

Strength Behavior of Temperature-Dependent MICP-Treated Soil

Yuze Wang*, Yong Wang, Charalampos Konstantinou

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1 **Strength behavior of temperature-dependent Microbially Induced Carbonate Precipitation**

2 **(MICP)-treated soil**

3
4 Yuze Wang^{a,b*}, Yong Wang^b, Charalampos Konstantinou^c

5
6 a. Shenzhen Key Laboratory of Natural Gas Hydrates, Southern University of Science
7 and Technology, Shenzhen, 518055, China;

8 b. Department of Ocean Science and Engineering, Southern University of Science and Technology,
9 Shenzhen, 518055, China;

10 c. Department of Civil and Environmental Engineering, University of Cyprus, Nicosia, Cyprus

11 Corresponding author: Yuze Wang; email address: wangyz@sustech.edu.cn

12
13 **Abstract**

14 Microbially Induced Carbonate Precipitation (MICP) is a novel soil strengthening technique that
15 involves a bio-geo-chemical process. Temperature plays a crucial role in influencing the biological
16 and chemical processes involved in the formation of carbonate precipitates, which in turn affect the
17 mechanical properties of the treated soil. The aim of this study was to investigate the impact of
18 temperature on the cementing structure of MICP-treated soils and its subsequent effects on their
19 strength parameters. The results revealed that temperature considerably affected the content, size,
20 and distribution of CaCO₃ crystals produced, resulting in variations in the friction angle, cohesion,
21 stiffness, peak strength, residual strength, and dilation of the MICP-treated soil samples. Lower
22 strength enhancement was observed when fewer and smaller carbonate crystals were produced at

23 4°C and 50°C. In contrast, higher numbers of larger crystal clusters were produced at 20°C and
24 35°C, which effectively bonded the soil particles. Increasing the number of bacterial injections at
25 50°C promoted the formation of larger crystals and enhanced strength effectively. This study
26 highlights the temperature effects on calcium carbonate growth in biocemented soils, which is a
27 critical step in determining the field-scale application of this innovative soil stabilization technique.

28

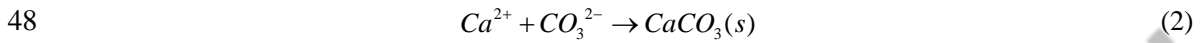
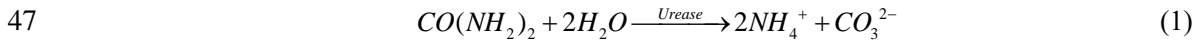
29 **Keywords:** environmental temperature; urease activity; MICP performance; consolidated drained
30 triaxial tests

31

32 **1 Introduction**

33 Microbially Induced Carbonate Precipitation (MICP) is a biochemical process that involves
34 microorganisms inducing an enzyme-catalyzed reaction to increase the supersaturation ratio of
35 carbonate minerals, such as calcium carbonate, and causing their precipitation in an aqueous
36 environment within soil pores or geo-structure cracks (Mitchell and Santamarina 2005; DeJong et
37 al. 2006). The solid surfaces of soil particles or geo-structures provide the nucleation sites for CaCO₃
38 crystal formation. Among the various MICP processes for soil stabilization, ureolysis-driven MICP
39 is the most extensively studied, owing to its high chemical conversion efficiency and ease of control.
40 In this process, ureolytic bacterial cells are introduced into the soil matrix through either bio-
41 augmentation or bio-stimulation (Gomez et al. 2017; Graddy et al. 2021), followed by multiple
42 injections of cementation solution containing CaCl₂, urea and nutrient broth. Ureolysis bacteria
43 express urease enzyme, which catalyzes the hydrolysis of urea (*Equation 1*). The addition of calcium
44 (Ca²⁺) to this system induces the precipitation of calcium carbonate (CaCO₃) as the CO₃²⁻ ions

45 produced by the hydrolysis of urea react with the supplied Ca^{2+} (Equation 2), whilst the addition of
46 nutrient broth to this system maintains the bacterial activity throughout the precipitation period:



49

50 The precipitated CaCO_3 crystals bind the soil particles, increasing soil strength and stiffness, whilst
51 maintaining a relatively high soil permeability, or filling the cracks in geo-structures and reducing
52 their permeability (Wang et al. 2023). Therefore, MICP has many applications such as soil
53 stabilisation (DeJong et al., 2006, 2010; van Paassen et al. 2010; Naveed et al. 2019), production of
54 cemented sandstones (Konstantinou et al. 2021a), improvement of liquefaction resistance (Montoya
55 et al. 2013; Xiao et al. 2018; Darby et al. 2019), control of soil erosion (Jiang et al. 2017; Chek et
56 al. 2021), maintenance of wellbore integrity, fracture sealing (Cuthbert et al. 2013; Phillips et al.
57 2018), and the modification and protection of materials (Tobler et al. 2018). Moreover, MICP has
58 been suggested for producing energy piles (Venuleo et al. 2016; Martinez et al. 2019), preventing
59 the corrosion of marine materials (Guo et al. 2019), separating oil and water (Tang et al. (2021),
60 forming deep foundations (Lin et al. 2016, 2021; Zamani et al. 2021), stabilizing gas hydrate-
61 bearing sediments (Hata et al. 2020), and achieving ocean negative carbon emission (Zhang et al.
62 2022).

63

64 As the use of Microbially Induced Carbonate Precipitation (MICP) expands and its applications
65 extend to various environmental conditions, the impact of factors such as pH, salinity, oxygen level,
66 and temperature on its effectiveness must be considered. Of these, temperature is a critical factor.

67 Despite the fact that various studies have shown that soil engineering properties, such as strength,
68 stiffness, deformation, and creep behavior, are minimally affected by temperature within the range
69 of room temperature to 90°C (Burghignoli et al. 1999; Liu et al. 2018), the impact of environmental
70 temperature on the effectiveness of MICP for soil stabilization is significant. This is mainly because
71 temperature affects the processes involved in MICP and the properties of carbonate crystals, which
72 indirectly impact the engineering behavior of MICP-treated soils. For instance, Cheng et al. (2017)
73 reported that 25°C was more efficient than 4°C and 50°C in generating the largest CaCO₃ crystals
74 and achieving the highest unconfined compressive strength (UCS) of MICP-treated soils. In addition,
75 earlier studies have shown that crystal number and characteristics were considerably influenced by
76 bacterial quantity and crystal growth dynamics, which were highly dependent on temperature in the
77 range of 4-50°C (Wang et al. 2022a).

78
79 Based on these findings, it is crucial to further investigate the impact of treatment temperature on
80 the cementing structure in MICP-treated soils, and how this influences their strength parameters.
81 Given that MICP is employed in a wide range of applications, from stabilization of gas hydrate-
82 bearing sediments where temperatures can range from 3.5 to 14.5°C at the seafloor (Fujii et al., 2015;
83 Li et al., 2018; Ye et al., 2020), to energy piles where temperatures can reach 30°C at shallow depths
84 and 80°C at higher depths, and wellbore integrity and fracture sealing where temperatures can be
85 35-55°C (Toribio et al. 2004) or even reaching or exceed 100°C based on typical geothermal
86 gradients (Tian et al. 2015; Jing et al. 2022) , this study focuses on a series of MICP treatment tests
87 conducted within the temperature range of 4-50°C. To investigate the strength parameters of MICP-
88 treated soils, triaxial tests are essential. Previous studies have examined the strength parameters of

89 soil after treatment at room temperature, such as Montoya and DeJong (2015), Cui et al. (2017),
90 Gao et al. (2019), and Nafisi et al. (2020). However, in this study, MICP is carried out at various
91 temperatures, and triaxial tests are performed to obtain the strength parameters of MICP-treated
92 samples, which was not conducted previously. Additionally, SEM imaging and X-ray diffraction
93 (XRD) analysis are used to analyze the effects of temperature and extra bacterial injections on
94 calcium carbonate crystals and their impact on soil parameters. This study discusses the implications
95 of these findings for resulting engineering properties and treatment protocols for subsurface
96 applications of MICP.

97

98 **2 Materials and methods**

99 **2.1 Sand properties and specimen preparation**

100 In this study, a fine silica sand conforming to the CHINA ISO standard was employed. The sand has
101 an average grain size of 0.125 mm and a coefficient of uniformity, C_u of 5.6, as depicted in Fig.1.
102 Based on the Unified Soil Classification System (ASTM, 2017), the sand is classified as poorly
103 graded. To prepare the samples, split acrylic cylindrical molds with an inner diameter of 38 mm and
104 a height of 80 mm were used (Fig. 2a). The weight of the dry sand was calculated to achieve a
105 targeted relative density (RD) of 50%. The sand was poured into the columns using a dry pluviation
106 method. The sand columns were then flushed multiple times with deionized water to remove any
107 air present until the flow rate stabilized and the specimen became completely saturated. The outlet
108 tube was closed with a drain valve to maintain the sand column's saturation until the biotreatment
109 process was initiated.

110

111 **2.2 Biological suspension and cementation solution**

112 The urease-active strain *S. pasteurii* (CGMCC1.3687) was chosen in this study, as it has
113 demonstrated superior urease activity compared to many other alternative ureolytic bacteria. The
114 strain was cultivated in a NH₄-YE liquid media (ATCC 1366) comprising 20 g/L yeast extract, 10
115 g/L ammonium sulfate, and 0.13 M tris buffer at 30°C for approximately 24 hours in a shaking
116 incubator at 200 rpm/min until it reached an optical density at 600 nm (OD₆₀₀) of 1.0. The bacterial
117 activity of the bacterial suspension tested at OD₆₀₀ of 1.0 was approximately 40 mM/h. The
118 cementation solution used consisted of 0.75 M urea, 0.5 M calcium chloride, and 3 g/L nutrient
119 broth. A urea to calcium chloride ratio greater than one was chosen as previous studies have
120 demonstrated its greater effectiveness (Montoya et al. 2013, 2015; Martinez et al. 2013). All
121 chemical reagents used in this study were of analytical grade.

122

123 **2.3 MICP treatment**

124 A gravity filtration method was used for the MICP treatment process, where injection was performed
125 from the top to the bottom using gravity (see Figure 2a and b). The staged injection method was
126 employed for the MICP treatment, and the MICP treatment parameters are listed in Table 1. To
127 improve bacterial attachment to soil particles, a 24-hour retention period was utilized for Test No.
128 1-4. Additionally, a 24-hour bacterial settling time was chosen to enhance bacterial settlement and
129 distribution homogeneity (Wang 2022a, b; Konstantinou et al. 2021). As bacterial urease activity
130 decreases rapidly at 50°C (Wang et al. 2022a), the number of bacterial suspension (BS) injections
131 at 50°C was increased to 3 and 6, respectively, in groups 5 and 6. Moreover, the time between BS
132 and cementation solution (CS) injections was reduced to 2 hours instead of 24 hours in these two

133 tests. In all tests, 1.0 pore volume (PV) of BS or CS were injected each time, and the total number
134 of CS injections was 6. During the retention times of the experiment, the samples were kept at the
135 specified temperature. To achieve four temperatures (4°C, 20°C, 35°C, and 50°C), the setup was
136 placed in a refrigerator, at room temperature, and in two ovens (see Figure 2c). After completion of
137 the biological treatment process for the soils in the columns, the specimens were flushed with two
138 pore volumes of deionized water to eliminate all excess soluble salts before removing the specimens
139 from the columns following established practices (Whiffin et al. 2007; Dejong et al. 2010).

140

141 **2.4 Triaxial tests**

142 Consolidated drained (CD) triaxial tests (ASTM 2020) were conducted to determine the strength
143 and stress-strain relationships of MICP-treated specimens. In the study by Montoya and DeJong
144 (2015) on MICP-treated sands, samples were prepared by first applying confining pressure,
145 followed by MICP treatment to match field soil conditions. However, in other studies such as Cui
146 et al. (2017) and Gao et al. (2018), samples were treated with MICP first, demolded, and then
147 positioned in the test apparatus before applying confining pressures for triaxial tests. Due to
148 temperature requirements, the procedure of Cui et al. (2017) and Gao et al. (2018) was followed in
149 this study. It should be noted that this procedure may damage some cementation bonds before
150 triaxial shear begins, potentially resulting in lower strength parameters than the true values.

151

152 First, the specimens were saturated with deionized water using a vacuum saturation method for 24
153 hours and then placed in the triaxial apparatus. Hydraulic saturation was applied for 12 hours at a
154 confining pressure of 20 kPa, and back-pressure saturation was sequentially applied until the B-

155 value exceeded 0.95. After consolidation, the confining pressure was maintained, and the specimens
156 were sheared under drained conditions at a constant displacement loading rate of 0.1 mm/min.
157 Stress-strain curves were obtained for each confining pressure to determine peak and residual
158 strength, with the residual strength defined as the stress level at which the yield strength stabilized
159 after a decrease from the peak strength due to accumulated damage (at a strain of 20% in this case).
160 The two key shear strength parameters, friction angle and cohesion strength, were obtained by
161 consolidating each of the four samples at effective confining pressures of 100, 200, 300, and 400
162 kPa based on the Mohr-Coulomb (MC) failure criterion (Wood 1990).

163

164 **2.5 CaCO₃ content measurement and chemical efficiency calculation**

165 After completion of the triaxial tests, samples weighing between 15 to 25 g were collected every 15
166 mm along the height of the sand column. These samples were subjected to oven-drying at 105°C for
167 at least 24 hours to determine their CaCO₃ content using the ASTM Method (ASTM 2014). To
168 perform the measurement, each sample was mixed with 30 ml of 3 M hydrochloric acid inside a
169 sealed chamber, ensuring no contact between samples. The chamber was gently agitated to allow
170 for the reaction between CaCO₃ and hydrochloric acid, which produced CO₂, leading to an increase
171 in pressure within the chamber. The reading was taken when the pressure gauge indicated no further
172 change in pressure, and the amount of CaCO₃ was calculated as follows:

$$173 \quad \text{CaCO}_3 (\text{g}) = 0.034 \cdot \text{CO}_2 \text{ pressure} + 0.0198 \quad (3)$$

174

175 The chemical conversion efficiency of MICP was then calculated, which is defined as the ratio of
176 the precipitated mass of CaCO₃ in the sand to the calculated mass of CaCO₃ from cementation

177 solutions (Wang 2018):

$$178 \quad \text{Efficiency}(\%) = \frac{m(\text{CaCO}_3) / m_1(\text{sand})}{c(\text{CaCl}_2) \cdot V \cdot M(\text{CaCO}_3) / m_2(\text{sand})} \times 100\% \quad (4)$$

179 where, $m(\text{CaCO}_3)/m_1(\text{sand})$ is the measured CaCO_3 content, $c(\text{CaCl}_2)$ is the concentration of CaCl_2
180 in the cementation solutions, V is the total volume of cementation solution injected into samples,
181 $M(\text{CaCO}_3)$ is the molar mass of CaCO_3 (100 g/mol), and $m_2(\text{sand})$ is the dry mass of sand used to
182 prepare sample columns.

183

184 **2.6 Scanning electron microscopy (SEM) imaging and X-ray diffraction (XRD) analyses**

185 After the MICP treatment, samples were oven-dried at 105°C and prepared for scanning electron
186 microscopy (SEM) imaging using a PHENOM XL Scanning Electron Microscope to investigate the
187 microscale properties of the CaCO_3 crystals that formed. Additionally, X-ray diffraction (XRD)
188 analysis was performed on the samples treated at different temperatures using a Regiku Miniflex
189 600 X-ray diffractometer.

190

191 **3 Results and discussion**

192 **3.1 CaCO_3 content and chemical efficiency**

193 In the case of a single bacterial suspension injection (Fig. 3a), the highest average CaCO_3 content
194 was obtained at 35°C (sample T35(BS1) in Fig. 3a), while temperatures of 4°C and 50°C resulted
195 in relatively lower average CaCO_3 contents (samples T4(BS1) and T50(BS1) in Fig. 3a). Injecting
196 bacterial suspension multiple times increases the average CaCO_3 content (Fig. 3b). The distribution
197 of CaCO_3 content along the height of the sand column becomes more heterogeneous as the average
198 CaCO_3 content increases (Fig. 3a and b). Least-squares regression was used to obtain the fit lines

199 of the data. The chemical conversion efficiency of the sample ranges from 17.3% to 67%, with the
200 peak obtained at 35°C (Fig. 3c). At 50°C, the chemical conversion efficiency is logarithmically fitted
201 with the number of BS injections, ranging from 17.3% to 56.5% (Fig. 3d).

202

203 The findings of this study are consistent with those of Wang et al. (2022b), except for the highest
204 CaCO₃ content, which was achieved at 35°C in this study, while Wang et al. (2022b) reported the
205 highest content at 20°C. The difference may be attributed to the variation in the interval between
206 consecutive injections of the cementation solution in the two studies. Wang et al. (2022b) used a 48-
207 hour interval, while this study employed a 24-hour interval, which was sufficient to precipitate 0.5
208 M of CS at both 20°C and 35°C. As bacterial activity decreases over time at higher temperatures,
209 the shorter injection interval maintained a relatively higher bacterial activity and CaCO₃ conversion
210 efficiency, indicating that the injection interval between BS and CS needs to be carefully considered
211 at higher temperatures (e.g., 50°C). Moreover, increasing the number of BS injections compensated
212 for bacterial decay caused by high temperature, maintained the supersaturation state in the solution,
213 and resulted in higher CaCO₃ content than the amount produced when BS was injected only once.
214 Therefore, a 24-hour injection interval is suitable for temperatures below 35°C, while a shorter
215 interval should be adopted at high-temperature conditions to ensure the desired treatment effect.

216

217 **3.2 Stress-strain and volumetric-strain responses**

218 Previous studies by Feng and Montoya (2016) and Xiao et al. (2019) have shown that samples
219 treated with microbial-induced calcium carbonate precipitation (MICP) at room temperature exhibit
220 strain softening behavior. This study confirms that strain softening is not dependent on treatment

221 temperature, as samples treated at temperatures ranging from 4 to 50°C showed similar behavior.
222 However, more pronounced softening was observed at 20°C and 35°C, while gradual softening was
223 observed at 4°C and 50°C after reaching peak strength (Fig. 4). Peak strength increased with
224 increasing confining pressure, as expected. At 50°C, an increase in the number of injections of
225 bacterial suspension resulted in increased peak strength and brittle behavior of the samples (Fig. 4).
226
227 All MICP-treated samples exhibited volumetric expansion at small strains and a dilative response
228 at large strain (Fig. 5). The amount of dilation decreased with increasing confining pressures (Fig.
229 5 a-d). Samples treated at 4°C and 50°C exhibited less contraction at small strains compared to those
230 treated at 20°C and 35°C. Among all temperatures, the sample treated at 35°C showed the largest
231 dilatancy when the strain was 15% at a given confining pressure (e.g., 100 kPa), followed by those
232 treated at 20°C, 4°C, and 50°C (Fig. 5a). The volumetric behavior of the samples was related to the
233 treatment temperature, which directly affects the CaCO₃ content. Higher carbonate content led to
234 higher dilatancy at large strains at a given confining pressure, as reported by other studies (e.g., Lin
235 et al., 2016; Xiao et al., 2019) conducted at room temperature. When the strain was small,
236 contraction was observed for all samples, but samples treated at 50°C showed substantial dilatancy
237 improvement as the number of injections of bacterial suspension was increased.

238

239 The stress paths and critical state lines in q-p' space are shown in Fig. 6 (a) and (b). The critical state
240 lines moved upwards with increasing CaCO₃ content produced in samples at varied temperatures or
241 with varying numbers of bacterial injections. This indicates that temperature and bacterial injection
242 number influence the CaCO₃ content produced, which consequently affects the strength behavior of

243 MICP-treated soil. The low strength behavior of MICP-treated soil at 50°C can be improved by
244 increasing the number of injections of bacterial suspension.

245

246 **3.3 Peak strength and residual strength**

247 The increase in confining pressure led to an increase in both peak strength (q_p) and residual strength
248 (q_R), as shown in Fig. 7 b and d. For instance, the peak strength of the sample treated at 20°C rose
249 from 623.6 kPa to 1576.8 kPa as the confining pressure was increased from 100 kPa to 400 kPa,
250 while the residual strength rose from 271.7 kPa to 812.2 kPa (Fig. 7). Increasing the number of BS
251 injections also resulted in a important improvement in both peak and residual strengths (Fig. 7 b and
252 d).

253

254 The effect of temperature and number of BS injections on peak and residual strengths can be
255 attributed to the variation in CaCO_3 content, as illustrated in Fig. 7 by the CaCO_3 contents. In the
256 residual state, the strength of the sample is mainly due to the non-broken calcium carbonate between
257 particles and the surface roughness of soil particles caused by CaCO_3 precipitation. The cementation
258 degraded substantially after reaching the peak state, especially in stronger specimens, but the non-
259 broken calcite still acts as cementation, and the roughness of sand particles may also contribute to
260 the residual strength.

261

262 Multiple BS injections resulted in an increase in peak and residual strengths because the calcite
263 crystal clusters grew in size. However, the sample with the highest strength was still the one with
264 one injection of BS at 35°C. This highlights the significance of adjusting the temperature

265 specifically for each MICP application. In cases where temperature cannot be controlled, altering
266 the MICP recipe, specifically the number of bacterial solution injections, is suggested.

267

268 **3.4 Effective strength parameters**

269 Fig. 8a shows that the effective cohesion and friction angle of the sand sample initially increased
270 and then decreased as the temperature increased from 4°C to 50°C. The effective strength parameters
271 were smaller at lower and higher temperatures compared to those in the temperature range of 4°C-
272 50°C. Within this range, the effective cohesion and friction angle varied from 3.1 kPa to 80.9 kPa
273 and from 26.8° to 36.6°, respectively. The effective cohesion and friction angle increased linearly
274 with an increase in the number of bacterial solution injections. Furthermore, the effective cohesion
275 increased exponentially with an increase in the average CaCO₃ content, while the effective friction
276 angle increased linearly with the average CaCO₃ content, as depicted in Figure 9. These results are
277 consistent with previous studies conducted at a standard temperature of 25°C (e.g., Cui et al. 2017;
278 Chou et al. 2020).

279

280 Temperature has a noteworthy effect on the strength of sand treated with MICP, with CaCO₃ content
281 and product uniformity being the primary factors (Feng and Montoya 2016; Cui et al. 2017; Nafisi
282 et al. 2020; Konstantinou et al. 2021). Previous triaxial testing studies have shown that an increase
283 in CaCO₃ content leads to an increase in both effective cohesion and friction angle (Cui et al. 2017;
284 Chou et al. 2020). For non-cemented soil, grain size distribution, angularity, and particle
285 interlocking primarily affect the effective cohesion and friction angle. However, for cemented soil,
286 CaCO₃ precipitation has a bonding effect on sand cementation, leading to a substantial increase in

287 cohesion, which defines the non-frictional part of shear resistance (DeJong et al. 2010). Additionally,
288 the 'added' calcium carbonate crystals increase particle interlocking, leading to an increase in the
289 friction angle. The study shows that the increase in cohesion is more pronounced than the increase
290 in friction angle in MICP-treated specimens due to the added cementation at the contact points.

291

292 **3.5 Microstructure characterization**

293 As the temperature increased from 4°C to 35°C, more CaCO₃ crystals precipitated on the surface of
294 sand particles or at the particle contacts. However, for the sample treated at 50°C, the number of
295 CaCO₃ crystals decreased dramatically. This is consistent with the macro-scale measurement of
296 CaCO₃ content shown in Figure 3. The size of the CaCO₃ crystals also varied with temperature.
297 Small crystals mainly precipitated in samples treated at 4°C, while larger crystal clusters were
298 formed in samples treated at 20°C and 35°C. On the other hand, crystal clusters were observed at
299 50°C, but their size was smaller than those obtained at 20°C and 35°C. The size of the crystal clusters
300 increased with the increase in the number of BS injections, with the average diameter of crystal
301 clusters reaching 25 μm and 40 μm for samples treated with BS injected 3 times and 6 times,
302 respectively (Figure 10).

303

304 The XRD diagrams confirmed the presence of calcite at the peaks with angles of 29.4° and 55°
305 (Figure 11). To further analyze the data, the areas under the two peaks were measured for each
306 condition and presented in Table 2 (units in cps*degrees). The findings showed that the optimum
307 point was at 35 degrees Celsius, where both areas under the two peaks were larger. The carbonate
308 measurements with the acid method also supported these results as shown previously.

309

310 **3.6 Correlation between CaCO₃ microstructure and soil strength behavior**

311 Figure 12 illustrates magnified SEM images (Figure 12a) and a schematic of cementing pattern
312 (Figure 12b). The distinguish of crystal cluster is also illustrated in Figure 12a. At 20°C, 35°C, and
313 50°C, smaller and more crystals form into clusters in the cementing pattern (compared to the case
314 with multiple BS injections), whereas at 4°C, the cluster pattern is not evident. Increasing the
315 number of BS injections at 50°C results in larger crystals. Figure 13 presents the quantification of
316 number and size of crystal or clusters, with crystal or clusters numbered according to size. Among
317 the four temperatures (4°C, 20°C, 35°C, and 50°C), 20°C (see Figure 12 and Figure 13b) produces
318 the largest crystal number range, while 35°C produces the largest crystal size range (see Figure 12
319 and Figure 13c). This indicates that 4°C and 50°C produce fewer and smaller crystals, while 20°C
320 and 35°C produce more and larger crystal clusters. Moreover, 20°C produces a larger number of
321 smaller clusters than 35°C (see Figure 12 and Figure 13 b-c). In the case of multiple bacterial
322 injections, larger crystals tend to form instead of forming large clusters (See Figure 12). The size of
323 crystal increases as the number of BS injections increases from 1 to 6 (Figure 13d-f). This might be
324 because the injected bacteria preferentially attach to calcite, which acts as a better nucleation site
325 for crystals than quartz.

326

327 Based on Figures 12 and 13, it can be concluded that when bacteria are only injected once, but the
328 temperature changes, clusters with different sizes and numbers tend to form with temperature
329 changes. However, when the temperature is fixed at 50°C, but the injection number of bacterial
330 suspension increases, large crystals tend to form. To compare the effect of clusters and larger crystals

331 on increasing the strength behavior of MICP treatment, the correlation between average CaCO_3 and
332 effective cohesion, effective friction angle, peak deviatoric stress, and residual deviatoric stress were
333 analyzed separately for tests 1-4 and 4-6 (results are shown in Figures 14 and 15).

334

335 The results indicate that when the CaCO_3 content is the same, compared to large CaCO_3 , crystal
336 clusters are more effective in increasing effective cohesion (Figure 14a), peak strength (Figure 15),
337 and residual strength (Figure 15). This may be because increasing cluster size not only improves
338 bonding efficiency, leading to an increase in cohesion but also enhances surface roughness, resulting
339 in a more effective increase in friction. However, large crystals are more effective in increasing the
340 effective friction angle at the low average CaCO_3 content range compared to crystal clusters and
341 less effective at the high average CaCO_3 content range. This may be because, at the low average
342 CaCO_3 content range, the big crystals are more effective in forming friction between soil particles,
343 compared to smaller but a larger number of crystals. However, at the high average CaCO_3 content
344 range, the big crystals are still effective in forming friction between soil particles, but because the
345 crystals are big, the number of crystals is fewer compared to crystal clusters, resulting in reduced
346 overall friction effectiveness. Further work can be done to study the effect of crystal and cluster
347 properties on MICP-treated soil, both experimentally and numerically.

348

349 **4 Conclusions**

350 This study aimed to investigate the effectiveness of microbially induced calcium carbonate
351 precipitation (MICP) in treating granular media at different temperatures. A series of consolidated
352 drained triaxial compression tests were conducted to evaluate the engineering properties of treated

353 soils, and micro-scale investigation was conducted to explore the characteristics of the precipitated
354 minerals responsible for these properties. Based on the findings, the following conclusions were
355 drawn:

356

357 Increasing the treatment temperature from 4°C to 35°C increased the CaCO₃ content of samples,
358 but at 50°C, the content decreased considerably. All MICP-treated samples exhibited strain softening
359 and volumetric dilatancy, with peak and residual strength values varying with treatment temperature.
360 Samples treated at 35°C exhibited the highest peak and residual strength, followed by 20°C, while
361 samples treated at lower and higher temperatures resulted in lower strength. Increasing the number
362 of bacterial solution injections improved the peak and residual strength of samples treated at 50°C.

363

364 The effective cohesion and friction angle followed a similar pattern as peak and residual strength,
365 increasing and then decreasing with temperature. The amount of CaCO₃ precipitation was attributed
366 to the difference in strength enhancement of MICP-treated sand at different temperatures.

367

368 SEM images and XRD results showed that rhombohedral calcite was the dominant type of CaCO₃
369 crystals produced at different temperatures, with most crystals forming as clusters on particle
370 surfaces or at particle contacts. Crystal cluster sizes increased as the temperature increased from 4°C
371 to 35°C and decreased at 50°C.

372

373 The mechanical behavior of MICP-treated sand was found to be closely related to treatment
374 temperature, and altering the MICP recipe, specifically the number of bacterial solution injections,

375 was suggested as a potential solution when temperature cannot be controlled. The efficiency of
376 MICP in soil strength enhancement was found to depend on the properties of produced CaCO₃, and
377 any environmental factors affecting these properties should be considered when predicting MICP
378 efficiencies.

379

380 The study showed that crystal clusters are better than large crystals in increasing effective cohesion,
381 peak strength, and residual strength when CaCO₃ content is the same, due to improved bonding
382 efficiency and surface roughness. However, large crystals are more effective in increasing the
383 effective friction angle at low average CaCO₃ content range. Further research is needed to explore
384 the impact of crystal and cluster properties on MICP-treated soil.

385

386 **Data Availability Statement**

387 Some or all data, models, or code that support the findings of this study are available from the
388 corresponding author upon reasonable request.

389

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568

Table 1 MICP treatment protocols

Test No.	Sample numbers	Dry density, ρ_d (g/cm ³)	Relative density, D_r (%)	Treating temperature (°C)	Injection number of bacterial suspension	Mean effective stress, p' (kPa)
T4	6	1.48	50	4	1	100, 200, 300, 400
T20	6	1.49	50	20	1	100, 200, 300, 400
T35	6	1.48	50	35	1	100, 200, 300, 400
T50 (BS1)	6	1.48	50	50	1	100, 200, 300, 400
T50 (BS3)	6	1.49	50	50	3	100, 200, 300, 400
T50 (BS6)	6	1.48	50	50	6	100, 200, 300, 400

Table 2 The areas under the two calcite peaks in the XRD diagrams for the various treatment

temperature conditions (units in cps*degrees)

	29.4°	55°
T4	150.825	180.325
T20	650.975	181.3
T35	806.8	184.225
T50-BS1	51.65	157.225
T50-BS3	107.725	160.325
T50-BS6	115.725	171.3

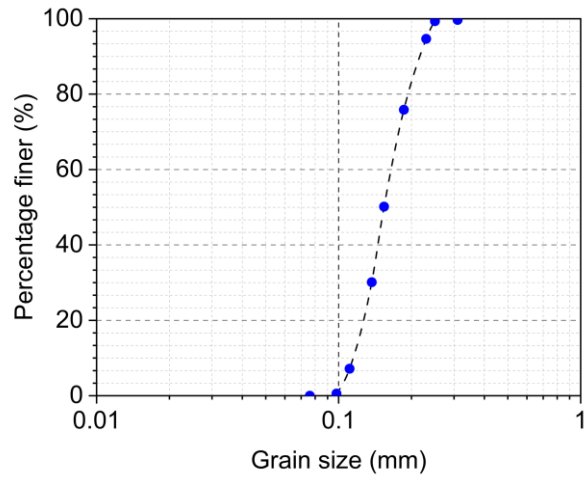


Figure 1 Particle size distribution of the sand used in this study

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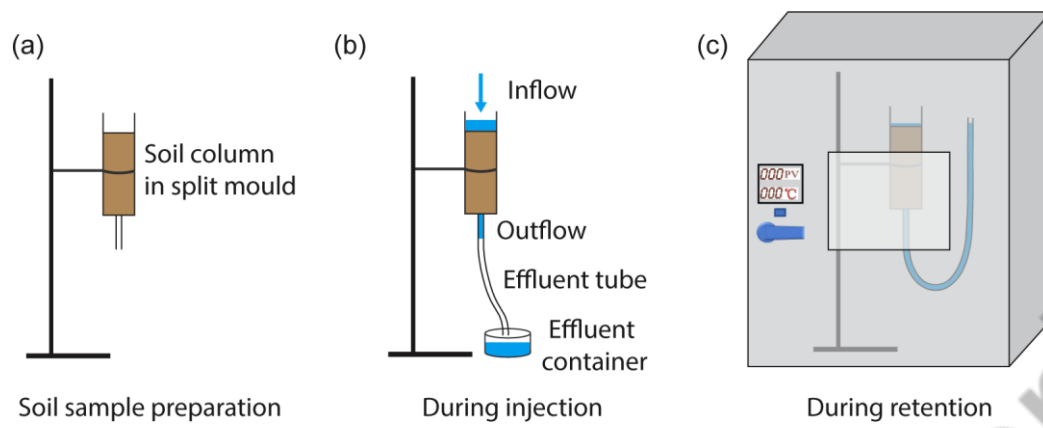


Figure 2 Schematic of the soil column experiments: (a) sand column preparation, (b) and (c) MICP treatment

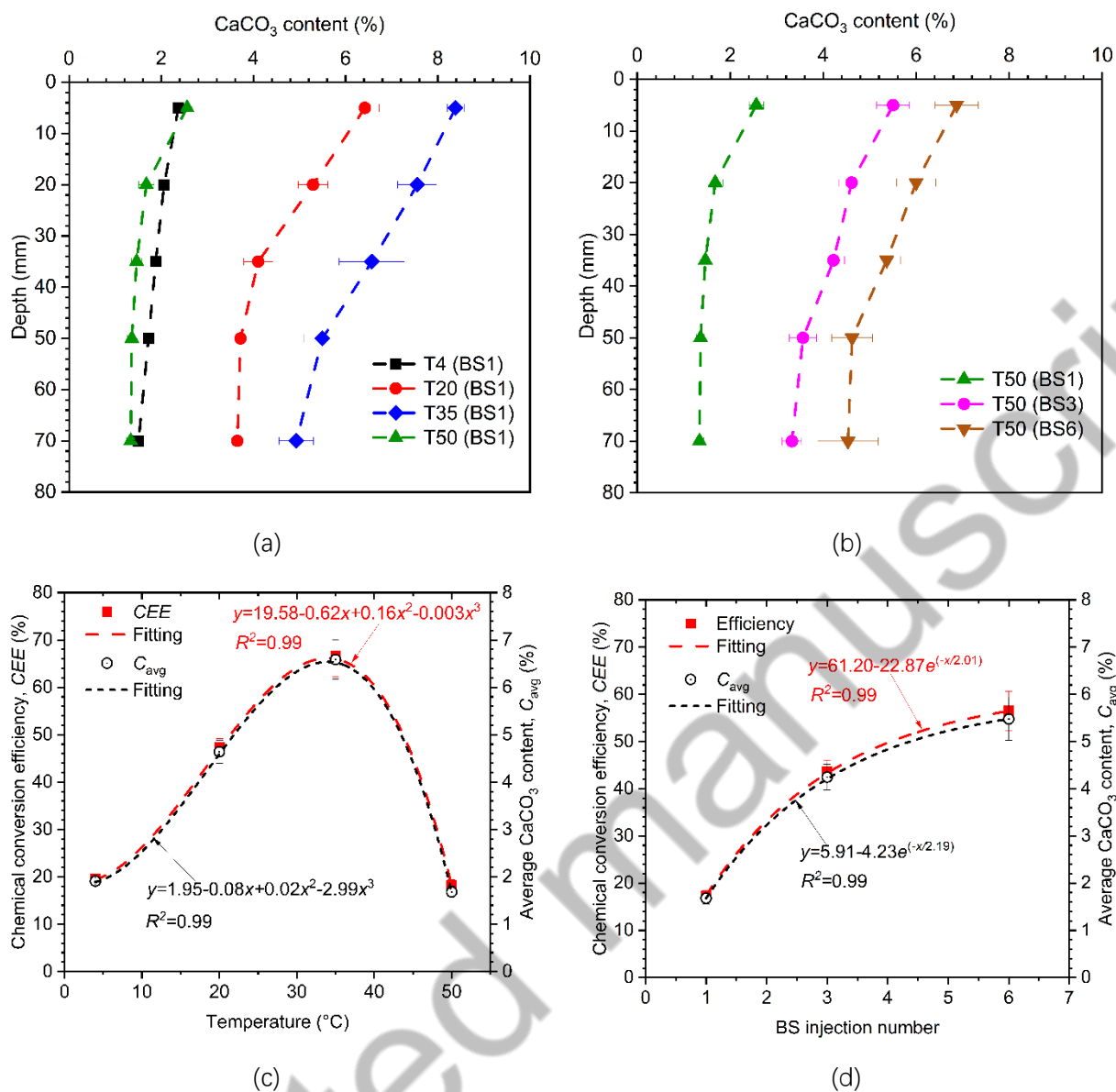


Figure 3 CaCO₃ distribution along sand columns and chemical transform efficiency of MICP: (a, c) specimens treated at different temperatures with bacteria introduced only once; (b, d) specimens treated at 50°C with bacteria introduced once, twice, and three times, respectively.

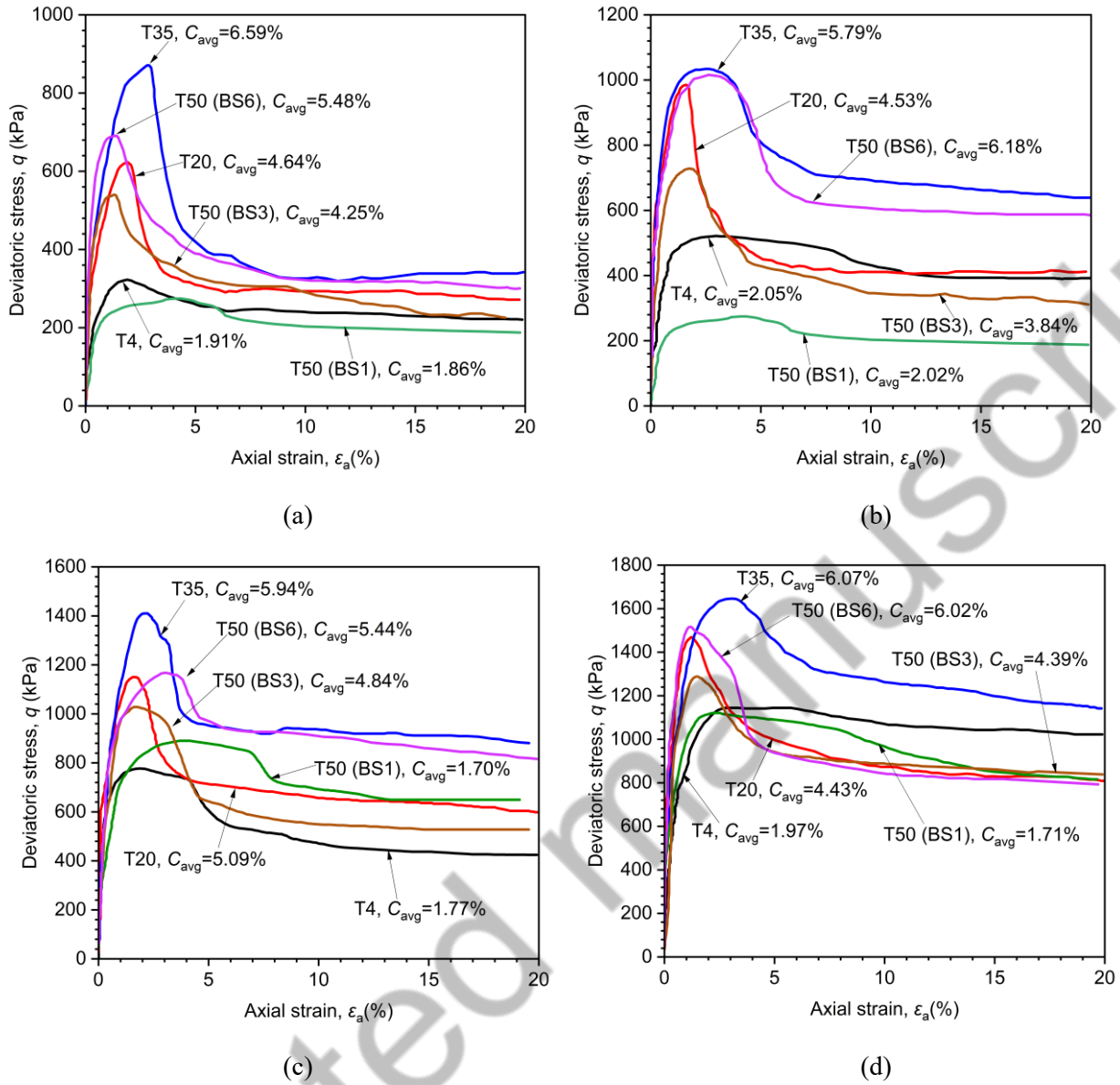
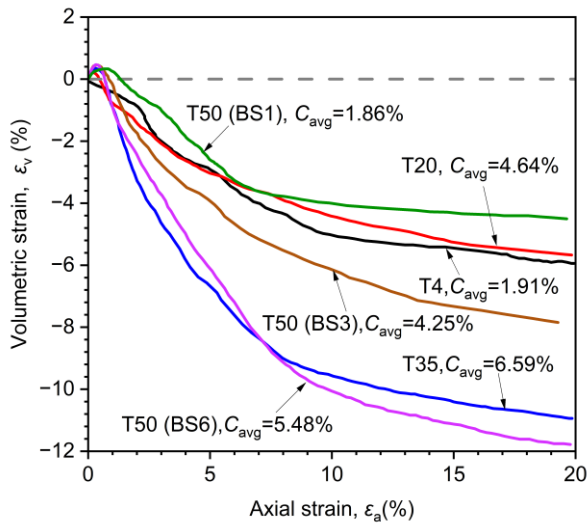
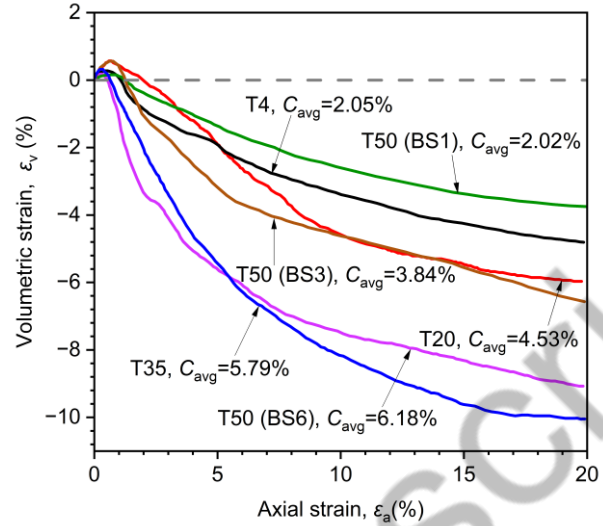


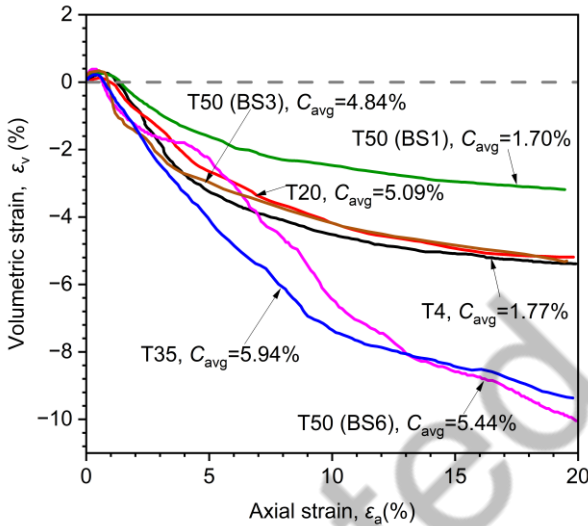
Figure 4 Stress-strain behaviour of MICP-treated sand at different confining pressures: (a) $p'_c = 100$ kPa; (b) $p'_c = 200$ kPa; (c) $p'_c = 300$ kPa; and (d) $p'_c = 400$ kPa; T , indicates the treatment temperature, BS , indicates the injection number of bacterial suspension; C_{avg} indicates the average CaCO_3 content



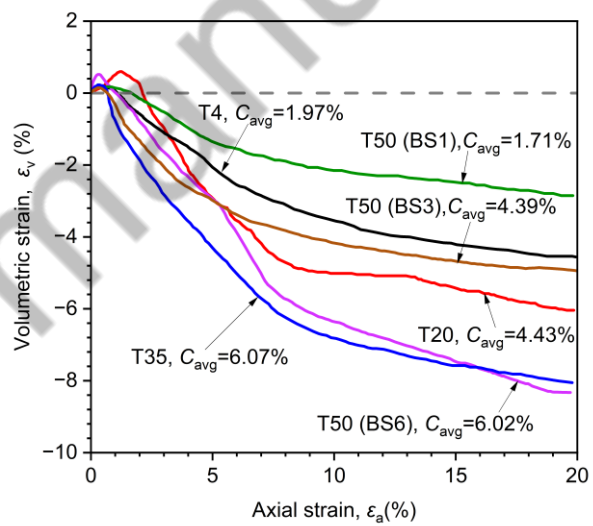
(a)



(b)

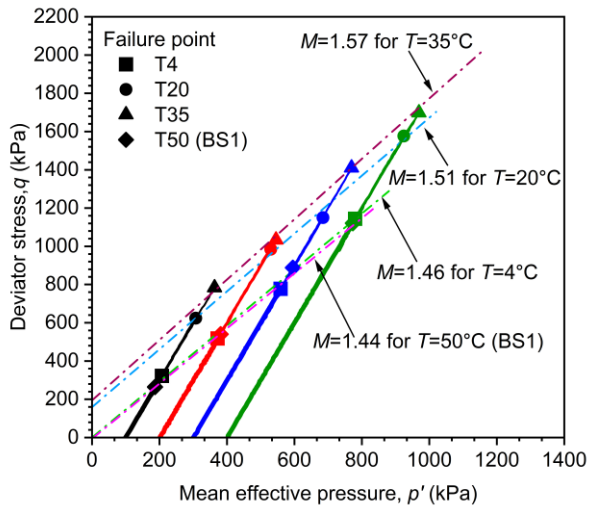


(c)

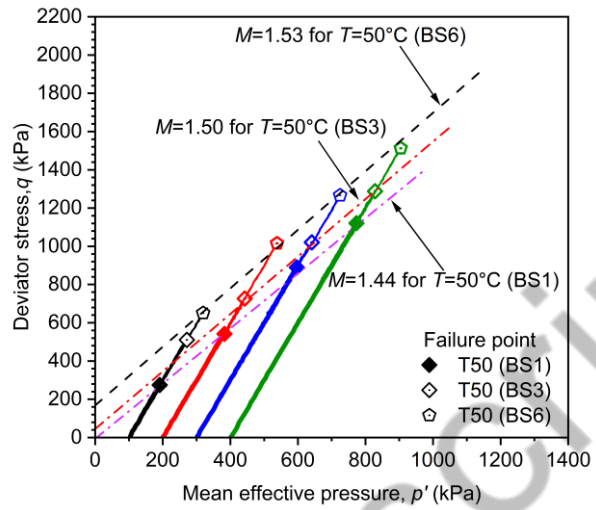


(d)

Figure 5 Volumetric behaviour of MICP-treated sand at different confining pressures: (a) $p_c' = 100$ kPa; (b) $p_c' = 200$ kPa; (c) $p_c' = 300$ kPa; and (d) $p_c' = 400$ kPa; T , indicates the treatment temperature; BS, indicates the injection number of bacterial suspension; C_{avg} indicates the average CaCO_3 content.

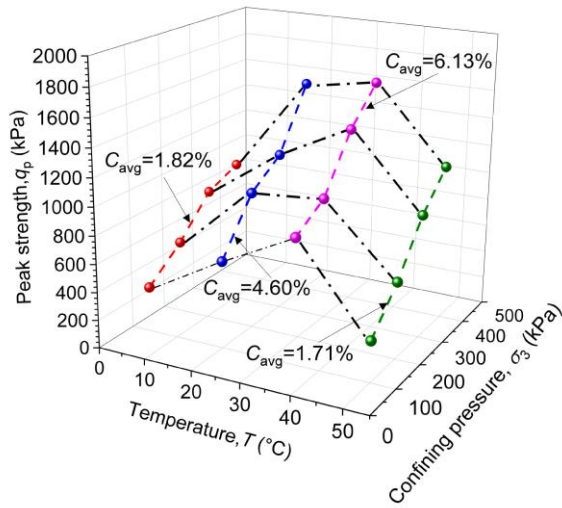


(a)

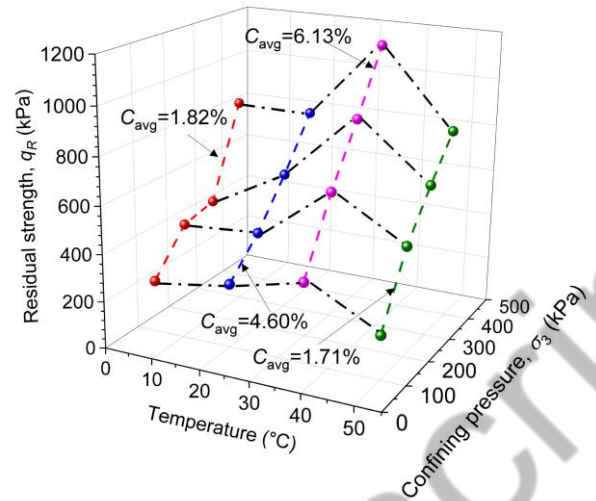


(b)

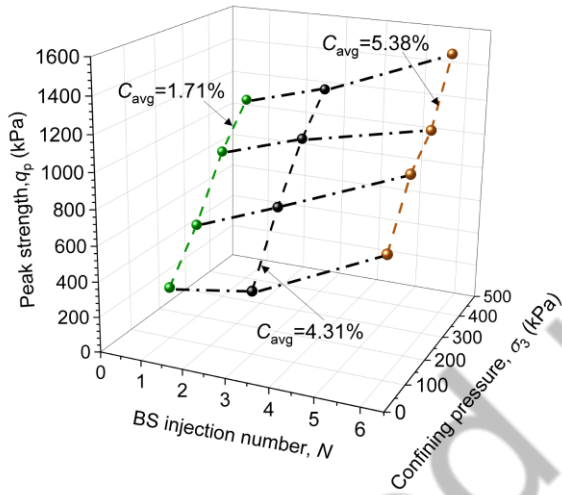
Figure 6 Stress path and critical state line in q - p' space: (a) specimens treated at temperatures range from 4°C to 50°C with bacteria introduced only once; (b) specimens treated at 50°C with bacteria introduced once, twice, and three times, respectively.



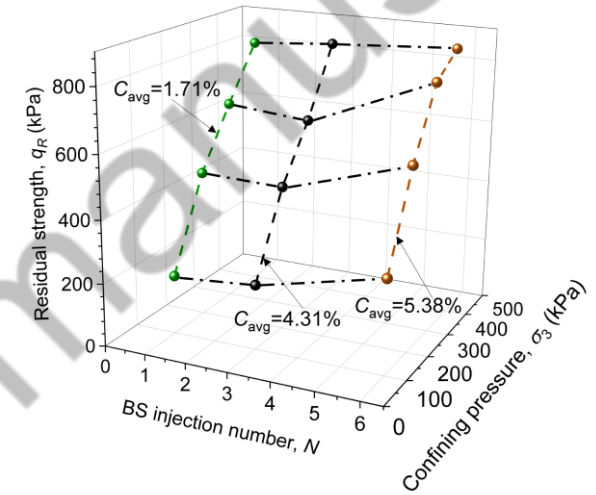
(a)



(b)

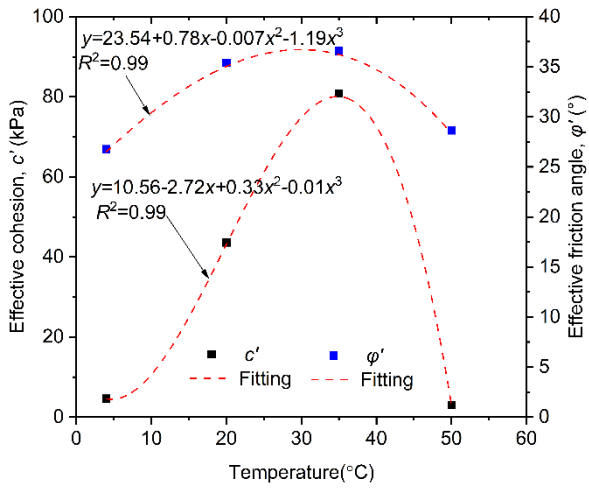


(c)

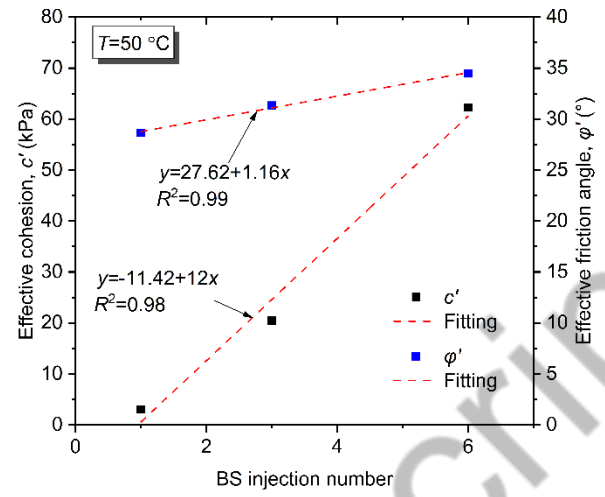


(d)

Figure 7 Measured and fitted relationship of q_p - T - σ_3 and q_R - T - σ_3 : (a, c) specimens treated at different temperatures with bacteria introduced only once; (b, d) specimens treated at 50°C with bacteria introduced once, twice, and three times, respectively.

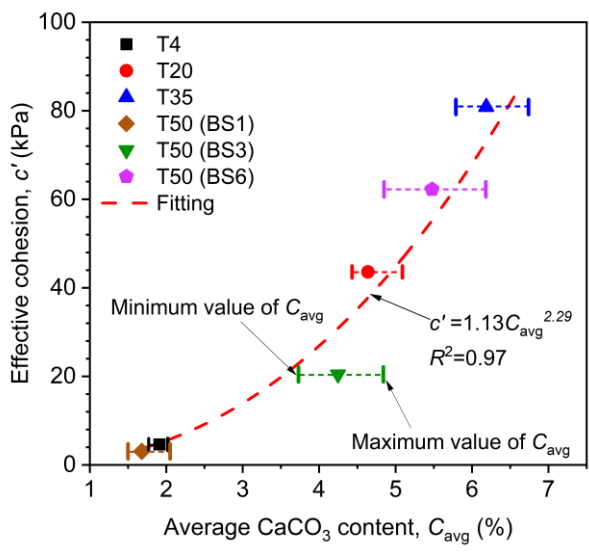


(a)

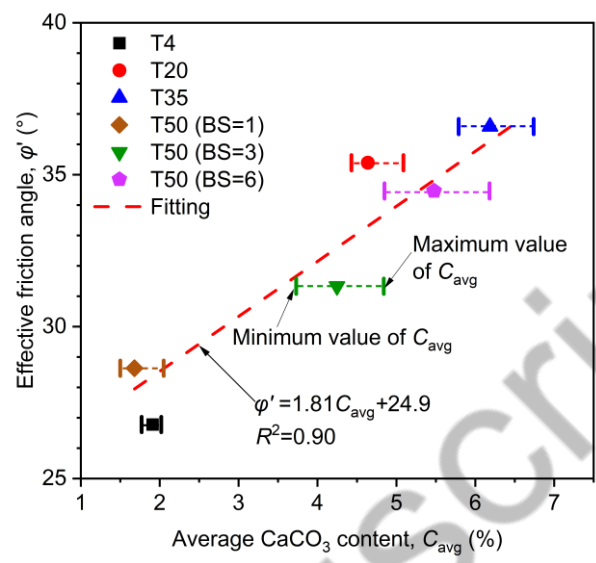


(b)

Figure 8 Relationship between strength parameters and average CaCO_3 content at (a) different temperature; (b) different BS injection numbers at 50°C (1)



(a)



(b)

Figure 9 Effects of average CaCO_3 content on (a) effective cohesion; and (b) friction angle.

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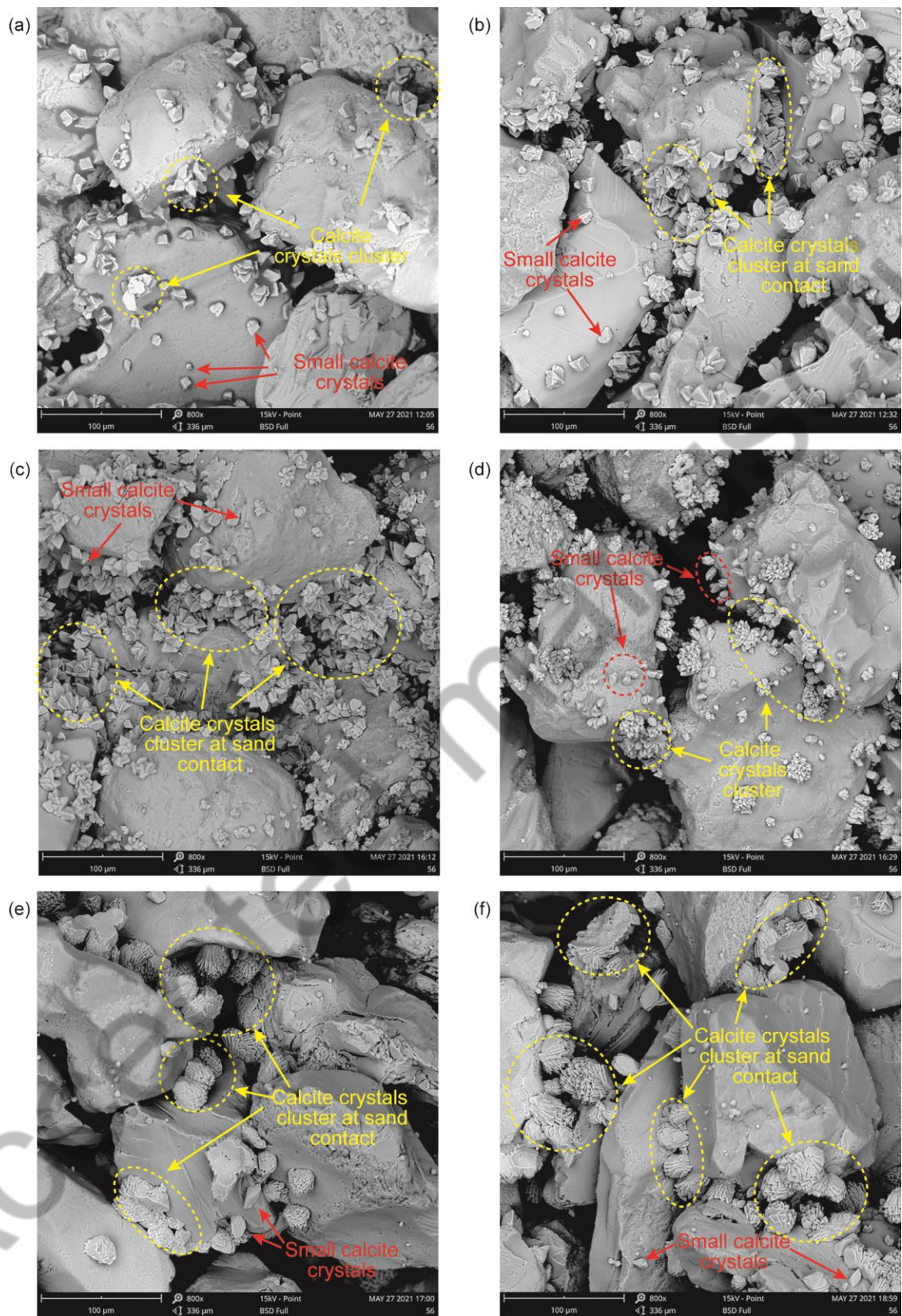
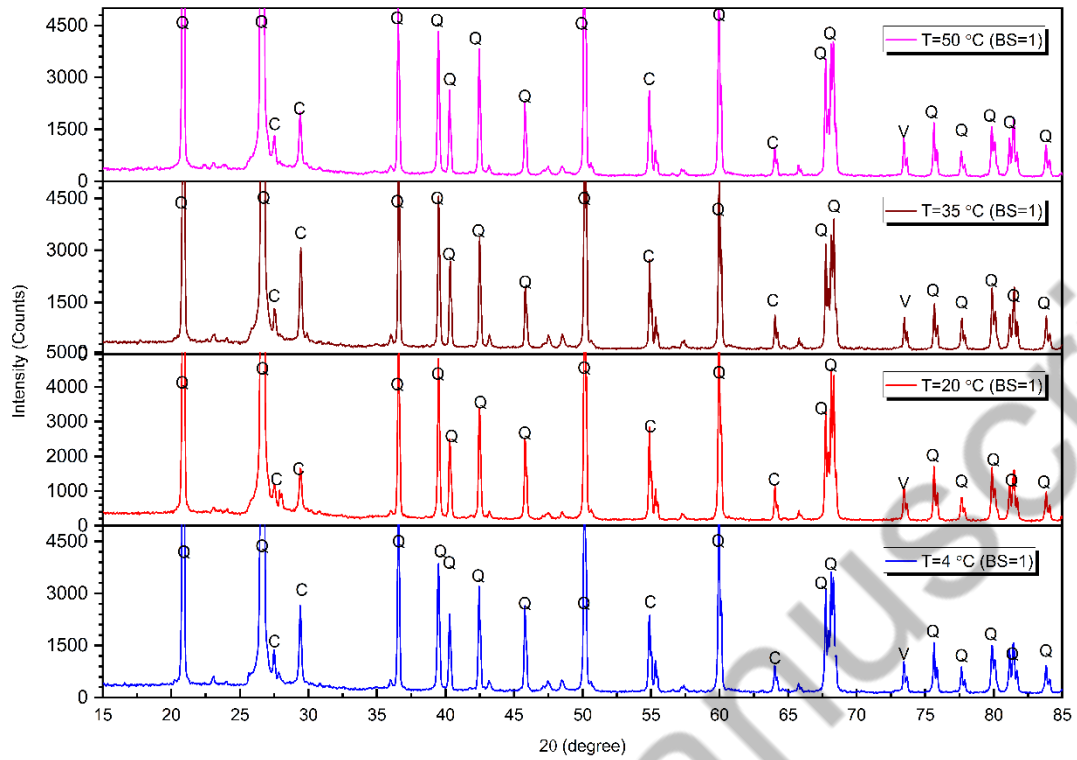
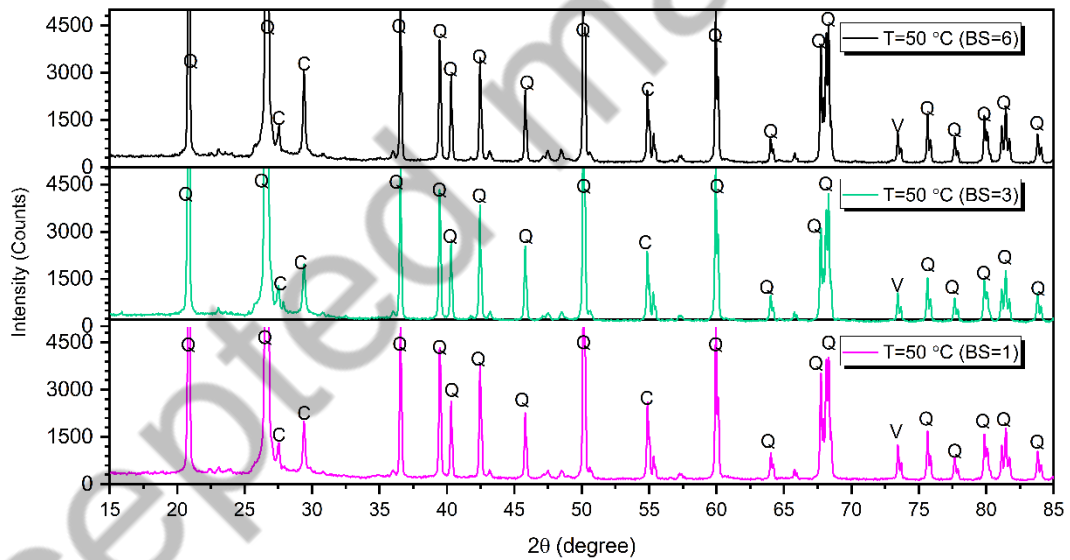


Figure 10 SEM images of specimens treated at different temperatures: (a) T=4°C (BS1); (b) T=20°C (BS1); (c) T=35°C (BS1); (d) T=50°C (BS1); (e) T=50°C (BS3); (f) T=50°C (BS6)

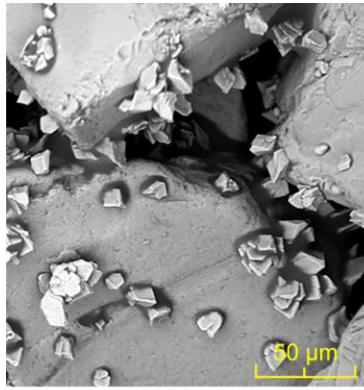


(a)

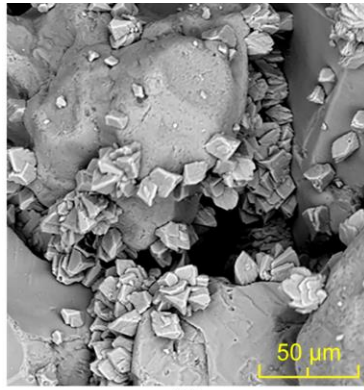


(b)

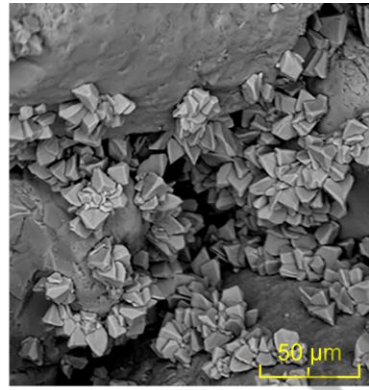
Figure 12 The XRD results of biocemented sand at: (a) different temperatures; and (b) different BS injection number at 50°C; Q indicates Quartz, Ca indicates calcite.



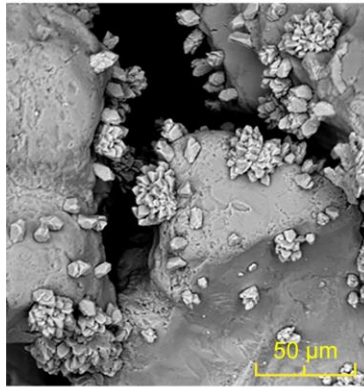
(a)



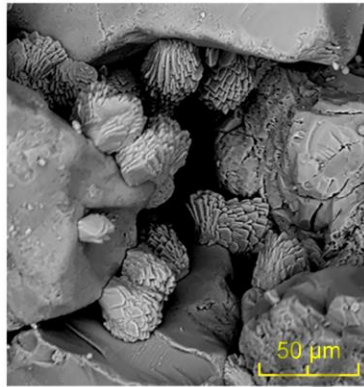
(b)



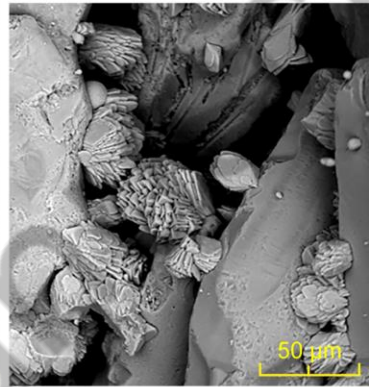
(c)



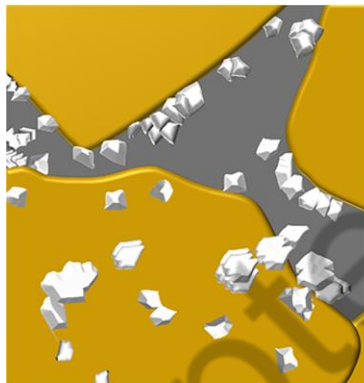
(d)



(e)



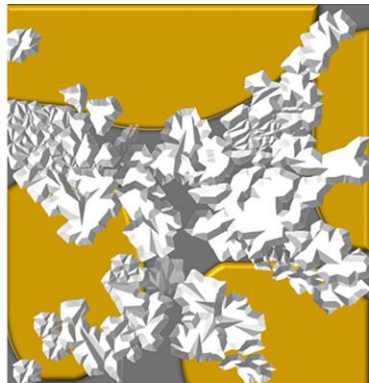
(f)



(a)



(b)



(c)



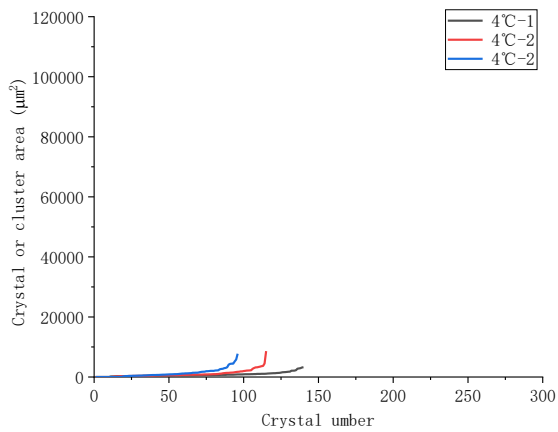
(d)



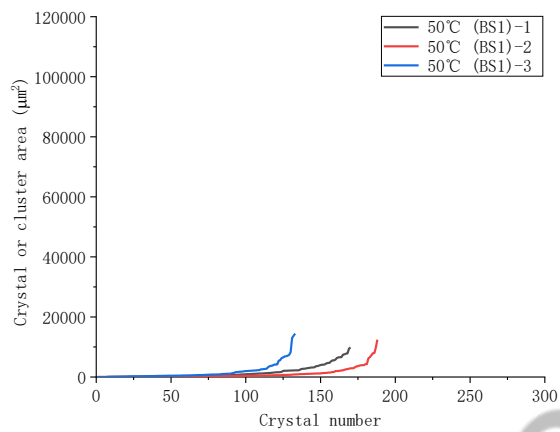
(e)



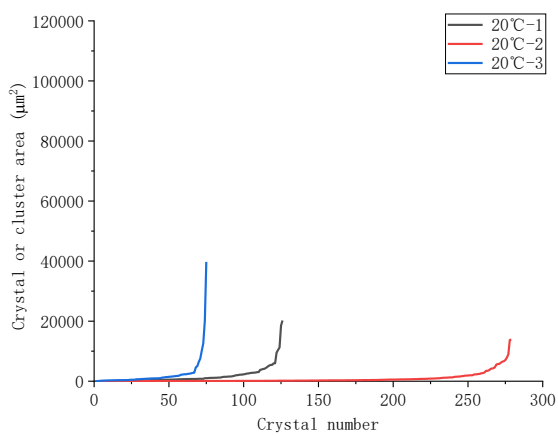
(f)



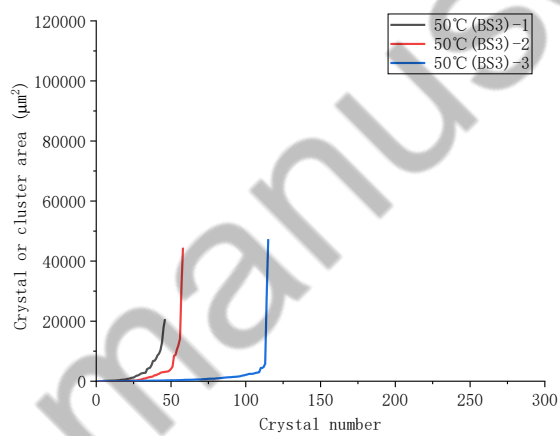
(a)



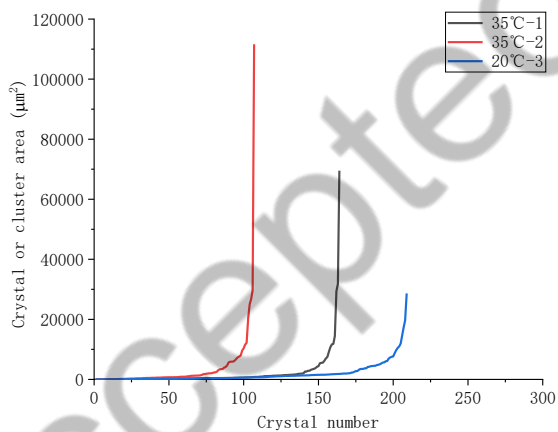
(d)



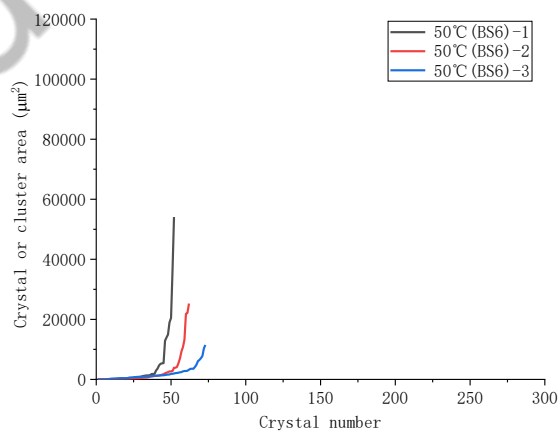
(b)



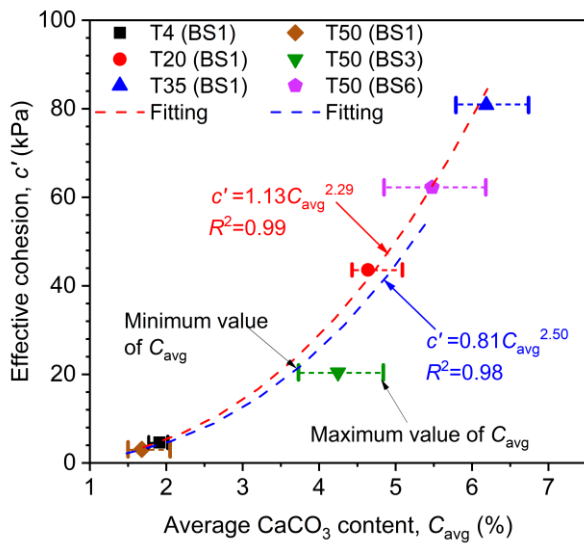
(e)



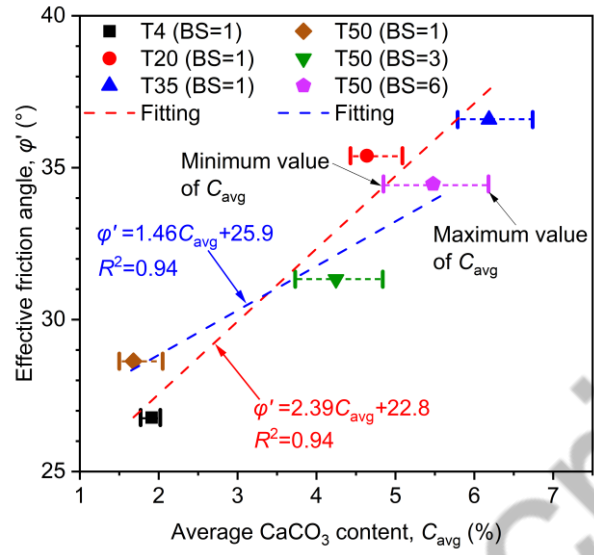
(c)



(f)

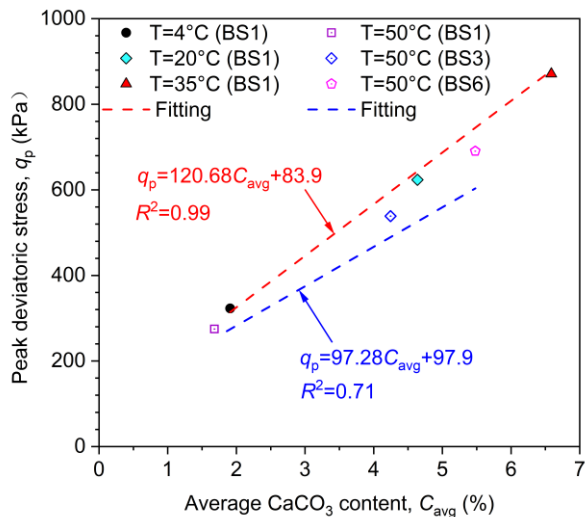


(a)

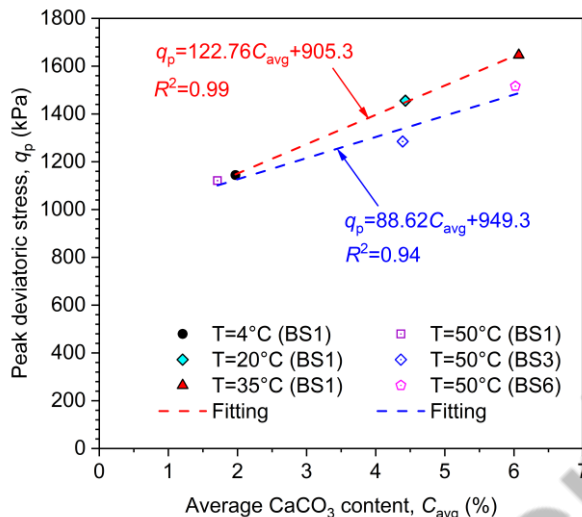


(b)

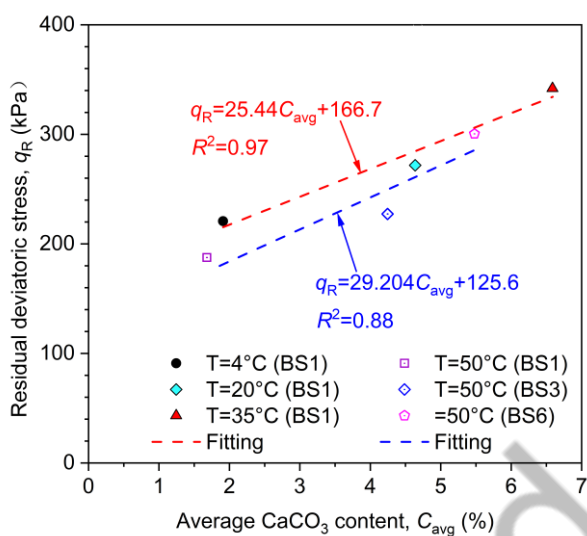
Figure 14 Effects of average CaCO_3 content on (a) effective cohesion; and (b) friction angle.



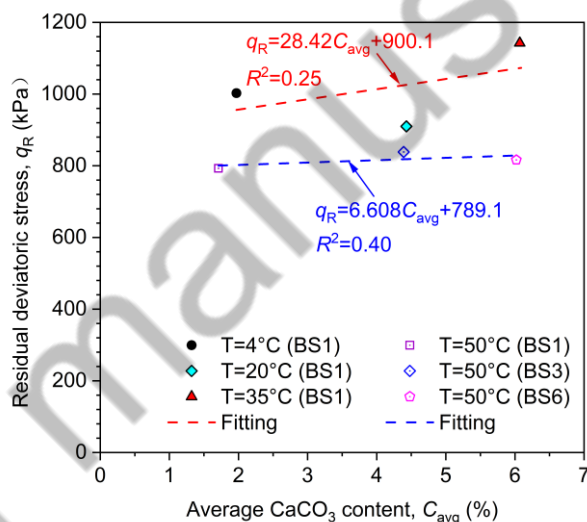
(a)



(b)



(c)



(d)

Figure 15 Effects of average CaCO_3 content on peak strength (a, c) and residual strength (b, d) at 100 kPa (a, c); and 400 kPa (b, d)