

Are Melamine and Its Derivatives the Alternatives for Per- and Polyfluoroalkyl Substance (PFAS) Fabric Treatments in Infant Clothes?

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Cite This: *Environ. Sci. Technol.* 2020, 54, 10207–10216



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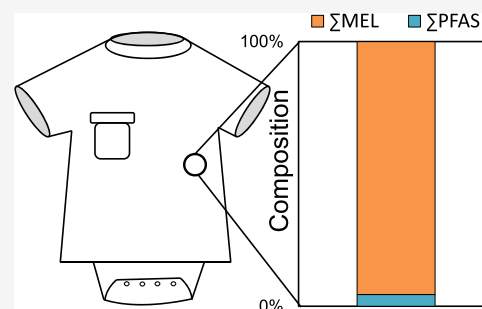


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ABSTRACT: Per- and polyfluoroalkyl substances (PFAS) and melamine (MEL)-based compounds are used in textile finishing as grease, stain, and water repellents. Here, we investigated the occurrence of a large suite of PFAS and MEL compounds in 86 infant clothing items. The Σ MEL concentrations ranged from below the method detection limit to 250,000 ng/g with a median concentration of 78.2 ng/g, significantly higher ($p < 0.05$) than the Σ PFAS levels (1.22–203 ng/g; median 3.62 ng/g). MEL and its derivatives were most abundant in nylon clothes (median 32,800 ng/g), followed by organic cotton (median 6120 ng/g). In a simulated laundering experiment, the Σ MEL concentrations in clothing decreased on an average by ~60 and 90% when washed in cool (20 °C) and warm (50 °C) water, respectively. This removal rate increased to 97% when the samples were washed with a detergent. The estimated daily intakes of MEL and PFAS through dermal absorption from nylon clothes were three orders of magnitude higher than those from the non-nylon clothes and decreased by more than half for washed clothes. Our findings demonstrate that MEL-based compounds are abundant in infant clothing and suggest that this group of compounds could be used as potential PFAS replacements in textile finishing.



INTRODUCTION

Fabric finishing is an important process in textile manufacturing and includes a variety of treatment applications used to impart specific functional properties to the material. More than 2500 chemicals are currently used in fabric finishing treatments,¹ including dyes, antioxidants, flame retardants, fiber purifiers, UV light absorbers, and grease, stain, and water repellents.^{2,3} The latter are essential for oil-, stain-, and waterproofing fibers and fabrics and have been in use since the 1950s when textiles with enhanced water resistance and grease-proof properties were first introduced to the market.⁴ The majority of these applications are fluoropolymer-based and contain per- and polyfluoroalkyl substances (PFAS) that are applied as surface coatings to fabrics or fibers, followed by polymerization and curing.⁵ A number of PFAS-based textile coating agents are marketed under trade names such as Teflon, Stainmaster, NanoTex, GoreTex, and Unidyne. The two major PFAS compounds, perfluorooctanesulfonic acid (PFOS) and perfluorooctanoic acid (PFOA), have been found in various textiles, including carpets, rain jackets, gloves, and backpacks.^{6–11} In addition, because of global restrictions on the production and use of persistent and toxic PFOS and PFOA, textile treatments using PFAS with shorter (<C8) carbon chains and nonionic (neutral) molecules, generally considered less environmentally persistent and bioaccumulative, have been increasingly used.^{12,13} For example, surface coating finishes such as Scotchgard PM-3622, PM-490, and PM-930 use side-

chain fluorinated polymers based on the derivatives of perfluorobutane sulfonyl fluoride,¹⁴ which can degrade to perfluoro-1-butanesulfonic acid (PFBS, C4).¹⁵ Neutral PFAS, such as fluorotelomer alcohols (FTOHs), especially those with six perfluorinated carbons, are used as cross-linking agents for textiles.¹⁶ Both short-chain and neutral PFAS were found in textile impregnating treatments with levels reaching 13.2 mg/L.¹⁷ As a result, dermal exposure from clothes and other textiles has been recognized as an important human exposure pathway to many PFAS.^{3,18}

Considering the growing concerns regarding the toxicity of the entire PFAS family,¹⁹ the textile industry has been working on implementing “green” strategies for textile finishing.⁵ The results of a 2015 survey of 17 textile manufacturing companies indicate that 4 of these companies eliminated fluorine-based technologies in waterproof textiles and switched to a PFAS-free polymer technology to achieve water-repellent properties.⁵ The Roadmap to Zero Discharge of Hazardous Chemicals (ZDHC) was launched in 2011 by a group of major apparel and footwear brands and retailers, aimed to lead the industry

Received: May 12, 2020

Revised: July 8, 2020

Accepted: July 14, 2020

Published: July 14, 2020



toward zero discharge of hazardous chemicals by 2020.²⁰ According to the ZDHC report on durable water repellents, melamine (MEL) and its derivatives are the major group of nonfluorinated chemicals being increasingly used in water-proofed fabrics instead of PFAS.²⁰

MEL has received global attention after a series of pet food and infant formula adulteration incidences that resulted in renal damage, and in some cases, renal failure and death of infants and pets.^{21–23} However, it is less known that, in addition to food adulteration, MEL and its derivatives are also widely used as textile additives. MEL-based polymers were first introduced in the 1930s and have been used since then as fabric surface coating resins.²⁴ MEL-based polymers comprise a large group that includes MEL–formaldehyde, methylated MEL–formaldehyde, MEL–urea–formaldehyde, MEL–urea–phenol–formaldehyde, and other resins. Similar to PFAS, these MEL-based resins are used to enhance water-repellent and wrinkle-resistant properties of textiles^{11,25} but are generally considered less toxic. In addition, MEL-based fibers can be used in flame-resistant protective clothing because of their low thermal conductivity, high flame resistance, and self-extinguishing properties.²⁶ Other MEL-based compounds, such as cyanuric acid (CYA), ammeline (AMN), and ammelide (AMD), are byproducts or impurities in the manufacturing of MEL²⁵ and can be used together with MEL to produce MEL-based resins. In addition, CYA is used as a disinfectant, bleaching agent, and fertilizer on its own.²⁷

Clothing has been recognized as an important source of chemical exposures through dermal absorption from direct contact of clothes with skin.²⁸ Infants wearing clothing items that are in direct contact with skin, such as cloth diapers and onesies, may be at a higher risk from nearly constant exposure.²⁹ Even low-dose environmental exposures are of concern in the early-life period and can result in health effects resonating across the entire life span.³⁰ However, the number of studies on chemical exposures from infant clothes is limited.^{31,32}

Here, we simultaneously measured two major groups of chemicals used as textile surface treatments, PFAS and MEL compounds, in a wide selection of infant clothing. We investigated the occurrence and distribution of 11 MEL derivatives and 40 ionic and neutral PFAS in infant clothing items purchased from local and online stores in the United States and assessed infants' dermal exposure to these chemicals from clothes. In addition, we conducted simulated laundering experiments to assess the release of MEL and PFAS compounds from clothing to the environment.

MATERIALS AND METHODS

Samples. Eighty-six pieces of new finished infant clothing (sizes 0–12 months) from nine national brands were purchased from local stores in Bloomington and Indianapolis, Indiana, United States, and online in June 2019. The clothes were manufactured during 2017–2019 and included various categories such as shirts ($n = 13$), pants ($n = 12$), socks ($n = 12$), cloth diapers ($n = 20$), dresses ($n = 2$), onesies ($n = 24$), and outerwear ($n = 3$) made of conventional and organic cotton, polyester, spandex, and nylon. The clothes were manufactured in nine countries, including Cambodia ($n = 6$), China ($n = 38$), India ($n = 6$), Indonesia ($n = 5$), Nicaragua ($n = 5$), El Salvador ($n = 4$), Thailand ($n = 2$), the United States ($n = 10$), and Vietnam ($n = 9$). Table S3 includes the details of

samples analyzed in this study, including fabric composition, clothing type, manufacturing origin, and year of production.

Sample Treatment. Three pieces of fabric (3×3 cm) were randomly cut out of each garment, finely cut, and mixed, and 100 mg of this sample was transferred into a 15 mL polypropylene tube and spiked with 50 ng of surrogate standards (Tables S1 and S2). The samples were then sonicated three times (1 h each) with 4 mL of methanol at room temperature. The supernatants were combined, solvent-exchanged to 500 μ L acetonitrile with 5 mM ammonium formate buffer (95:5, v/v, pH = 4.0), filtered through a 0.2 μ m nylon syringe filter, and spiked with 50 ng of internal standards (Tables S1 and S2).

Simulated Laundering Experiments. Laundering experiments consisted of a simulated washing process that included either washing without detergent in water at 20 and 50 °C or washing with detergent in water at 20 and 50 °C. Clothing items ($n = 4$, all made of nylon) with the highest \sum PFAS and \sum MEL concentrations detected were included in these experiments. Fabric pieces (approximately 5×5 cm) cut from these selected clothing items were added to 400 mL of preheated high-performance liquid chromatography water at 20 and 50 °C, each in a separate beaker, and shaken in an incubator shaker at 150 rpm for 1 h. The clothing pieces were then squeezed to remove excess water and air-dried at room temperature covered with an aluminum foil in a fume hood. The experiment using a detergent was conducted in a similar way, but one drop of concentrated liquid laundry detergent was added to the water. The concentrations of MEL derivatives and PFAS were determined in washed fabric samples using the methods described above.

Instrumental Analysis. The complete list of the target compounds and surrogate and internal standards is given in Tables S1 and S2. The instrumental method for PFAS was previously reported⁷ and is briefly described in the Supporting Information (Tables S1 and S2, and Figure S1). MEL and its derivatives were analyzed using an ultraperformance liquid chromatograph coupled with a triple-quadrupole mass spectrometer (Agilent 1290 Infinity II UPLC-6470 QQQ-MS). Chromatographic separation of MEL, CYA, AMN, and AMD was achieved on an Acquity UPLC HILIC column (50, 2.1 mm i.d., 1.7 μ m thickness, Waters, Milford, MA). Ammonium formate buffer (mobile phase A) (5 mM, adjusted to pH 4.0 using formic acid) and acetonitrile (mobile phase B) were used as the mobile phases. The column temperature and the flow rate were 40 °C and 0.4 mL/min, respectively. The mobile-phase gradient was as follows: 0 min, 95% B; 5.6 min, 65% B; 9 min, 65% B; 9.1 min, 95% B; and 15.1 min, 95% B. The instrument was equilibrated for 4 min after every run, and the nebulizer, gas flow, gas temperature, capillary voltage, sheath gas temperature, and sheath gas flow were set to be 20 psi, 11 L/min, 260 °C, 3500 V, 300 °C, and 12 L/min, respectively. An Acquity UPLC BEH C₁₈ column (50, 2.1 mm i.d., 1.7 μ m thickness, Waters, Milford, MA) was used for the separation of other MEL derivatives (Tables S1 and S2) at 40 °C. Mobile phases included 0.1% formic acid in water (mobile phase A) and 0.1% formic acid in methanol (mobile phase B), and the flow rate was 0.4 mL/min. The gradient was as follows: 0 min, 30% B; 0.5 min, 30% B; 3.5 min, 70% B; 8 min, 74% B; 14 min, 100% B; and 15.5 min, 100% B. The nebulizer, gas flow, gas temperature, capillary voltage, sheath gas temperature, and sheath gas flow were set to be 40 psi, 10 L/min, 260 °C, 3500 V, 375 °C, and 12 L/min, respectively.

Table 1. Detection Frequencies (DF, %) and Median Concentrations of MEL-Based Compounds and PFAS in Infant Clothing (ng/g) [Calculated for Concentrations above the Method Detection Limit (MDL)]

	cotton		organic cotton		cotton/polyester		cotton/spandex		polyester		nylon		overall	
	DF	median	DF	median	DF	median	DF	median	DF	median	DF	median	DF	median
MEL	95	36.4	100	20.8	95	43.7	61	16.6	64	9.70	100	17,300	85	35.7
CYA	58	48.8	100	5990	40	15.6	28	32.8	27	15.0	38	171	47	77.6
AMD	58	2.47	50	42.1	20	10.3	22	56.0	9	3.74	75	401	36	28.2
AMN	16	708	<MDL		20	270	6	262	18	219	100	14,900	21	1530
TBMMAT	79	0.631	<MDL		20	0.133	11	0.503	27	0.0835	100	50.3	37	0.451
Σ MEL		81.4		6120		61.1		23.9		6.20		32,800		78.2
PFAS														
Short-Chain Ionic														
PFBA	<MDL		100	0.707	<MDL		<MDL		<MDL		<MDL		12	0.707
PFBS	11	0.0965	<MDL		15	0.0769	6	0.162	9	0.0656	25	0.251	11	0.0775
PFHxA	26	0.116	<MDL		35	0.107	44	0.0797	55	0.142	50	0.138	35	0.110
PFHxS	63	0.282	100	0.564	65	0.309	61	0.297	64	0.377	100	0.306	71	0.329
PFHpA	37	0.0777	20	0.0329	45	0.0845	33	0.0893	36	0.0696	38	0.0646	36	0.0765
Long-Chain Ionic														
PFOA	68	0.704	90	0.610	65	0.662	67	0.517	64	0.739	50	0.671	67	0.614
PFOS	11	0.466	<MDL		5	0.229	<MDL		<MDL		<MDL		3	0.432
PFNA	16	0.140	<MDL		<MDL		<MDL		<MDL		25	0.184	6	0.157
PFDA	58	0.206	80	0.161	55	0.242	56	0.293	36	0.156	25	0.289	54	0.220
PFUGA	89	0.475	100	0.484	95	0.520	100	0.397	82	0.410	75	0.540	92	0.479
PFDoA	53	0.248	100	0.164	45	0.316	61	0.213	27	0.301	63	0.366	56	0.230
PFTtDA	79	0.242	90	0.198	65	0.261	83	0.268	73	0.266	13	0.493	71	0.242
PFTeDA	95	0.331	100	0.376	100	0.443	100	0.319	100	0.447	100	0.362	99	0.367
PFHxDA	100	1.07	100	0.855	100	1.47	100	1.04	100	1.39	100	1.39	100	1.17
Σ ionic PFAS		3.27		4.02		3.46		2.91		3.28		3.54		3.42
Neutral														
6:2 FTOH	<MDL		<MDL		<MDL		6	1.84	27	17.3	38	149	8	27.1
8:2 FTOH	<MDL		<MDL		<MDL		17	10.4	<MDL		63	56.7	9	27.2
10:2 FTOH	<MDL		<MDL		<MDL		<MDL		<MDL		25	12.2	2	12.2
Σ neutral PFAS								10.4		17.3		89.5		41.9
Σ PFAS		3.27		4.02		3.46		3.19		3.28		86.9		3.62

The injection volume was 5 μL . The details of multiple reaction ion monitoring transitions, fragmentors, and collision energies for all analytes are provided in Tables S1 and S2.

Quality Assurance and Control. Procedural blanks ($n = 7$) and matrix spike recovery samples ($n = 7$) were analyzed with every batch of 12 samples. The average absolute matrix spike recoveries were 94 ± 18 , 85 ± 19 , and $65 \pm 20\%$ (mean \pm standard deviation) for MEL, ionic, and neutral PFAS compounds, respectively. The average recoveries for MEL and PFAS surrogate standards in different fabrics are provided in Table S4. Blank levels of target compounds constituted less than 1% of the respective levels in samples, on an average. All data were blank-corrected by subtracting blank concentrations from sample concentrations. Method detection limits (MDLs) were set as 3 times the standard deviation of the target analyte levels detected in the blanks and ranged from 0.01 to 0.35 ng/g. For compounds not detected in the blanks, MDLs were based on a signal-to-noise ratio of three. Quantification of target analytes was performed by isotope dilution using calibration curves with concentration ranges of 1–500 ng/mL. Correlation coefficients in linearity tests were all >0.99 , and samples with concentrations exceeding the linearity ranges were diluted to achieve the levels within the concentration ranges of the calibration curves.

Data Analysis. The removal rate of a chemical after the laundering experiment was calculated using eq 1

$$\text{Removal rate (\%)} = \frac{(C_{\text{before}} - C_{\text{after}}) \times 100\%}{C_{\text{before}}} \quad (1)$$

where C_{before} is the concentration of a chemical in the fabric before laundering (ng/g) and C_{after} is the concentration of a chemical in the fabric after laundering (ng/g).

Estimated daily intakes (EDIs, pg/kg body weight [bw]/day [d]) via dermal absorption of target chemicals from non-washed and washed clothing were calculated using eqs 2 and 3, respectively

$$\text{EDI}_{\text{non-washed}} = \frac{(C \times D \times SA \times F_{\text{mig}} \times F_{\text{contact}} \times F_{\text{pen}} \times N) \times T}{\text{BW}} \quad (2)$$

$$\text{EDI}_{\text{washed}} = \frac{(C \times (1 - \text{CC}/100) \times D \times SA \times F_{\text{mig}} \times F_{\text{contact}} \times F_{\text{pen}} \times N) \times T}{\text{BW}} \quad (3)$$

where C is the median concentration of a chemical in the fabric (ng/g), D is the density of the fabric (23.4 mg/cm^2), SA is the skin contact surface area (2040, 2310, 2660, and 3160 cm^2 for <1, 1–3, 3–6, and 6–12 month old infants, respectively),³³ F_{mig} is the migration rate of a chemical to the skin per day (0.005/day),³⁴ F_{contact} is the fraction of contact skin area (1, unitless),³⁴ F_{pen} is the penetration rate of a chemical (0.01, unitless),³⁴ T is the contact time between the fabric and skin (assumed to be 1 day),¹⁸ N is the average number of events per day (assumed to be 1/day),¹⁸ BW is the body weight (4.8, 5.9, 7.4, and 9.2 kg for <1, 1–3, 3–6, and 6–12 month old infants, respectively³³), and CC is the change in the concentration after laundering (%).

Basic and descriptive statistics were calculated using Minitab 19. Plots were generated using SigmaPlot 13 (Systat Software Inc.). Analysis of variance (ANOVA) was performed on log-transformed concentrations. The results of ANOVA are shown as letters; the concentrations of analytes sharing the same letters are not significantly different at $p < 0.05$.

RESULTS AND DISCUSSION

Melamine (MEL) and Its Derivatives. Five out of 11 targeted MEL-based compounds were detected in clothing items analyzed in this study. The other six analytes (Table S1) were not detected and are not included in the further discussion. MEL was detected in 85% of the samples, while the rest of the compounds were detected in up to 47%. The ΣMEL concentrations ranged from <MDL to 250,000 ng/g with a median concentration of 78.2 ng/g (Table 1). Ammeline (AMN) was the most abundant chemical with a median concentration of 1530 ng/g [calculated for concentrations above the MDL], although it was detected only in 21% of the samples. AMN-based resins, such as the AMN–formaldehyde resin, can be used in textiles along with MEL-based resins, which can be a possible explanation for this finding.³⁵ The concentrations of cyanuric acid (CYA), MEL, and ammelide (AMD) (medians 77.6, 35.7, and 28.2 ng/g, respectively) were lower than those of AMN, but the concentrations of these MEL-based compounds were not statistically different from one another (Figure 1). 2,4,6-Tris[bis(methoxymethyl)amino]-1,3,5-triazine (TBMMAT) was detected in 37% of the samples and was the least abundant compound among MEL derivatives (median 0.451 ng/g).

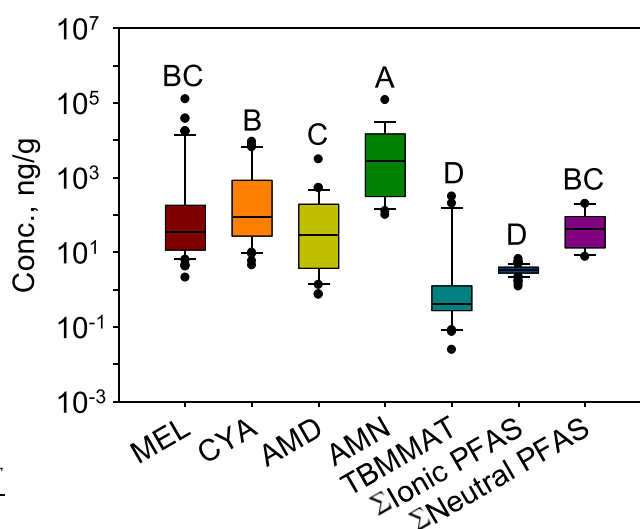


Figure 1. Concentrations of MEL and its derivatives, $\Sigma\text{ionic PFAS}$, and $\Sigma\text{neutral PFAS}$ in infant clothes (ng/g). Concentrations are shown as boxplots representing the 25th and 75th percentiles; black lines represent the median; and the whiskers represent the 10th and 90th percentiles. The letters represent the results of ANOVA; concentrations are ranked from the highest to the lowest in alphabetic order, and boxes sharing the same letters are not significantly different at $p < 0.05$.

MEL concentrations in this study were comparable to those previously reported in textiles and infant clothes from the United States (Table S5, median 53 ng/g)³¹ and benzothiazoles in unfinished textiles and infant clothing (median 51.1 ng/g).³⁶ These concentrations were higher than the concentrations of bisphenol-A (BPA) and bisphenol-S (BPS) in clothes from China (medians 26.9 and 7.38 ng/g, respectively)³⁷ but lower than the BPA levels detected in children's socks (median 396 ng/g),²⁹ BPS levels in pantyhose (median

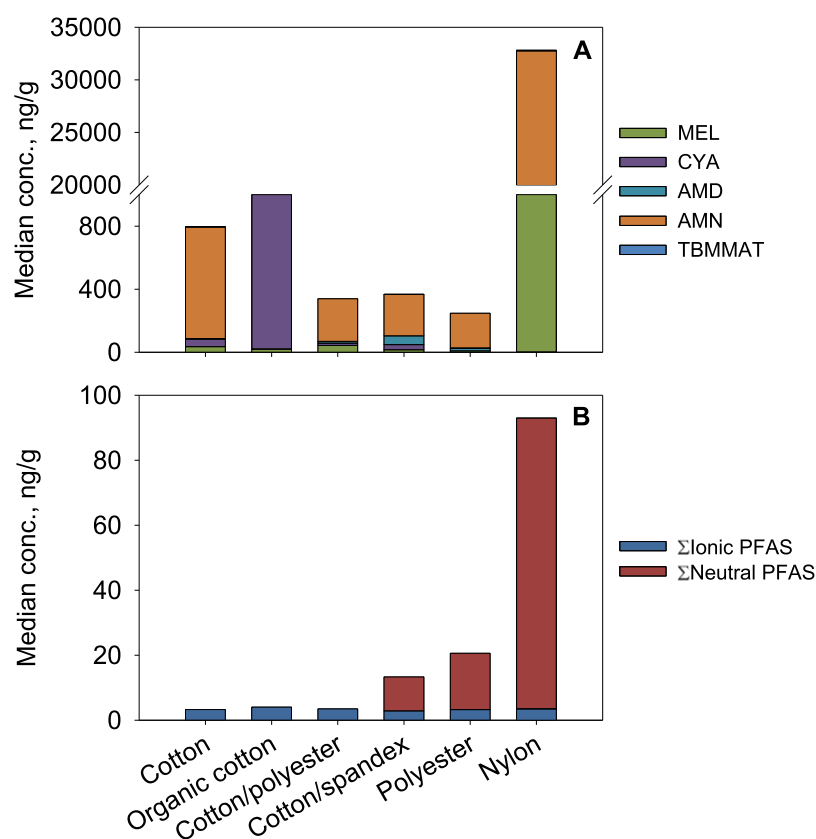


Figure 2. Median concentrations of (A) MEL-based compounds and (B) PFAS in different fabrics (ng/g).

1430 ng/g),³⁸ and phthalates in cotton infant clothing (690–2740 ng/g).³⁹

When grouped based on the fabric type, the highest Σ MEL concentrations were found in clothing made of nylon ($n = 8$) ranging from 6150 to 250,000 ng/g with a median concentration of 32,800 ng/g (Table 1). The Σ MEL levels in nylon and organic cotton were significantly higher ($p < 0.05$; Figure 2) than those in other fabric types including cotton, cotton mixed with polyester (cotton/polyester), cotton mixed with spandex (cotton/spandex), and polyester. The Σ MEL concentrations in other fabric types were statistically indistinguishable from one another. MEL and AMN were the most abundant chemicals found in nylon (medians 17,300 and 14,900 ng/g, respectively). The concentrations of these compounds in nylon were 2–4 orders of magnitude higher than those in other fabric types (Table 1), suggesting that MEL and its derivatives may be used in the manufacturing of nylon clothing to enhance water-repellent properties and thermal stability.^{40–42} In fact, the process of using MEL–formaldehyde resins in nylon finishing for imparting water-repellent properties has been patented,⁴³ and the amount of MEL in nylon as a result of these treatments can reach up to 50% of a fabric's dry weight.⁴³ Hence, the high levels of MEL found in nylon samples may be due to the intentional use of MEL in this type of fabric.

Interestingly, organic cotton also had high Σ MEL concentrations ranging from 96.2 to 9220 ng/g (median 6120 ng/g). These Σ MEL concentrations mostly comprised of CYA with concentrations of up to 9120 ng/g, which was more than 2 orders of magnitude higher than that in conventional cotton (<MDL—471 ng/g). Commercial nitrogen-based fertilizers, such as urea, nitrolime, and ammonia-

based natural fertilizers, contain high levels of CYA (up to 250,000 ng/g) relative to MEL (up to 90,400 ng/g)⁴⁴ and are used in growing organic cotton, which must meet strict federal regulations on the use of less toxic pesticides and fertilizers.⁴⁵ Heavy use of nitrogen-based fertilizers containing large amounts of CYA may result in plant uptake^{44,46,47} and possibly contribute to CYA levels in organic cotton.

The Σ MEL concentrations in clothing labeled as 100% cotton ranged from 2.51 to 1770 ng/g with a median concentration of 81.4 ng/g. It has been reported that textiles labeled as “100% cotton” usually contain 77% cotton, 10% shrink-resistance resins, and 4% dyes and other additives.¹ The median concentrations of Σ MEL were 61.1, 23.9, and 6.20 ng/g for cotton/polyester, cotton/spandex, and polyester clothes, respectively. The Σ MEL concentrations in polyester and spandex clothing were statistically indistinguishable from each other, perhaps because of the limited sample size of each group.

When grouped based on clothing type (outerwear, diapers, dresses, onesies, pants, shirts, and socks), the highest Σ MEL concentrations were found in outerwear (median 70,800 ng/g; Figure 3 and Table S6). The outerwear items were made of nylon, which may be coated with MEL–urea–formaldehyde resins to increase the water-repellency and heat resistance of the fabric.^{41,48} Cloth diapers were the second category that had high Σ MEL concentrations (median 4570 ng/g, Table S6), which is concerning considering that diapers are in close contact with skin and worn almost constantly during the first 12–24 months of life.

The Σ MEL concentrations in clothes manufactured in the United States (median 6120 ng/g) were significantly higher than those from other countries ($p < 0.05$; Figure S2 and Table

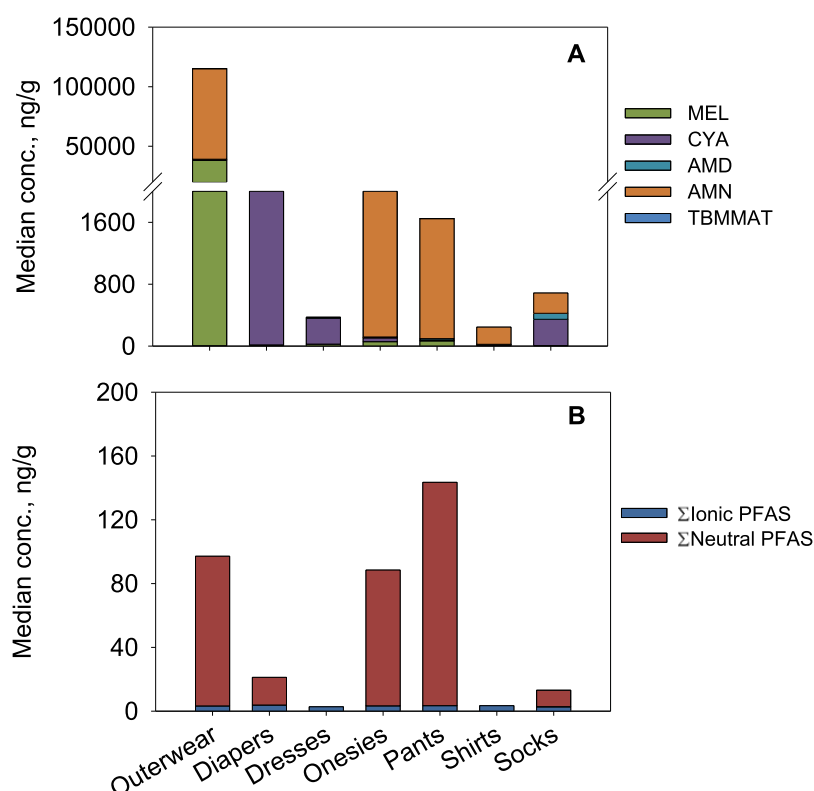


Figure 3. Median concentrations of (A) MEL-based compounds and (B) PFAS in different clothing categories (ng/g).

S7). However, it should be noted that because of the difficulty of finding clothes manufactured in the United States, these samples only included cloth diapers made of organic cotton. No significant differences in Σ MEL concentrations were found among clothing items manufactured in other countries. Overall, our findings suggest that fabric composition was the major factor affecting the concentrations of MEL-based compounds in infant clothing.

PFAS. Among 40 targeted PFAS, 17 were detected (Table 1). These included five short-chain (C4–C7) and nine long-chain (C8–C16) ionic PFAS. In addition, three neutral FTOHs were also detected (Table 1). Detection frequencies for these PFAS ranged from 2 to 100%, and at least, one PFAS was detected in each sample. The Σ PFAS concentrations (the sum of all 17 detected PFAS concentrations) ranged from 1.22 to 203 ng/g with a median concentration of 3.62 ng/g (Table 1), which was up to 3 orders of magnitude lower than the levels previously reported in performance clothes (2400–219,000 ng/g).^{11,49} While FTOHs were detected at the highest levels among the measured PFAS (median 41.9 ng/g), they were found in only 2–9% of the samples. The FTOH concentrations in this study were up to 3 orders of magnitude lower than those reported for outdoor adult clothing (2380–219,000 ng/g).^{11,32,49} Interestingly, perfluorooctane sulfonamides and perfluorooctane sulfonamidoethanols, the two PFAS groups that used to be applied in electrochemical fluorination textile treatment techniques before the 2000s,⁵⁰ were not detected in these samples.

In contrast, ionic PFAS were detected in most of the samples, but at lower concentrations. Overall, the median Σ ionic PFAS (3.42 ng/g) was up to 10 times lower than the median Σ neutral PFAS concentration (41.9 ng/g). Concentrations of ionic PFAS ranged from 1.22 to 6.45 ng/g and were

similar to previous reports.^{50,51} Long-chain (C11–C15) PFAS were frequently detected, also similar to previous studies.^{5,10,52,53} The C16 homologue was the most abundant ionic PFAS at a median concentration of 1.17 ng/g. PFOS was detected in only 3% of the samples at a median concentration of 0.432 ng/g, while PFOA was found in 67% of the samples at a higher median of 0.614 ng/g. Short-chain ionic PFAS with 4–7 carbons were found in 11–71% of the samples, but at low concentrations (medians 0.0765–0.707 ng/g). The concentrations of long-chain ionic PFAS were significantly higher than the levels of the short-chain PFAS compounds (Figure S3, $p < 0.05$).

The highest Σ PFAS concentrations were found in clothing made of nylon (Figure 2, $p < 0.05$) and ranged from 4.23 to 203 ng/g with a median concentration of 86.9 ng/g (Table 1 and Figure 2). The Σ PFAS concentrations in nylon were dominated by FTOHs. Among FTOHs, 6:2 FTOH was the most abundant with a median concentration of 149 ng/g. In studies from the early 2000s, high FTOH levels (200,000–9,000,000 μ g/L) were found in textile impregnating fluids¹⁷ and in waterproofing products (up to 3.8% by weight).⁵⁴ The levels in our study were much lower compared to these earlier studies and suggest that PFAS in clothing items in this study were either added at much lower levels or due to the impurities in manufacturing processes or atmospheric deposition.

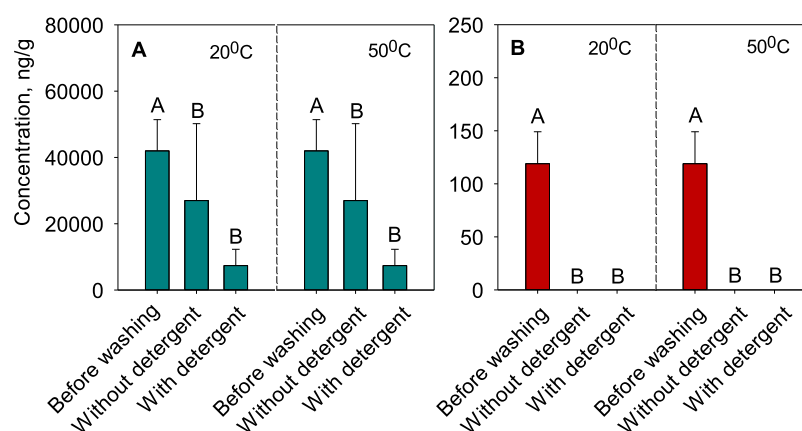
When grouped based on clothing type, the highest Σ PFAS concentrations were found in pants (median 140 ng/g; Figure 3 and Table S6), followed by outerwear (median 93.8 ng/g). No significant differences in PFAS levels were found based on the clothing type and manufacturing origin (Figures 3 and S2).

A recent study from Zhu and Kannan measured ionic PFAS in infant clothing from the United States and reported a median concentration of ionic PFAS at 0.953 ng/g.³² While

Table 2. Σ MEL and Σ PFAS Concentrations (Mean \pm Standard Error, ng/g) before and after Laundering and the Removal Rate (%) as a Result of Washing^a

conditions	Σ MEL	removal rate	Σ PFAS	removal rate
before washing	42,000 \pm 9410		119 \pm 30.1	
washing, 20 °C	27,000 \pm 23,200	56.6 \pm 31.6	<MDL	100
washing, 20 °C, detergent	7400 \pm 4900	86.2 \pm 6.46	<MDL	100
washing, 50 °C	3090 \pm 2960	95.4 \pm 4.17	<MDL	100
washing, 50 °C, detergent	1800 \pm 1610	97.2 \pm 2.23	<MDL	100

^a Σ MEL represents the sum of MEL and AMN concentrations as these two compounds were the most abundant in samples used for this experiment. MDL: method detection limit.

**Figure 4.** Mean (with standard errors) Σ MEL (A) and Σ PFAS concentrations (B) in clothes washed in different conditions (ng/g). The letters represent the results of ANOVA; the concentrations are ranked from the highest to the lowest in alphabetic order; and the bars sharing the same letters are not significantly different at $p < 0.05$.

neutral PFAS were not directly measured in the latter study, the total oxidizable precursor (TOP) assay was applied to test for possible PFAS precursors,³² resulting in a 10-fold increase in the concentrations of the ionic PFAS after the TOP assay and suggesting that PFAS precursors were present in the analyzed clothing samples. Our study further supports these findings and suggests that these precursors may include FTOHs, which were found at high levels in several clothing items analyzed in this study.

Overall, both MEL derivatives and PFAS were found in all of the clothing items analyzed in this study. However, the detection frequencies and concentrations of MEL derivatives were significantly higher than those of PFAS ($p < 0.05$), suggesting potential higher use of MEL derivatives in textile production. There is a general trend in the textile industry to move away from using fluorine-based applications for grease-, stain-, and waterproof fabric finishing, while still achieving desired end results.⁵ According to U.S. EPA's Chemview database, more than 30 companies have reported the use of MEL as a textile surface coating.⁵⁵ Interestingly, more than 15,000 patents on textile finishing using MEL-based compounds can be found as a result of a Google Patent search. The production volumes of MEL resins are not openly available, but the production amounts of individual MEL-based compounds that are used as components of these mixtures range from 23,000 to 120,000 tons/year in the United States.⁵⁵

Our findings suggest that textiles can be a significant source of human exposure to MEL-based compounds. MEL has generally been recognized as a less toxic alternative for many textile finishing agents and specifically for long-chain PFAS.²⁰ However, the European Chemical Agency is currently considering classifying MEL as a bladder toxin and

carcinogen.⁵⁶ In 2019, the International Cancer Research Agency (IARC) classified MEL as a possible carcinogen to humans.⁵⁷ The German Chemical Society considers MEL as a persistent, mobile, and toxic (PMT) compound similar to tris(2-chloro-1-methylethyl)phosphate and 1,4-dioxane.^{58,59} When combined, MEL and CYA can crystallize in kidneys and induce renal damage and failure.^{60–62} Recent studies found MEL and CYA in breast milk⁶³ and children's urine,⁶⁴ suggesting widespread exposure in early life. The urinary levels of MEL and CYA in children were higher than those in adults, indicating higher exposures in children,^{64,65} and were suggestive of kidney damage at current exposure levels.⁶⁴

Simulated Laundering. Clothing items with the highest Σ MEL and Σ PFAS concentrations were included in a simulated laundering experiment in order to evaluate the release of MEL and PFAS during the laundry process under different conditions, including washing with and without detergent in cool (20 °C) and warm (50 °C) water. Σ MEL and Σ PFAS concentrations in fabric decreased significantly after washing, especially when washed in warm water (Table 2). When washed in cool water, the average Σ MEL concentrations decreased on an average by $\sim 60\%$, from 42,000 \pm 9410 to 27,000 \pm 23,200 ng/g (Table 2 and Figure 4), while when washed in warm water, the Σ MEL concentrations decreased on an average by $\sim 95\%$ to 3090 \pm 2960 ng/g, which was significantly lower than the concentrations before laundering ($p < 0.05$). The Σ MEL concentrations decreased by an additional 30% when washed in cool water with detergent (7400 \pm 4900 ng/g; Table 2 and Figure 4), indicating that the application of laundry detergent facilitates the release of MEL-based compounds from clothes into water. Σ MEL further decreased by only $\sim 2\%$ when

Table 3. EDIs (pg/kg bw/day) of Σ PFAS, MEL, and CYA via Dermal Absorption from Clothing (Not Washed and Washed at Different Temperatures with Detergent) for 0–1, 1–3, 3–6, and 6–12 Month Old Infants (Based on Median Concentrations)

age (months)	not washed				washed, 20 °C, detergent				washed, 50 °C, detergent			
	0–1	1–3	3–6	6–12	0–1	1–3	3–6	6–12	0–1	1–3	3–6	6–12
Non-Nylon												
Σ PFAS	1.64	1.51	1.39	1.33	0	0	0	0	0	0	0	0
MEL	15.0	13.8	12.7	12.1	2.07	1.91	1.75	1.67	0.0580	0.0534	0.0491	0.0469
CYA	29.2	26.9	24.7	23.6	4.02	3.71	3.40	3.25	0.113	0.104	0.0953	0.0911
Nylon												
Σ PFAS	44.5	41.0	37.6	36.0	0	0	0	0	0	0	0	0
MEL	8.590	7.910	7.260	6.940	1.190	1.090	1.000	958	33.2	30.6	28.1	26.8
CYA	85.1	78.4	72.0	68.8	11.7	10.8	9.93	9.49	0.329	0.303	0.278	0.266

washed in warm water with detergent (1800 ± 1610 ng/g). Similar levels of MEL released into wastewater were reported for MEL-based tableware^{66–68} and may be attributed to high solubilities of MEL and its derivatives ($3.24\text{--}1000$ mg/L).⁵⁴

In contrast, PFAS were completely washed out after laundering in cool water without detergent and were not detected in any of the samples after washing (Table 2 and Figure 4). Strong migration of PFAS has also been reported in previous studies.^{9,52} Leaking of chemicals from clothing to water during laundry is significantly correlated with a chemical's K_{ow} .⁶⁹ For example, as much as 80% of organophosphate esters with $\log K_{ow} < 4$ leaked from clothing to water in a simulated laundry experiment, while the leakage rate was only 10% for brominated flame retardants with $\log K_{ow} > 6$.⁶⁹ The high release rates for both MEL and PFAS suggest that treated clothes and other textiles can be an important source of these chemicals to wastewater treatment plants and eventually to surface waters.

Exposure Assessment. EDIs of Σ PFAS, MEL, and CYA via dermal absorption from clothes for 0–1, 1–3, 3–6, and 6–12 month old infants are presented in Table 3. The Σ PFAS EDI from non-nylon clothes ranged from 1.33 to 1.64 pg/kg bw/d, which was 10–20 times lower than that for MEL and CYA. For nylon clothes, these EDIs increased to 36.0–44.5 pg/kg bw/d. The MEL EDI from nylon clothes was about 3 orders of magnitude higher than that from non-nylon clothing. The highest PFAS, MEL, and CYA EDIs were estimated for 0–1 month old infants, suggesting that newborns are the highest exposure risk group. The EDIs from infant clothing for BPA were estimated as 201–248 pg/kg bw/d²⁹ and were similar to the EDI for MEL from non-nylon clothes in this study (12.1–15.0 pg/kg bw/d) and much lower than the MEL EDI from nylon clothes (6940–8590 pg/kg bw/d). The EDIs for MEL and CYA via dermal absorption were up to 4–100 times lower than those from breastfeeding, consumption of infant formula, and dust ingestion (Table S8).^{63,70,71} The EDIs calculated using concentrations in clothes after washing decreased by more than half for MEL and CYA and 100% for Σ PFAS. Lowest EDIs were determined in samples washed with detergent at 50 °C, suggesting that laundering in warm water using a detergent could be a suitable way to reduce dermal exposure to chemicals in clothes, but it raises concerns about the fate of these chemicals in wastewater treatment processes.

Limitations and Implications. This study has several limitations. The sample size of each fabric and clothing type was limited, especially for clothes made of nylon. Further studies on different categories of nylon clothing are needed to

verify our findings of high levels of MEL in this fabric type. The absorption of PFAS and MEL-based compounds from the indoor environment onto textiles cannot be ruled out but was not evaluated in this study. In addition, the laundering experiment was performed using nonverified simulators, and no repeat washes were performed. The extraction methods used in this study did not include different exhaustive extraction methods (e.g., TOP assay), which may result in underestimation of the nonextractable MEL and PFAS compounds.

Nevertheless, this is the first comprehensive study that reports two groups of textile additives, MEL and PFAS compounds, in infant clothing. All clothing items analyzed in this study had significantly higher levels of MEL-based compounds than PFAS, suggesting a potential use of MEL and its derivatives in infant clothes and possibly other textiles. Although MEL-based formulations are considered as safe PFAS alternatives by the textile industry, the European Chemical Agency⁵⁶ and the International Cancer Research Agency have recently listed MEL as a possible human carcinogen.⁵⁷ Clearly, further research is needed on risks associated with existing environmental exposures to this group of chemicals.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.est.0c03035>.

Target analytes; sample details; summary of previously published data on emerging contaminants in textiles; representative chromatograms of FTOHs; concentrations of MEL-based compounds and PFAS in different categories of clothing items; details of exposure assessment; and PFAS composition in clothes (PDF)

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Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We thank Christina Gu and Emma-Li Kissinger for help with sample collection and processing.

REFERENCES

- (1) Lacasse, K.; Baumann, W. *Textile Chemicals: Environmental Data and Facts*; Springer Berlin Heidelberg: New York, 2004.
- (2) O Ecotextiles. Choosing a Fabric for Your New Sofa. <https://oecotextiles.wordpress.com/tag/bisphenol-a/> (accessed Sept 23, 2019).
- (3) Rovira, J.; Domingo, J. L. Human health risks due to exposure to inorganic and organic chemicals from textiles: A review. *Environ. Res.* **2019**, *168*, 62–69.
- (4) Schellenberger, S.; Hill, P. J.; Levenstam, O.; Gillgard, P.; Cousins, I. T.; Taylor, M.; Blackburn, R. S. Highly fluorinated chemicals in functional textiles can be replaced by re-evaluating liquid repellency and end-user requirements. *J. Cleaner Prod.* **2019**, *217*, 134–143.
- (5) The Danish Environmental Protection Agency. *Polyfluoroalkyl Substances (PFASs) in Textiles for Children, Survey of Chemical Substances in Consumer Products* No. 136, 2015.
- (6) Supreeyasunthorn, P.; Boontanon, S. K.; Boontanon, N. Perfluorooctane sulfonate (PFOS) and perfluorooctanoic acid (PFOA) contamination from textiles. *J. Environ. Sci. Health, Part A: Toxic/Hazard. Subst. Environ. Eng.* **2016**, *51*, 472–477.
- (7) Zheng, G.; Boor, B. E.; Schreder, E.; Salamova, A. Indoor exposure to per- and polyfluoroalkyl substances (PFAS) in the childcare environment. *Environ. Pollut.* **2020**, *258*, 113714.
- (8) Beesoon, S.; Genuis, S. J.; Benskin, J. P.; Martin, J. W. Exceptionally high serum concentrations of perfluorohexanesulfonate in a canadian family are linked to home carpet treatment applications. *Environ. Sci. Technol.* **2012**, *46*, 12960–12967.
- (9) Hill, P. J.; Taylor, M.; Goswami, P.; Blackburn, R. S. Substitution of PFAS chemistry in outdoor apparel and the impact on repellency performance. *Chemosphere* **2017**, *181*, 500–507.
- (10) Gremmel, C.; Frömel, T.; Knepper, T. P. Systematic determination of perfluoroalkyl and polyfluoroalkyl substances (PFASs) in outdoor jackets. *Chemosphere* **2016**, *160*, 173–180.
- (11) Kotthoff, M.; Müller, J.; Jüriling, H.; Schlummer, M.; Fiedler, D. Perfluoroalkyl and polyfluoroalkyl substances in consumer products. *Environ. Sci. Pollut. Res.* **2015**, *22*, 14546–14559.
- (12) Field, J. A.; Seow, J. Properties, occurrence, and fate of fluorotelomer sulfonates. *Crit. Rev. Environ. Sci. Technol.* **2017**, *47*, 643–691.
- (13) Wang, Z.; DeWitt, J. C.; Higgins, C. P.; Cousins, I. T. A never-ending story of per- and polyfluoroalkyl substances (PFASs)? *Environ. Sci. Technol.* **2017**, *51*, 2508–2518.
- (14) Wang, Z.; Cousins, I. T.; Scheringer, M.; Hungerbühler, K. Fluorinated alternatives to long-chain perfluoroalkyl carboxylic acids (PFCAs), perfluoroalkane sulfonic acids (PFASs) and their potential precursors. *Environ. Int.* **2013**, *60*, 242–248.
- (15) Wang, Z.; Cousins, I. T.; Scheringer, M.; Buck, R. C.; Hungerbühler, K. Global emission inventories for C4–C14 perfluoroalkyl carboxylic acid (PFCA) homologues from 1951 to 2030, Part I: Production and emissions from quantifiable sources. *Environ. Int.* **2014**, *70*, 62–75.
- (16) Knepper, T. P.; Frömel, T.; Gremmel, C.; Driezum, I. v.; Weil, H.; Vestergren, R.; Cousins, I. T. Understanding the Exposure Pathways of Per- and Polyfluoroalkyl Substances (PFASs) via Use of PFASs-Containing Products—Risk Estimation for Man and Environment; Federal Environment Agency Germany. 2014, <http://www.umweltbundesamt.de/publikationen/understanding-the-exposure-pathways-of-per> (accessed Sept 23, 2019).
- (17) Herzke, D.; Posner, S.; Olsson, E. Survey, Screening and Analyses of PFCs in Consumer Products. TA-2578/2009. Swerea/IVF Project report 09/47. 2009, https://tema.miljodirektoratet.no/no/Publikasjoner/Publikasjoner/2011/Mars/Survey_screening_and_analyses_of_PFCs_in_consumer_products/ (accessed Sept 23, 2019).
- (18) Rovira, J.; Nadal, M.; Schuhmacher, M.; Domingo, J. L. Human exposure to trace elements through the skin by direct contact with clothing: Risk assessment. *Environ. Res.* **2015**, *140*, 308–316.
- (19) Sunderland, E. M.; Hu, X. C.; Dassuncao, C.; Tokranov, A. K.; Wagner, C. C.; Allen, J. G. A review of the pathways of human exposure to poly- and perfluoroalkyl substances (PFASs) and present understanding of health effects. *J. Exposure Sci. Environ. Epidemiol.* **2019**, *29*, 131–147.
- (20) Water Repellency Project. Durable Water and Soil Repellent Chemistry in the Textile Industry—A Research Report. 2012, https://outdoorindustry.org/wp-content/uploads/2015/05/FINAL_ZDHC_P05_DWR-Research_Nov20121.pdf (accessed Sept 23, 2019).
- (21) Chen, J.-s. A worldwide food safety concern in 2008—Melamine-contaminated infant formula in China caused urinary tract stone in 290 000 children in China. *Chin. Med. J.* **2009**, *122*, 243–244.
- (22) Dobson, R. L. M.; Motlagh, S.; Quijano, M.; Cambron, R. T.; Baker, T. R.; Pullen, A. M.; Regg, B. T.; Bigalow-Kern, A. S.; Vennard, T.; Fix, A.; Reimschuessel, R.; Overmann, G.; Shan, Y.; Daston, G. P. Identification and characterization of toxicity of contaminants in pet food leading to an outbreak of renal toxicity in cats and dogs. *Toxicol. Sci.* **2008**, *106*, 251–262.
- (23) Gossner, C. M.-E.; Schlundt, J.; Ben Embarek, P.; Hird, S.; Lo-Fo-Wong, D.; Beltran, J. J. O.; Teoh, K. N.; Tritscher, A. The melamine incident: Implications for international food and feed safety. *Environ. Health Perspect.* **2009**, *117*, 1803–1808.
- (24) Hauser, W. D. S. P. J. *Chemical Finishing of Textiles*; Woodhead Publishing, 2004.
- (25) Rovina, K.; Siddiquee, S. A review of recent advances in melamine detection techniques. *J. Food Compos. Anal.* **2015**, *43*, 25–38.
- (26) Weil, E. D.; Levchik, S. V. Flame retardants in commercial use or development for textiles. *J. Fire Sci.* **2008**, *26*, 243–281.
- (27) Cantú, R.; Evans, O.; Kawahara, F. K.; Shoemaker, J. A.; Dufour, A. P. An HPLC method with UV detection, pH control, and reductive ascorbic acid for cyanuric acid analysis in water. *Anal. Chem.* **2000**, *72*, 5820–5828.
- (28) Molander, S.; Kristin, F.; Johan, T.; Haglund, P.; Holmgren, T.; Tomas, R.; Jenny, W. Calculating the Swedish economy wide emissions of additives from plastic materials. Poster Present. 33rd Annual Meeting SETAC North America, 2012.
- (29) Xue, J.; Liu, W.; Kannan, K. Bisphenols, benzophenones, and bisphenol A diglycidyl ethers in textiles and infant clothing. *Environ. Sci. Technol.* **2017**, *51*, 5279–5286.
- (30) Wu, Y.; Zhang, Y. Analytical chemistry, toxicology, epidemiology and health impact assessment of melamine in infant formula: Recent progress and developments. *Food Chem. Toxicol.* **2013**, *56*, 325–335.
- (31) Zhu, H.; Kannan, K. Determination of melamine and its derivatives in textiles and infant clothing purchased in the United States. *Sci. Total Environ.* **2020**, *710*, 136396.
- (32) Zhu, H.; Kannan, K. Total oxidizable precursor assay in the determination of perfluoroalkyl acids in textiles collected from the United States. *Environ. Pollut.* **2020**, *265*, 114940.
- (33) EPA. *Exposure Factors Handbook*; Washington, DC, 2011.
- (34) BfR. *Introduction to the Problems Surrounding Garment Textiles, Updated BfR Opinion No. 041/2012*, 2017; Vol 140, pp 308–316.
- (35) Ji, S.; Crews, G. M.; Pittman, C. U., Jr.; Wang, Y.; Ran, R. Ammeline—melamine—formaldehyde resins. Preparation and properties. *J. Polym. Sci., Part A: Polym. Chem.* **1996**, *34*, 2543–2561.
- (36) Liu, W.; Xue, J.; Kannan, K. Occurrence of and exposure to benzothiazoles and benzotriazoles from textiles and infant clothing. *Sci. Total Environ.* **2017**, *592*, 91–96.

- (37) Wang, L.; Zhang, Y.; Liu, Y.; Gong, X.; Zhang, T.; Sun, H. Widespread occurrence of bisphenol A in daily clothes and its high exposure risk in humans. *Environ. Sci. Technol.* **2019**, *53*, 7095–7102.
- (38) Li, A. J.; Kannan, K. Elevated concentrations of bisphenols, benzophenones, and antimicrobials in pantyhose collected from six countries. *Environ. Sci. Technol.* **2018**, *52*, 10812–10819.
- (39) Li, H.-L.; Ma, W.-L.; Liu, L.-Y.; Zhang, Z.; Sverko, E.; Zhang, Z.-F.; Song, W.-W.; Sun, Y.; Li, Y.-F. Phthalates in infant cotton clothing: Occurrence and implications for human exposure. *Sci. Total Environ.* **2019**, *683*, 109–115.
- (40) Yang, H.; Yang, C. Q.; He, Q. The bonding of a hydroxy-functional organophosphorus oligomer to nylon fabric using the formaldehyde derivatives of urea and melamine as the bonding agents. *Polym. Degrad. Stab.* **2009**, *94*, 1023–1031.
- (41) Weber, B.; Bremser, W.; Hiltrop, K. Creating new materials with melamine resins. *Prog. Org. Coat.* **2009**, *64*, 150–155.
- (42) Mecker, L. C.; Tyner, K. M.; Kauffman, J. F.; Arzhantsev, S.; Mans, D. J.; Gryniwicz-Ruzicka, C. M. Selective melamine detection in multiple sample matrices with a portable Raman instrument using surface enhanced Raman spectroscopy-active gold nanoparticles. *Anal. Chim. Acta* **2012**, *733*, 48–55.
- (43) Roth, P. B. Method of Finishing Nylon Textile Material. U.S. Patent 3,197,270 A, 1962.
- (44) Zhu, H.; Wang, Y.; Sun, H.; Kannan, K. Fertilizers as a source of melamine and cyanuric acid in soils: A nationwide survey in China. *Environ. Sci. Technol. Lett.* **2019**, *6*, 55–61.
- (45) Gam, H. J.; Cao, H.; Farr, C.; Kang, M. Quest for the eco-apparel market: a study of mothers' willingness to purchase organic cotton clothing for their children. *Int. J. Consum. Stud.* **2010**, *34*, 648–656.
- (46) Terman, G. L.; Dement, J. D.; Hunt, C. M. Fertilizer nitrogen sources, crop response to urea and urea pyrolysis products. *J. Agric. Food Chem.* **1964**, *12*, 151.
- (47) Shimadzu. Analysis of Melamine and Its Related Substances in Fertilizers. 2017, <https://www.shimadzu.com/an/literature/hplc/jpl217002.html> (accessed Sept 23, 2019).
- (48) Fang, Z. H.; Duan, H. Y.; Zhang, Z. H.; Wang, J.; Li, D. Q.; Huang, Y. X.; Shang, J. J.; Liu, Z. Y. Novel heat-resistance UV curable waterborne polyurethane coatings modified by melamine. *Appl. Surf. Sci.* **2011**, *257*, 4765–4768.
- (49) Brigden, K.; Hetherington, S.; Wang, M.; Santillo, D.; Johnston, P. Hazardous Chemicals in Branded Textile Products on Sale in 25 Countries/Regions During 2013. Greenpeace Res. Lab. Tech. Rep. 2013, <http://www.greenpeace.to/greenpeace/wp-content/uploads/2014/02/Technical-Report-01-2014.pdf> (accessed Sept 23, 2019).
- (50) Robel, A. E.; Marshall, K.; Dickinson, M.; Lunderberg, D.; Butt, C.; Peaslee, G.; Stapleton, H. M.; Field, J. A. Closing the mass balance on fluorine on papers and textiles. *Environ. Sci. Technol.* **2017**, *51*, 9022–9032.
- (51) Tokranov, A. K.; Nishizawa, N.; Amadei, C. A.; Zenobio, J. E.; Pickard, H. M.; Allen, J. G.; Vecitis, C. D.; Sunderland, E. M. How do we measure poly- and perfluoroalkyl substances (PFASs) at the surface of consumer products? *Environ. Sci. Technol. Lett.* **2019**, *6*, 38–43.
- (52) Commission for Environmental Cooperation. Furthering the Understanding of the Migration of Chemicals from Consumer Products. 2017, <http://www3.cec.org/islandora/en/item/11777-furthering-understanding-migration-chemicals-from-consumer-products-en.pdf> (accessed Sept 23, 2019).
- (53) van der Veen, I.; Weiss, J. M.; Hanning, A.-C.; De Boer, J.; Leonards, P. E. G. Development and validation of a method for the quantification of extractable perfluoroalkyl acids (PFAAs) and perfluorooctane sulfonamide (FOSA) in textiles. *Talanta* **2016**, *147*, 8–15.
- (54) Dinglasan-Panlilio, M. J. A.; Mabury, S. A. Significant residual fluorinated alcohols present in various fluorinated materials. *Environ. Sci. Technol.* **2006**, *40*, 1447–1453.
- (55) EPA. Chemview. <https://chemview.epa.gov/chemview> (accessed Sept 23, 2019).
- (56) European Chemicals Agency. Melamine-Toxicological Summary. <https://echa.europa.eu/registration-dossier/-/registered-dossier/15978/7/8> (accessed Sept 23, 2019).
- (57) Grosse, Y.; Loomis, D.; Guyton, K. Z.; El Ghissassi, F.; Bouvard, V.; Benbrahim-Tallaa, L.; Mattock, H.; Straif, K. Some chemicals that cause tumours of the urinary tract in rodents. *Lancet Oncol.* **2017**, *18*, 1003–1004.
- (58) Chemsec. Chemicals Added to the SIN List. 2019, <https://sinlist.chemsec.org/the-new-sin-list-chemicals/> (accessed Sept 23, 2019).
- (59) Rüdél, H.; Körner, W.; Letzel, T.; Neumann, M.; Nödler, K.; Reemtsma, T. Persistent, mobile and toxic substances in the environment: a spotlight on current research and regulatory activities. *Environ. Sci. Eur.* **2020**, *32*, 5.
- (60) Puschner, B.; Poppenga, R. H.; Lowenstine, L. J.; Filigenzi, M. S.; Pesavento, P. A. Assessment of melamine and cyanuric acid toxicity in cats. *J. Vet. Diagn. Invest.* **2007**, *19*, 616–624.
- (61) Filigenzi, M. S.; Puschner, B.; Aston, L. S.; Poppenga, R. H. Diagnostic determination of melamine and related compounds in kidney tissue by liquid chromatography/tandem mass spectrometry. *J. Agric. Food Chem.* **2008**, *56*, 7593–7599.
- (62) Chang, L.; She, R.; Ma, L.; You, H.; Hu, F.; Wang, T.; Ding, X.; Guo, Z.; Soomro, M. H. Acute testicular toxicity induced by melamine alone or a mixture of melamine and cyanuric acid in mice. *Reprod. Toxicol.* **2014**, *46*, 1–11.
- (63) Zhu, H.; Kannan, K. Occurrence of melamine and its derivatives in breast milk from the United States and its implications for exposure in infants. *Environ. Sci. Technol.* **2019**, *53*, 7859–7865.
- (64) Sathyanarayana, S.; Flynn, J. T.; Messito, M. J.; Gross, R.; Whitlock, K. B.; Kannan, K.; Karthikraj, R.; Morrison, D.; Huie, M.; Christakis, D.; Trasande, L. Melamine and cyanuric acid exposure and kidney injury in US children. *Environ. Res.* **2019**, *171*, 18–23.
- (65) Zhu, H.; Kannan, K. Inter-day and inter-individual variability in urinary concentrations of melamine and cyanuric acid. *Environ. Int.* **2019**, *123*, 375–381.
- (66) Chien, C.-Y.; Wu, C.-F.; Liu, C.-C.; Chen, B.-H.; Huang, S.-P.; Chou, Y.-H.; Chang, A.-W.; Lee, H.-H.; Pan, C.-H.; Wu, W.-J.; Shen, J.-T.; Chang, M.-Y.; Huang, C.-H.; Shiea, J.; Hsieh, T.-J.; Wu, M.-T. High melamine migration in daily-use melamine-made tableware. *J. Hazard. Mater.* **2011**, *188*, 350–356.
- (67) Lund, K. H.; Petersen, J. H. Migration of formaldehyde and melamine monomers from kitchen- and tableware made of melamine plastic. *Food Addit. Contam.* **2006**, *23*, 948–955.
- (68) Wu, M.-T.; Wu, C.-F.; Chen, B.-H. Behavioral intervention and decreased daily melamine exposure from melamine tableware. *Environ. Sci. Technol.* **2015**, *49*, 9964.
- (69) Saini, A.; Thaysen, C.; Jantunen, L.; McQueen, R. H.; Diamond, M. L. From clothing to laundry water: Investigating the fate of phthalates, brominated flame retardants, and organophosphate esters. *Environ. Sci. Technol.* **2016**, *50*, 9289–9297.
- (70) Zhu, H.; Kannan, K. Continuing occurrence of melamine and its derivatives in infant formula and dairy products from the United States: Implications for environmental sources. *Environ. Sci. Technol. Lett.* **2018**, *5*, 641–648.
- (71) Zhu, H.; Kannan, K. Distribution profiles of melamine and its derivatives in indoor dust from 12 countries and the implications for human exposure. *Environ. Sci. Technol.* **2018**, *52*, 12801–12808.