICP processes holds tremendous potential. Microfluidics enables real-time visualization of MICP processes, providing insights into temporal and spatial dynamics [16,17]. It also allows precise control over environmental conditions, facilitating the investigation of factors such as pH, temperature, and nutrient gradients on MICP. Furthermore, microfluidics operates at a microscale, making it an ideal tool for replicating and studying MICP processes at an appropriate scale. It generates highly detailed and precise data, [18]helping researchers unravel the intricacies of MICP mechanisms and interactions, and encouraging interdisciplinary collaboration among microbiologists, engineers,

and geoscientists, thereby facilitating a comprehensive understanding of MICP complexity and its practical applications.

2.Background of Microfluidics

Microfluidics is a highly versatile and interdisciplinary field that involves the science and technology of manipulating and controlling fluids in channels or devices with dimensions ranging from micrometers to millimeters [19]. It has rapidly evolved from its early beginnings and has found applications in diverse research domains. Initially, microfluidics was not conceived for the study of MICP but rather emerged to address fundamental challenges in various fields [20,21]. The history of microfluidics can be traced back to the early 1960s when researchers began exploring the potential of small-scale fluid manipulation. The advent of microfluidic devices [22,23]. The semiconductor industry, played a pivotal role in the development of microfluidic devices [22,23]. The seminal work of George A. Whitpical et al. in the late 1980s and the early 1990s laid the foundation for microfluidics by demonstrating the capabilities of microscale channels and their potential for a wide range of applications [24,25].

Over the years, microfluidics has found relevance in numerous research fields. In analytical chemistry, it revolutionized sample handling, mixing, and analysis, enabling high-throughput screening and miniaturized laboratory-on-a-chip systems [26,27]. In biology, microfluidic devices have been instrumental in studying cell behavior, single-cell analysis, and DNA sequencing [28–30]. Additionally, microfluidics has made significant contributions to the fields of materials science, physics, and even environmental monitoring [31,32].

While microfluidics was originally developed for different applications, its precision and control over fluid dynamics have made it a valuable tool for investigating complex processes, such as MICP. Researchers have adopted microfluidic systems to simulate and analyze the intricate mechanisms of MICP in controlled laboratory settings [33,34]. These microscale platforms offer a unique advantage for studying how microorganisms interact with calcium ions, urea hydrolysis, and carbonate precipitation [35–37]. By providing a controlled environment, microfluidic experiments can yield valuable insights into the kinetics and factors influencing MICP, thereby enhancing our understanding of this biogeochemical process. Its precise control over fluids has opened up new avenues for studying complex phenomena, including MICP, contributing to advancements in various scientific disciplines.

3.Advantage of using microfluidics to study MICP

Microfluidics represents a powerful and versatile tool for investigating MICP mechanisms. Its precise control, real-time visualization capabilities, scalability, reproducibility, and high-throughput screening potential have revolutionized MICP research [38–40]. By leveraging these advantages, researchers can unlock the full potential of MICP for diverse applications ranging from soil stabilization to environmental remediation and contribute to the advancement of biogeochemical engineering. Over the last five years, the technology for utilizing microfluidic chips to study MICP has seen significant development. Wang et al. [37][18] pioneered the use of microfluidic chips in MICP studies to observe bacterial behavior and patterns of crystal change. Subsequently, various types of microfluidic chips have been gradually introduced into MICP studies, including droplet microfluidics [41], soil particle-filled microfluidic chips [42] and long-distance (1-meter) microfluidic chips (Figure 1).

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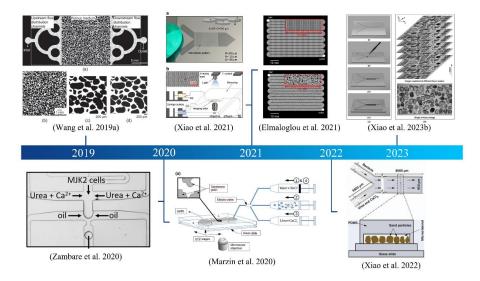


Figure 1. History of microfluidic chips applied to MICP research.

Microfluidic devices provide an unparalleled level of control compared to traditional MICP research methods. Researchers can precisely manipulate factors such as fluid flow rates, reactant concentrations, and environmental parameters like pH and oxygen levels [39,43]. This control enables the creation of well-defined microenvironments that closely mimic natural conditions, facilitating accurate and controlled MICP experiments. An advantage of microfluidics is its compatibility with real-time visualization techniques, including microscopy. Within microfluidic channels, researchers can observe and record MICP processes as they unfold at the microscale level. This dynamic visualization not only offers valuable insights into the temporal and spatial aspects of MICP but also enables the monitoring of microbial activity, mineral nucleation, and growth [34,43,44]. Microfluidic systems inherently operate at the microscale, aligning with the dimensions of MICP processes. This feature allows for the detailed investigation of microbial behavior, urea hydrolysis, and mineral precipitation on the same scale as they occur in natural settings [35,44]. Microfluidic platforms can be adapted for high-throughput screening of various microbial strains, environmental conditions, and reactant combinations [17]. This accelerates the discovery process by enabling researchers to rapidly identify the most efficient MICP-inducing conditions and microbial strains [35]. Optimizing MICP parameters is crucial for practical applications, and microfluidic devices facilitate this optimization by simulating diverse environmental conditions relevant to MICP applications. Researchers can recreate the complexities of soil, groundwater, and subsurface reservoirs to closely mimic real-world scenarios [45,46]. This capacity for environmental simulations provides critical insights into the applicability and adaptability of MICP in geotechnical and environmental settings [47]. With the help of microfluidics, researchers have gained highresolution insights into the intricacies of MICP mechanisms, thereby enhancing our understanding of this process. The mechanisms of MICP that can be investigated include bacterial growth after injection (Figure 2a), bacterial aggregation and detachment after the injection of the cementation solution (Figures 2b and c), the correlation between bacterial distribution and CaCO₃ distribution (Figure 2d), the growth of CaCO₃ crystals with the injection

of the cementation solution (Figure 2e), and the crystal morphology and bacterial traces in the microfluidic chip under a scanning electron microscope (Figure 2f).

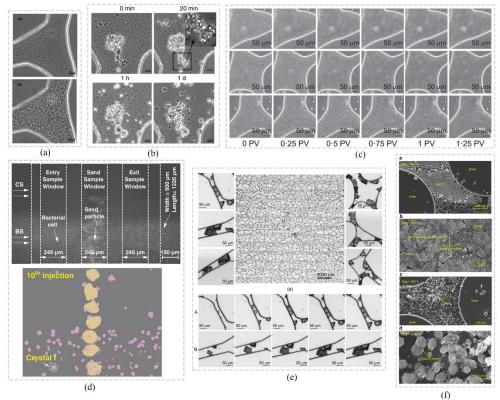


Figure 2. Mechanisms of bacterial behavior and calcium carbonate precipitation revealed by using microfluidic chips: (a) Bacteria continue to grow within 24 h after injection into a microfluidic chip [43]; (b) bacteria aggregation in a microfluidic chip [48]; (c) detachment of bacteria with the injection of cementation solution [18]; (d) distribution characteristics of bacteria and crystals in a microfluidic chip [35];(e) crystal growth with the injection of cementation solution [18]; (f) crystal morphology and bacterial traces in the microfluidic chip under scanning electron microscope [38].

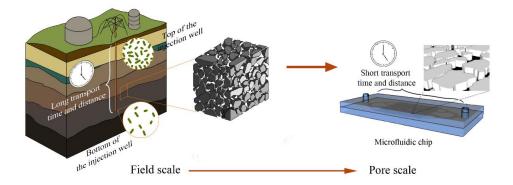
4. Challenges related to MICP study by using microfluidics

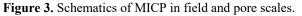
Microfluidics holds tremendous potential for advancing our understanding of MICP mechanisms. However, researchers must navigate various challenges related to microbial compatibility, device design, scaling up, heterogeneity, data interpretation, environmental realism, and long-term studies. Addressing these challenges is essential for harnessing the full potential of MICP in geotechnical and environmental applications, offering innovative solutions for soil stabilization, groundwater remediation, and more.

While microfluidic studies provide valuable insights into MICP mechanisms, one of the primary challenges is the significant scale difference between microfluidic devices and the natural environment where MICP occurs. Microfluidic channels are typically on the order of micrometers to millimeters, whereas MICP processes in the field take place at a much larger scale, often involving soil volumes measured in cubic meters [49,50]. This scale discrepancy can result in differences in fluid dynamics, transport phenomena, and reaction rates [51]. For example, the large-scale distribution of bacteria is affected by transportation distance, time, and complex geological structures, which cannot be easily simulated using microfluidic chip

experiments (Figure 3). Bridging the gap between microscale experiments and field-scale implementation necessitates careful consideration of factors such as material compatibility, microbial growth, and environmental conditions.

Microfluidic studies of MICP often involve the use of specific microbial strains. Challenges arise in maintaining microbial viability within microchannels because the confined environments may differ significantly from natural habitats. Ensuring compatibility between microbial requirements and microfluidic conditions is crucial for obtaining representative MICP results. Additionally, in natural environments, MICP processes often involve diverse microbial communities with intricate interactions [49,53]. For practical reasons, microfluidic studies may use simplified microbial populations or monocultures, which can neglect the complex interplay among microorganisms, potentially leading to deviations in MICP behavior.





Microfluidic devices often simplify [34] the complexity of natural MICP environments. Soil properties, such as grain size distribution, mineral composition, and porosity, can vary widely in natural settings. Achieving realistic heterogeneity in microfluidic setups can be challenging because natural environments exhibit significant variability in factors such as pH, nutrient availability, and microbial populations [54]. Researchers must carefully design microfluidic experiments to capture this heterogeneity realistically. However, microfluidic studies can sometimes lack full environmental compatibility, potentially leading to deviations from real-world MICP processes. For example, natural MICP environments in soil or groundwater exhibit spatial and temporal heterogeneity in parameters such as pH, nutrient concentrations, and microbial populations. In contrast, microfluidic studies may oversimplify these factors, leading to deviations from the dynamic and heterogeneous conditions found in real-world environments. Ensuring that microfluidic experiments accurately replicate the environmental conditions relevant to MICP applications is crucial for their practicality.

Many microfluidic experiments are conducted over relatively short timeframes due to practical constraints. In field MICP processes, MICP can occur over longer periods, potentially spanning months or even years [57,58]. The differences in temporal scales can impact the kinetics and efficiency of MICP and may not accurately represent real-world scenarios.

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5.Conclusions

This article highlights the critical role of microfluidics in advancing our understanding of MICP. Microfluidic systems offer precise control over environmental conditions, enabling researchers to investigate how factors like pH, temperature, and nutrient gradients influence MICP. Real-time visualization capabilities within microfluidic channels provide insights into the temporal and spatial dynamics, microbial interactions, and mineral nucleation and growth. Furthermore, microfluidics facilitates the high-throughput screening of microbial strains and environmental conditions, expediting the optimization of MICP parameters for practical applications. Nonetheless, employing microfluidic technology to investigate MICP still presents several challenges. These challenges encompass bridging the substantial scale disparity between microfluidic experiments and natural MICP environments, ensuring the vitality of microorganisms within microchannels, replicating the intricate microbial communities existing in the natural environment within microfluidic chips, and addressing the temporal-scale differences between microfluidic experiments and field-based experiments, among others.

By addressing these challenges and continuing to innovate in microfluidic design and experimentation, researchers can unlock the full potential of MICP as a transformative biotechnological tool for geotechnical and environmental engineering. Designing microfluidic devices that mimic natural MICP environments while offering precise control is a non-trivial task [35,59]. Achieving the right balance between complexity and simplicity, as well as accommodating different types of experiments, requires careful consideration. To address the current challenges, advancements in the fabrication process of microfluidic chips are required to meet the experimental demands of MICP. Meanwhile, it is essential to integrate microscale experiments with macroscale experiments, wherein microscale mechanisms and macroscale responses are closely interconnected and mutually validated. This integration serves as a bridge between on-site MICP experiments and large-scale MICP applications. Despite these challenges, the integration of microfluidics with MICP research holds immense promise. It offers a pathway for enhancing our understanding of MICP mechanisms and optimizing its applications in soil stabilization, concrete crack repair, and other geotechnical and environmental endeavors.

References

- Mujah D, Shahin M A and Cheng L 2017 State-of-the-art review of biocementation by microbially induced calcite precipitation (MICP) for soil stabilization *Geomicrobiol. J.* 34 524–37
- [2] Martinez B C, DeJong J T, Ginn T R, Montoya B M, Barkouki T H, Hunt C, Tanyu B and Major D 2013 Experimental optimization of microbial-induced carbonate precipitation for soil improvement J. Geotech. Geoenvironmental Eng. 139 587–98
- [3] Gowthaman S, Iki T, Nakashima K, Ebina K and Kawasaki S 2019 Feasibility study for slope soil stabilization by microbial induced carbonate precipitation (MICP) using indigenous bacteria isolated from cold subarctic region SN Appl. Sci. 1 1–16
- [4] Gowthaman S, Mitsuyama S, Nakashima K, Komatsu M and Kawasaki S 2019 Biogeotechnical approach for slope soil stabilization using locally isolated bacteria and inexpensive low-grade chemicals: A feasibility study on Hokkaido expressway soil, Japan Soils Found. 59 484–99
- [5] Gowthaman S, Nakashima K, Nakamura H and Kawasaki S 2020 Influence of wet-dry and freeze-thaw cycles on the physical and mechanical properties of MICP treated slope soil ARMA US Rock Mechanics/Geomechanics Symposium (ARMA) p ARMA-2020

doi:10.1088/1755-1315/1337/1/012039

[6] Gowthaman S, Nakashima K and Kawasaki S 2020 Freeze-thaw durability and shear responses of cemented slope soil treated by microbial induced carbonate precipitation *Soils Found*. 60 840–55

1337 (2024) 012039

- [7] Van Paassen L A 2011 Bio-mediated ground improvement: from laboratory experiment to pilot applications *Geo-frontiers 2011: advances in geotechnical* engineering pp 4099–108
- [8] van Paassen L A, Ghose R, van der Linden T J M, van der Star W R L and van Loosdrecht M C M 2010 Quantifying biomediated ground improvement by ureolysis: large-scale biogrout experiment J. Geotech. geoenvironmental Eng. 136 1721–8
- [9] Le Metayer-Levrel G, Castanier S, Orial G, Loubière J-F and Perthuisot J-P 1999 Applications of bacterial carbonatogenesis to the protection and regeneration of limestones in buildings and historic patrimony *Sediment. Geol.* 126 25–34
- [10] Liu B, Zhu C, Tang C-S, Xie Y-H, Yin L-Y, Cheng Q and Shi B 2020 Bioremediation of desiccation cracking in clayey soils through microbially induced calcite precipitation (MICP) Eng. Geol. 264 105389
- [11] Intarasoontron J, Pungrasmi W, Nuaklong P, Jongvivatsakul P and Likitlersuang S 2021 Comparing performances of MICP bacterial vegetative cell and microencapsulated bacterial spore methods on concrete crack healing *Constr. Build. Mater.* **302** 124227
- [12] Ramakrishnan V, Panchalan R K, Bang S S and City R 2005 Improvement of concrete durability by bacterial mineral precipitation *Proc. ICF* vol 11 (Citeseer) p 9736
- [13] Omoregie A I, Khoshdelnezamiha G, Senian N, Ong D E L and Nissom P M 2017 Experimental optimisation of various cultural conditions on urease activity for isolated Sporosarcina pasteurii strains and evaluation of their biocement potentials *Ecol. Eng.* **109** 65–75
- [14] Qabany A Al and Soga K 2014 Effect of chemical treatment used in MICP on engineering properties of cemented soils *Bio-and Chemo-Mechanical Processes* in Geotechnical Engineering: Géotechnique Symposium in Print 2013 (ICE Publishing) pp 107–15
- [15] Cheng L, Cord-Ruwisch R and Shahin M A 2013 Cementation of sand soil by microbially induced calcite precipitation at various degrees of saturation *Can. Geotech. J.* 50 81–90
- [16] Sun X, Miao L, Tong T and Wang C 2019 Study of the effect of temperature on microbially induced carbonate precipitation Acta Geotech. 14 627–38
- [17] Wang Y, Wang Y, Soga K, DeJong J T and Kabla A J 2023 Microscale investigations of temperature-dependent microbially induced carbonate precipitation (MICP) in the temperature range 4–50 C Acta Geotech. 18 2239–61
- [18] Wang Y, Soga K, Dejong J T and Kabla A J 2019 A microfluidic chip and its use in characterising the particle-scale behaviour of microbial-induced calcium carbonate precipitation (MICP) Géotechnique 69 1086–94
- [19] Beebe D J, Mensing G A and Walker G M 2002 Physics and applications of microfluidics in biology Annu. Rev. Biomed. Eng. 4 261–86
- [20] Nguyen N-T, Wereley S T and Shaegh S A M 2019 Fundamentals and applications of microfluidics (Artech house)
- [21] Sackmann E K, Fulton A L and Beebe D J 2014 The present and future role of microfluidics in biomedical research *Nature* 507 181–9
- [22] Harrison D J, Fluri K, Seiler K, Fan Z, Effenhauser C S and Manz A 1993 Micromachining a miniaturized capillary electrophoresis-based chemical analysis system on a chip *Science (80-.)*. 261 895–7
- [23] West J, Becker M, Tombrink S and Manz A 2008 Micro total analysis systems: latest achievements Anal. Chem. 80 4403–19
- [24] Duffy D C, McDonald J C, Schueller O J A and Whitesides G M 1998 Rapid prototyping of microfluidic systems in poly (dimethylsiloxane) Anal. Chem. 70 4974–84
- [25] Martinez A W, Phillips S T, Butte M J and Whitesides G M 2007 Patterned paper as a platform for inexpensive, low-volume, portable bioassays *Angew. Chemie* **119**

doi:10.1088/1755-1315/1337/1/012039

1340-2

- [26] Mitchell M C, Spikmans V and de Mello A J 2001 Microchip-based synthesis and analysis: control of multicomponent reaction products and intermediates *Analyst* 126 24–7
- [27] Zou Z, Jang A, MacKnight E, Wu P-M, Do J, Bishop P L and Ahn C H 2008 Environmentally friendly disposable sensors with microfabricated on-chip planar bismuth electrode for in situ heavy metal ions measurement *Sensors Actuators B Chem.* 134 18–24
- [28] Kopp M U, Mello A J de and Manz A 1998 Chemical amplification: continuous-flow PCR on a chip Science (80-.). 280 1046–8
- [29] Moschou D, Vourdas N, Kokkoris G, Papadakis G, Parthenios J, Chatzandroulis S and Tserepi A 2014 All-plastic, low-power, disposable, continuous-flow PCR chip with integrated microheaters for rapid DNA amplification Sensors Actuators B Chem. 199 470–8
- [30] Murugan D, Bhatia H, Sai V V R and Satija J 2020 P-FAB: a fiber-optic biosensor device for rapid detection of COVID-19 *Trans. Indian Natl. Acad. Eng.* 5 211–5
- [31] Rattanarat P, Dungchai W, Cate D, Volckens J, Chailapakul O and Henry C S 2014 Multilayer paper-based device for colorimetric and electrochemical quantification of metals *Anal. Chem.* 86 3555–62
- [32] Alizadehgiashi M, Gevorkian A, Tebbe M, Seo M, Prince E and Kumacheva E 2018
 3D-Printed Microfluidic Devices for Materials Science Adv. Mater. Technol. 3 1800068
- [33] Wang Y, Konstantinou C, Soga K, Biscontin G and Kabla A J 2022 Use of microfluidic experiments to optimize MICP treatment protocols for effective strength enhancement of MICP-treated sandy soils Acta Geotech. 17 3817–38
- [34] Elmaloglou A, Terzis D, De Anna P and Laloui L 2022 Microfluidic study in a meterlong reactive path reveals how the medium's structural heterogeneity shapes MICP-induced biocementation Sci. Rep. 12 19553
- [35] Xiao Y, He X, Stuedlein A W, Chu J, Matthew Evans T and Van Paassen L A 2022 Crystal growth of MICP through microfluidic chip tests J. Geotech. Geoenvironmental Eng. 148 6022002
- [36] Lauchnor E G, Schultz L N, Bugni S, Mitchell A C, Cunningham A B and Gerlach R 2013 Bacterially induced calcium carbonate precipitation and strontium coprecipitation in a porous media flow system *Environ. Sci. Technol.* 47 1557–64
- [37] Wang Y, Soga K and Jiang N-J 2017 Microbial induced carbonate precipitation (MICP): the case for microscale perspective *Proceedings of the 19th international conference on soil mechanics and geotechnical engineering* pp 1099–102
- [38] Xiao Y, He X, Wu W, Stuedlein A W, Evans T M, Chu J, Liu H, van Paassen L A and Wu H 2021 Kinetic biomineralization through microfluidic chip tests Acta Geotech. 16 3229–37
- [39] Wang Y, Konstantinou C, Tang S and Chen H 2023 Applications of microbialinduced carbonate precipitation: A state-of-the-art review *Biogeotechnics* 100008
- [40] Xiao Y, He X, Ma G, Zhao C, Chu J and Liu H 2023 Biomineralization and mineralization using microfluidics: A comparison study J. Rock Mech. Geotech. Eng.
- [41] Marzin T, Desvages B, Creppy A, Lépine L, Esnault-Filet A and Auradou H 2020 Using microfluidic set-up to determine the adsorption rate of sporosarcina pasteurii bacteria on sandstone *Transp. Porous Media* 132 283–97
- [42] Zambare N M, Naser N Y, Gerlach R and Chang C B 2020 Mineralogy of microbially induced calcium carbonate precipitates formed using single cell drop-based microfluidics Sci. Rep. 10 17535
- [43] Wang Y, Soga K, DeJong J T and Kabla A J 2019 Microscale visualization of microbial-induced calcium carbonate precipitation processes J. Geotech. Geoenvironmental Eng. 145 4019045
- [44] Shu S, Chen H and Meng H 2022 Modelling Microbially Induced Carbonate Precipitation (MICP) in Microfluidic Porous Chips Geofluids 2022
- [45] Xiao Y, Cao B, Shi J, Wu H, He X, Zhao C, Chu J and Liu H 2023 State-of-the-art

doi:10.1088/1755-1315/1337/1/012039

review on the application of microfluidics in biogeotechnology *Transp. Geotech.* 101030

- [46] Kim D, Mahabadi N, Jang J and van Paassen L A 2020 Assessing the kinetics and pore-scale characteristics of biological calcium carbonate precipitation in porous media using a microfluidic chip experiment *Water Resour. Res.* 56 e2019WR025420
- [47] Mortensen B M, Haber M J, DeJong J T, Caslake L F and Nelson D C 2011 Effects of environmental factors on microbial induced calcium carbonate precipitation J. *Appl. Microbiol.* 111 338–49
- [48] Wang Y, Soga K, DeJong J T and Kabla A J 2021 Effects of bacterial density on growth rate and characteristics of microbial-induced CaCO 3 precipitates: Particle-scale experimental study J. Geotech. Geoenvironmental Eng. 147 4021036
- [49] Gomez M G, Anderson C M, Graddy C M R, DeJong J T, Nelson D C and Ginn T R 2017 Large-scale comparison of bioaugmentation and biostimulation approaches for biocementation of sands J. Geotech. geoenvironmental Eng. 143 4016124
- [50] Gomez M G, Martinez B C, DeJong J T, Hunt C E, deVlaming L A, Major D W and Dworatzek S M 2015 Field-scale bio-cementation tests to improve sands *Proc. Inst. Civ. Eng. Improv.* 168 206–16
- [51] Weinhardt F, Deng J, Hommel J, Vahid Dastjerdi S, Gerlach R, Steeb H and Class H 2022 Spatiotemporal Distribution of Precipitates and Mineral Phase Transition During Biomineralization Affect Porosity–Permeability Relationships: Microfluidic investigations *Transp. Porous Media* 143 527–49
- [52] Tang Y, Gan M, Xie Y, Li X and Chen L 2014 Fast screening of bacterial suspension culture conditions on chips Lab Chip 14 1162–7
- [53] Graddy C M R, Gomez M G, DeJong J T and Nelson D C 2021 Native bacterial community convergence in augmented and stimulated ureolytic MICP biocementation *Environ. Sci. Technol.* 55 10784–93
- [54] Cheng L, Shahin M A and Mujah D 2017 Influence of key environmental conditions on microbially induced cementation for soil stabilization J. Geotech. Geoenvironmental Eng. 143 4016083
- [55] Gowthaman S, Nakashima K and Kawasaki S 2021 Durability analysis of biocemented slope soil under the exposure of acid rain J. Soils Sediments 21 2831– 44
- [56] Oliveira P J V, Freitas L D and Carmona J P S F 2017 Effect of soil type on the enzymatic calcium carbonate precipitation process used for soil improvement J. Mater. Civ. Eng. 29 4016263
- [57] Saneiyan S, Ntarlagiannis D, Werkema Jr D D, Colwell F S and Ohan J 2016 Long term monitoring of microbial induced soil strengthening processes AGU Fall Meeting Abstracts vol 2016 pp NS24A-03
- [58] Torres-Aravena A E, Duarte-Nass C, Azócar L, Mella-Herrera R, Rivas M and Jeison D 2018 Can microbially induced calcite precipitation (MICP) through a ureolytic pathway be successfully applied for removing heavy metals from wastewaters? Crystals 8 438

[59] Jiang N-J and Soga K 2017 The applicability of microbially induced calcite precipitation (MICP) for internal erosion control in gravel–sand mixtures *Géotechnique* **67** 42–55