

Organophosphate Esters and Their Metabolites in Breast Milk from the United States: Breastfeeding Is an Important Exposure Pathway for Infants

Guomao Zheng, Erika Schreder, Jennifer C. Dempsey, Nancy Uding, Valerie Chu, Gabriel Andres, Sheela Sathyanarayana, and Amina Salamova*



Cite This: *Environ. Sci. Technol. Lett.* 2021, 8, 224–230



Read Online

ACCESS |



Metrics & More

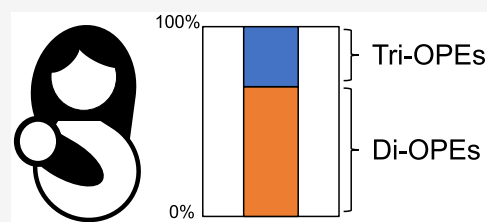


Article Recommendations



Supporting Information

ABSTRACT: Organophosphate esters (OPEs) are among the synthetic chemicals found in the highest concentrations in the indoor environment due to their use as flame retardants and plasticizers. In fish and wildlife, metabolites of OPEs have been found to build up in tissues. In this study, 28 triester OPEs (tri-OPEs) and their seven corresponding diester (di-OPE) and three hydroxyl metabolites were measured in breast milk collected from 50 U.S. mothers. Tris(1-chloro-2-propyl) phosphate, used in foam for insulation and furniture and the target compound with the largest U.S. production volume, was the most abundant tri-OPE (median level of 1.47 ng/mL). Di-*n*-butyl phosphate (DNBP), the metabolite of tri-*n*-butyl phosphate (TNBP), which has broad uses in adhesives, plastics, and hydraulic fluids, was the most abundant OPE metabolite (median level of 7.44 ng/mL) detected in these samples. Overall, the Σ di-OPE concentrations (median level of 8.32 ng/mL) were twice as high as the Σ tri-OPE concentrations (median of 3.85 ng/mL). The estimated daily intakes of tri- and di-OPEs through lactation were up to 50 times higher than those through diet and dust ingestion. This is the first study to simultaneously determine OPEs and their metabolites in breast milk, and our findings indicate that breastfeeding is a significant source of OPE exposure for infants.



INTRODUCTION

Organophosphate esters (OPEs) are man-made industrial chemicals that have been widely used as flame retardants, plasticizers, and antifoaming agents in a range of industrial processes and consumer products for the past several decades.^{1,2} With the phase-out of polybrominated diphenyl ether (PBDE) flame retardants, the level of global consumption of some OPEs [e.g., tris(2-chloroisopropyl) phosphate (TCIPP) and tris(1,3-dichloropropyl) phosphate (TDCIPP)] as common PBDE replacements has increased significantly over the past several years.³ Because OPEs are not chemically bound to products in which they are used, they can evaporate or weather off to the surrounding environment.⁴ As a result, OPEs have become ubiquitous in the environment, including in air,^{5–7} indoor dust,^{8–10} sediment,^{11,12} soil,¹³ water,^{8,14} and wildlife,¹⁵ and humans are exposed through inhalation,^{16,17} dermal exposure,¹⁸ food¹⁹ and water intake,^{20,21} and accidental ingestion of soil or dust.²² OPE exposure has been linked specifically to the presence of flame retardant products in the indoor environment. For example, children living in homes with higher numbers of baby products containing flame retardants have been found to have higher levels of exposure to certain OPEs,²³ and the presence of OPE flame retardants in couches has been associated with levels in house dust.²⁴ In daycares, levels of flame retardants in dust declined after flame retardant nap mats were removed and replaced with a flame retardant-free product.²⁵

A growing body of evidence suggests that exposure to OPE flame retardants is associated with a range of adverse health effects, including cancers and cytotoxicity as well as neurodevelopmental and endocrine disorders.^{2,26–32} Some information about the toxicity of OPE plasticizers is also available. For example, increases in the frequency of liver tumors were found in two-year cancer studies in which mice were exposed to tris(2-ethylhexyl) phosphate (TEHP); the same type of study found increased incidence and severity of urinary bladder tumors upon exposure to tri-*n*-butyl phosphate (TNBP).^{33,34} Laboratory animals exposed to tris(2-butoxyethyl) phosphate (TBOEP) showed effects on the heart and liver as well as on the nervous system, including nerve degeneration and decreased nerve conduction velocity.³⁵

Many OPEs can rapidly metabolize in the human liver with half-lives ranging from several hours to days.³⁶ Some oxidative dealkylation and hydroxylation products of triester OPEs (tri-OPEs), including di- and hydroxyl-OPEs (di-OPEs and OH-OPEs, respectively), have been detected in human urine.^{37–39}

Received: November 24, 2020

Revised: December 23, 2020

Accepted: December 31, 2020

Published: January 6, 2021



Table 1. Detection Frequencies (DF, percent), the Mean [with their standard errors (SE)], Median, Minimum, and Maximum Concentrations of Tri- and Di-OPEs in Breast Milk (nanograms per milliliter; $n = 50$), and Contributions (percent) of Each Individual Compound to the Σ tri-OPE and Σ di-OPE Concentrations

	DF	mean \pm SE	median	minimum	maximum	contribution
Tri-OPEs						
TEP	29	0.133 \pm 0.0210	<0.180	<0.180	1.11	2.9
TNBP	82	0.784 \pm 0.161	0.439	<0.140	5.15	17
TBOEP	73	0.325 \pm 0.0862	0.177	<0.131	3.66	7.1
TEHP	96	0.960 \pm 0.121	0.784	<0.006	3.35	21
TCIPP	82	2.28 \pm 0.385	1.47	<0.455	14.8	50
TCEP	2	0.00585	<0.010	<0.010	0.00585	0.13
TPHP	61	0.0820 \pm 0.0094	0.0494	<0.062	0.241	1.8
EHDP	39	0.0244 \pm 0.00801	<0.001	<0.001	0.341	0.53
Σ tri-OPEs	100	4.59 \pm 0.496	3.85	0.482	16.4	100
Di-OPEs						
DNBP	68	11.7 \pm 1.67	7.44	<3.80	40.5	97
BBOEP	36	0.0816 \pm 0.0300	<0.010	<0.010	1.31	0.68
BDCIPP	28	0.0164 \pm 0.00536	<0.001	<0.001	0.219	0.14
BCIPP	4	0.00139 \pm 0.000629	<0.001	<0.001	0.0256	0.01
DPHP	88	0.211 \pm 0.0368	0.126	<0.103	1.39	1.7
Σ di-OPEs	98	12.1 \pm 1.68	8.32	0.47	41.1	100

Moreover, several di-OPEs such as bis(1,3-dichloroisopropyl) phosphate (BDCIPP) and diphenyl phosphate (DPHP) have been recently found in dust and food,^{40,41} suggesting coexisting exposures to both tri- and di-OPEs. Although relatively little is known about the toxicity of OPE metabolites, some may have stronger toxic effects than their respective parent compounds due to longer half-lives in the body.⁴² Exposure to BDCIPP and DPHP, the metabolites of TDCIPP and triphenyl phosphate (TPHP), respectively, has been associated with poorer semen quality in men.⁴³ *p*-OH-TPHP (*p*-hydroxyl triphenyl phosphate) and DPHP can affect the thyroid by competitively displacing thyroxine from the human trans-thyretin transport protein.⁴⁴ DPHP is more able to alter transcripts in lipid metabolism in chicken embryonic hepatocytes than its parent compound, TPHP.⁴⁵ In addition, DPHP induced dose- and sex-specific perturbation of metabolic profiles in adult mice following neonatal exposure.⁴⁶

Children are generally more susceptible to adverse effects of environmental exposures, including OPEs, due to their rapid development and growth.^{47,48} For infants, breastfeeding is a major exposure pathway to environmental contaminants.^{49–51} Previous studies have reported that early life OPE exposure through lactation is higher than that for PBDEs exposure.^{52–55} In addition, the levels of BDCIPP and DPHP in infants' urine were significantly higher than in adults^{23,56} and breastfed infants had higher levels of the two TCIPP metabolites in urine than older children,⁵⁷ suggesting higher postnatal exposures to OPE metabolites through breastfeeding. However, coexisting exposures to both OPEs and their metabolites in breast milk have not been investigated. In this study, 28 tri-OPEs and their corresponding metabolites, including seven di-OPEs and three OH-OPEs, were simultaneously analyzed in breast milk samples collected from U.S. mothers and daily intakes through breastfeeding were estimated for both compound groups. This is the first study to assess the lactational exposure to OPE metabolites.

MATERIALS AND METHODS

Sample Collection. Breast milk samples ($n = 50$) were collected from breastfeeding primiparous women residing in or

near Seattle, WA, between March and October 2019. Participants were instructed to manually express breastmilk into a provided precleaned glass jar. The samples were picked up from participants within 24 h after collection and stored at $-4\text{ }^{\circ}\text{C}$ until they were shipped to Indiana University, where they were stored at $-20\text{ }^{\circ}\text{C}$ until they were analyzed. All recruitment and sample collection protocols were approved by the Indiana University Institutional Review Board.

Laboratory Analysis. Samples were thawed, spiked with surrogate standards, extracted using ultrasonication, and cleaned on Oasis HLB (for tri-OPEs) and Oasis WAX (for di- and OH-OPEs) cartridges. The samples were analyzed for 28 tri-OPEs, seven di-OPEs, and three OH-OPEs, and all of the analytes are listed in Table S1. The analysis was performed on an ultraperformance liquid chromatograph coupled to a triple-quadrupole mass spectrometer (Agilent 1290 Infinity II UPLC–6470 QQQ-MS). The details of the analytical methods, quality control measures, and data analysis are included in the Supporting Information and in a previous publication.⁵⁸

RESULTS AND DISCUSSION

Population Characteristics. A summary of demographic and behavioral characteristics of the sampled population is provided in Tables S2 and S3. Participants were 24–42 years of age (mean \pm standard deviation: 34 ± 4 years), with average residency in Seattle, WA, of 13 ± 11 years. Ninety-six percent of participants received higher education, and more than half lived in middle or upper-middle class neighborhoods. Fifty-eight percent of women had a body mass index within the normal range of 18.5–24.9, while 36% were overweight or obese.

Concentrations of Tri-OPEs. Eight of the 28 targeted tri-OPEs were detected in breast milk (Table 1 and Figure S1), including four alkyl OPEs [triethyl phosphate (TEP), TNBP, TBOEP, and TEHP], two chlorinated OPEs [TCIPP and tris(2-chloroethyl) phosphate (TCEP)], and two aryl OPEs [TPHP and 2-ethylhexyl diphenyl phosphate (EHDP)]. The total tri-OPE concentrations (Σ tri-OPE, the sum of eight tri-OPE concentrations) ranged from 0.482 to 16.4 ng/mL with a

median concentration of 3.85 ng/mL, which was comparable to that reported in breast milk from Sweden (3.37 ng/g)⁵³ and the United States (3.46 ng/g).⁵⁵ In comparison with other environmental contaminants, the levels of OPEs were higher than those of PBDEs (2.05 ng/g)^{59,60} and other emerging contaminants such as per- and polyfluoroalkyl substances (PFAS) and melamine derivatives detected at a median range of 0.16–2.2 ng/mL in breast milk from the United States.^{61,62}

The flame retardant TCIPP was detected in 82% of the samples at a median concentration of 1.47 ng/mL and was the most abundant tri-OPE, contributing 50% to the Σ tri-OPE concentrations. While TCIPP was also reported as a dominant OPE in breast milk collected between 1997 and 2006 in Sweden (median level of 1.53 ng/mL) and in a small Australian sample from 2015 (5.3–14 ng/mL), it was detected much less frequently in samples from U.S. mothers taken between 2009 and 2012.⁵⁵ In a study measuring TCIPP levels in whole blood and serum as well as metabolites in urine, significant levels of the parent compound were found in serum.⁶³ Taken together, the results suggest that measurement of TCIPP metabolites in urine may not adequately characterize exposure. Interestingly, the other two chlorinated OPEs, TCEP and TDCIPP, were rarely or not detected in any of the samples, with only a single detection of TCEP. Recent studies in the United States,⁵⁵ Spain,⁵⁴ and Vietnam⁵² failed to detect TCEP and TDCIPP, possibly due to a shift in use away from these two designated carcinogens.⁶⁴ Two alkyl-OPE plasticizers, TEHP and TNBP, were detected in most of the samples (96% and 82%, respectively) at median concentrations of 0.784 and 0.439 ng/mL, accounting for 21% and 17% of the Σ tri-OPE concentrations, respectively. These TEHP concentrations were lower than those in breast milk from Australian women (1.2–6.2 ng/mL).⁶⁵ TNBP concentrations in this study were lower than those found in breast milk from Vietnam,⁵² Sweden,⁵³ and Spain⁵⁴ (median levels of 2.00–10.9 ng/mL) but comparable to those from the United States (0.525 ng/mL).⁵⁵ TBOEP, TEP, TPHP, and EHDP were detected at lower concentrations (median levels of <0.001–0.0494 ng/mL), similar to previous reports from Asia, Europe, and the United States.^{52–55}

TNBP, TPHP, TEHP, and TCIPP concentrations were significantly and positively associated with each other, suggesting similar sources for these compounds [$r = 0.37$ – 0.63 ; $p < 0.05$ (Figure S2)]. However, these associations need to be interpreted with caution because of the fast biotransformation rates for some of these OPEs.

Concentrations of OPE Metabolites. Five di-OPEs were detected in these samples, including di-*n*-butyl phosphate (DNBP), bis(2-butoxyethyl) phosphate (BBOEP), BDCIPP, bis(1-chloro-2-propyl) phosphate (BCIPP), and DPHP. None of the targeted OH-OPEs were detected in any of the samples. The five di-OPEs were detected in 4–88% of the samples with the total di-OPE concentrations (Σ di-OPE, the sum of five di-OPE concentrations) ranging from 0.47 to 41.1 ng/mL. The median Σ di-OPE concentration was 8.32 ng/mL, more than twice as high as the median Σ tri-OPE concentration in these samples. DNBP was the most abundant OPE metabolite (median of 7.44 ng/mL) and contributed 97% to the Σ di-OPE concentrations. The rest of the di-OPEs were detected at much lower concentrations. A rapid biotransformation of TNBP to DNBP in liver microsomes in crucian carp,⁶⁶ herring gulls,⁶⁷ and rats⁶⁸ has been previously reported and may explain the relatively high levels of DNBP found in breast milk. For

example, among several targeted tri-OPEs, TNBP was metabolized most quickly in herring gulls' liver microsomes [depletion rate of 73 ± 4 pmol min^{−1} (mg of protein)^{−1}], followed by TBOEP (53 ± 8 pmol min^{−1} mg^{−1}), TCIPP (27 ± 1 pmol min^{−1} mg^{−1}), TPHP (22 ± 2 pmol min^{−1} mg^{−1}), and TDCIPP (8 ± 1 pmol min^{−1} mg^{−1}).⁶⁷ While previous reports of the fast biotransformation of TNBP in wildlife may offer an explanation for the high levels of the metabolite rather than the parent compound detected in breast milk in this study, it should be noted that there is no evidence of the TNBP metabolism pathway in humans and the differences that may exist among species should be considered.^{42,69} Finding high concentrations of DNBP is concerning given the toxicity of the parent compound. TNBP has been listed as a suspected carcinogen by the European Union, and there is evidence of hormone disruption potential.^{70,71} The compound has varied uses that may result in exposure, including as a plasticizer in vinyl, as a solvent in inks, dyes, and adhesives, such as for plywood, in floor waxes and finishes, in paints, in aircraft hydraulic fluid, and in pesticides.^{72,73}

Although no associations were found among the concentrations of di-OPEs, DNBP and DPHP concentrations were significantly and positively correlated with the concentrations of their respective tri-OPE parent compounds, TNBP ($r = 0.45$; $p < 0.05$) and TPHP [$r = 0.37$; $p < 0.05$ (Figure S2)], suggesting similar sources for the parent compounds and their respective metabolites and/or substantial metabolism to the di-OPE. Significant associations have also been previously observed for tri- and di-OPE concentrations in fish.⁵⁸

Parent:Metabolite Concentration Ratios. A relative abundance of each di-OPE was calculated as a ratio of the molar concentration of this di-OPE and the sum of the molar concentrations of that di-OPE and its parent tri-OPE [di-OPE/(di-OPE + tri-OPE)] (Figure 1). The relative abundance

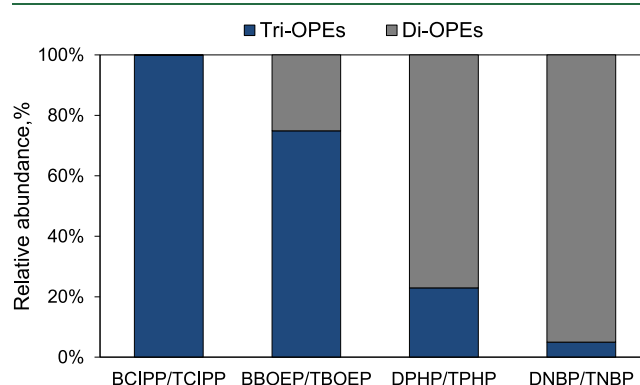


Figure 1. Relative percent abundances of tri-OPEs and their respective di-OPE metabolites in breast milk (based on molar concentrations).

ratios can provide critical information about the metabolic rate of a parent compound and the stability of a metabolite.^{25,58} Four pairs of the parent tri-OPEs and their respective di-OPE metabolites are discussed here, including BCIPP and TCIPP, BBOEP and TBOEP, DPHP and TPHP, and DNBP and TNBP. BCIPP, a metabolite of TCIPP, and BBOEP, a metabolite of TBOEP, had relative abundances of 0.10% and 25%, respectively, indicating that the tri-OPE parent compound is more abundant than its di-OPE metabolite. The predominance of TCIPP can be attributed to its slow biotransformation in humans⁷⁴ and its increased usage in the

U.S. market.⁵⁷ In contrast, TBOEP metabolizes rapidly in humans (half-life of <4 h),⁷⁵ suggesting that an alternative metabolism pathway resulting in a metabolite other than BBOEP may dominate. In an *in vitro* study using human liver and serum enzymes, TBOEP metabolized to several hydroxylated metabolites, such as bis(2-butoxyethyl) hydroxyethyl phosphate (BBOEHEP), which was more abundant than BBOEP.⁷⁶ Urinary BBOEHEP concentrations were at least 2 orders of magnitude higher than those of BBOEP when TBOEP was administered orally.⁷⁵ Compared to BCIPP and BBOEP, DPHP and DNBP (the metabolites of TPHP and TNBP, respectively) had higher relative abundances of 77% and 95%, respectively. These findings suggest that TPHP and TNBP can be metabolized quickly to DPHP and DNBP, respectively, which are then likely to be transferred from maternal blood to breast milk. However, it should be noted that other OPEs such as resorcinol bis(diphenyl phosphate) and EHDP can also metabolize to DPHP, and TPHP may not be the only source of this di-OPE. The higher abundance of OPE metabolites in breast milk can increase their lactational intake for infants and may offer an explanation for the previously found high DPHP and BDCIPP concentrations in infants' urine.^{23,56}

Relationships with Behavioral and Demographic Characteristics. Significant negative correlations were observed among TNBP, TEHP, DNBP, Σ di-OPE, and Σ tri-OPE concentrations in breast milk and maternal age [$r = -0.463$ to -0.298 ; $p < 0.05$ (Table S4)]. Somewhat higher Σ tri-OPE concentrations were observed among women with lower handwashing frequency, although the difference was not significant (Figure S3). No significant associations were found between tri- and di-OPEs levels and other demographic factors.

Exposure Assessment. Lactational estimated daily intake (EDI) rates of tri- and di-OPEs were calculated for infants between the ages of 0 and 12 months using established lactation rates for each age group⁷⁷ (Table S5). The highest Σ tri-OPE EDI was observed for infants who were <1 month of age [$689 \text{ ng [kg of body weight (bw)]}^{-1} \text{ day}^{-1}$], followed by 1–3 months of age [$643 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$], 3–6 months of age [$505 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$], and 6–12 months of age [$381 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$], similar to previous studies.⁵⁵ The lactational Σ tri-OPE EDI was higher than that via accidental dust ingestion,²² dermal absorption,⁷⁸ and inhalation,⁷⁸ indicating that breastfeeding is a significant source of OPE exposure for infants. The highest intake was estimated for TCIPP [$189\text{--}342 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$], which was up to 50 times higher than that for other tri-OPEs.

The Σ di-OPE EDI was $1820 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$ for infants who were <1 month of age, which was up to 3 times higher than that for Σ tri-OPEs. The Σ di-OPE EDIs via dietary intake for adults⁴⁰ and accidental dust ingestion for toddlers⁴¹ were up to 50 times lower [130 and $54.4 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$, respectively] than the lactational EDIs. Lactational intake of DNBP reached $1760 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$. These findings are concerning as DNBP exposure has been associated with chronic kidney disease.⁷⁹ Dust ingestion EDIs of di-OPEs for infants were also estimated on the basis of a previous study that measured di-OPEs in indoor dust (details for these calculations are included in the Supporting Information).⁴¹ The EDIs of BBOEP and BDCIPP via dust ingestion ranged from 3.43 to $6.58 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$ and from 0.51 to $0.98 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$ for infants who were 0–12 months of age, respectively (Table S6),⁴¹ which were 2–

7 times higher than those through lactation. The DPHP EDI through dust ingestion ranged from 14.1 to $27.1 \text{ ng (kg of bw)}^{-1} \text{ day}^{-1}$ (Table S6), which was comparable to the lactational intake.

This is the first study to simultaneously determine OPEs and their metabolites in breast milk. The results highlight considerable coexisting exposures to OPE flame retardants and plasticizers and their metabolites with significant toxicity concerns ranging from cancer to nervous system effects, disruption of thyroid, and lipid metabolism and suggest that breastfeeding is a significant exposure pathway to these compounds.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.estlett.0c00916>.

List of target analytes; details of sampling, data and instrumental analyses, and quality control measures; demographic and behavioral characteristics of participants; results of the Spearman correlations between chemical concentrations in breast milk and various demographic and behavioral characteristics; distribution of tri- and di-OPE concentrations in breast milk; estimated lactational daily intake rates; estimated daily intake rates for di-OPE via dust ingestion; and Pearson correlation matrix for tri- and di-OPE concentrations (PDF)

■ AUTHOR INFORMATION

Corresponding Author

Amina Salamova – Paul H. O'Neill School of Public and Environmental Affairs, Indiana University, Bloomington, Indiana 47405, United States; orcid.org/0000-0003-2174-030X; Email: asalamov@indiana.edu

Authors

Guomao Zheng – Paul H. O'Neill School of Public and Environmental Affairs, Indiana University, Bloomington, Indiana 47405, United States; orcid.org/0000-0002-5235-9950

Erika Schreder – Toxic-Free Future, Seattle, Washington 98103, United States

Jennifer C. Dempsey – Toxic-Free Future, Seattle, Washington 98103, United States

Nancy Uding – Toxic-Free Future, Seattle, Washington 98103, United States

Valerie Chu – Toxic-Free Future, Seattle, Washington 98103, United States

Gabriel Andres – Toxic-Free Future, Seattle, Washington 98103, United States

Sheela Sathyanarayana – Department of Pediatrics, University of Washington/Seattle Children's Research Institute, Seattle, Washington 98107, United States

Complete contact information is available at: <https://pubs.acs.org/doi/10.1021/acs.estlett.0c00916>

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

The authors thank the participants for their generosity in sharing their time and breast milk for this study. Carina Wells and Mikyla Sakurai provided significant contributions in recruiting participants and collecting samples. The authors thank Yan Wu and Marta Venier for their contributions to the development of the analytical methods and Duke University for helping to obtain [^{13}C]DPHP.

■ REFERENCES

- (1) van der Veen, I.; de Boer, J. Phosphorus flame retardants: properties, production, environmental occurrence, toxicity and analysis. *Chemosphere* **2012**, *88*, 1119–53.
- (2) Wei, G. L.; Li, D. Q.; Zhuo, M. N.; Liao, Y. S.; Xie, Z. Y.; Guo, T. L.; Li, J. J.; Zhang, S. Y.; Liang, Z. Q. Organophosphorus flame retardants and plasticizers: Sources, occurrence, toxicity and human exposure. *Environ. Pollut.* **2015**, *196*, 29–46.
- (3) Liu, R.; Mabury, S. A. Unexpectedly high concentrations of a newly identified organophosphate ester, tris(2,4-di-tert-butylphenyl) phosphate, in indoor dust from Canada. *Environ. Sci. Technol.* **2018**, *52*, 9677–9683.
- (4) Zhao, L.; Jian, K.; Su, H.; Zhang, Y.; Li, J.; Letcher, R. J.; Su, G. Organophosphate esters (OPEs) in Chinese foodstuffs: Dietary intake estimation via a market basket method, and suspect screening using high-resolution mass spectrometry. *Environ. Int.* **2019**, *128*, 343–352.
- (5) Okeme, J. O.; Yang, C.; Abdollahi, A.; Dhal, S.; Harris, S. A.; Jantunen, L. M.; Tsirlin, D.; Diamond, M. L. Passive air sampling of flame retardants and plasticizers in Canadian homes using PDMS, XAD-coated PDMS and PUF samplers. *Environ. Pollut.* **2018**, *239*, 109–117.
- (6) Zhou, L.; Hiltcher, M.; Gruber, D.; Püttmann, W. Organophosphate flame retardants (OPFRs) in indoor and outdoor air in the Rhine/Main area, Germany: Comparison of concentrations and distribution profiles in different microenvironments. *Environ. Sci. Pollut. Res.* **2017**, *24*, 10992–11005.
- (7) Shoeib, M.; Ahrens, L.; Jantunen, L.; Harner, T. Concentrations in air of organobromine, organochlorine and organophosphate flame retardants in Toronto, Canada. *Atmos. Environ.* **2014**, *99*, 140–147.
- (8) Harrad, S.; Brommer, S.; Mueller, J. F. Concentrations of organophosphate flame retardants in dust from cars, homes, and offices: An international comparison. *Emerg. Contam.* **2016**, *2*, 66–72.
- (9) Ali, N.; Eqani, S. A. M. A. S.; Ismail, I. M. I.; Malarvannan, G.; Kadi, M. W.; Albar, H. M. S.; Rehan, M.; Covaci, A. Brominated and organophosphate flame retardants in indoor dust of Jeddah, Kingdom of Saudi Arabia: Implications for human exposure. *Sci. Total Environ.* **2016**, *569–570*, 269–277.
- (10) Abdallah, M. A.-E.; Covaci, A. Organophosphate flame retardants in indoor dust from Egypt: Implications for human exposure. *Environ. Sci. Technol.* **2014**, *48*, 4782–4789.
- (11) Zhong, M.; Wu, H.; Mi, W.; Li, F.; Ji, C.; Ebinghaus, R.; Tang, J.; Xie, Z. Occurrences and distribution characteristics of organophosphate ester flame retardants and plasticizers in the sediments of the Bohai and Yellow Seas, China. *Sci. Total Environ.* **2018**, *615*, 1305–1311.
- (12) Cao, D.; Guo, J.; Wang, Y.; Li, Z.; Liang, K.; Corcoran, M. B.; Hosseini, S.; Bonina, S. M. C.; Rockne, K. J.; Sturchio, N. C.; Giesy, J. P.; Liu, J.; Li, A.; Jiang, G. Organophosphate esters in sediment of the Great Lakes. *Environ. Sci. Technol.* **2017**, *51*, 1441–1449.
- (13) Kurt-Karakus, P.; Alegria, H.; Birgul, A.; Gungormus, E.; Jantunen, L. Organophosphate ester (OPEs) flame retardants and plasticizers in air and soil from a highly industrialized city in Turkey. *Sci. Total Environ.* **2018**, *625*, 555–565.
- (14) Li, J.; Yu, N.; Zhang, B.; Jin, L.; Li, M.; Hu, M.; Zhang, X.; Wei, S.; Yu, H. Occurrence of organophosphate flame retardants in drinking water from China. *Water Res.* **2014**, *54*, 53–61.
- (15) Guo, J.; Venier, M.; Salamova, A.; Hites, R. A. Bioaccumulation of Dechloranes, organophosphate esters, and other flame retardants in Great Lakes fish. *Sci. Total Environ.* **2017**, *583*, 1–9.
- (16) Cao, D.; Lv, K.; Gao, W.; Fu, J.; Wu, J.; Fu, J.; Wang, Y.; Jiang, G. Presence and human exposure assessment of organophosphate flame retardants (OPEs) in indoor dust and air in Beijing, China. *Ecotoxicol. Environ. Saf.* **2019**, *169*, 383–391.
- (17) Schreder, E. D.; Uding, N.; La Guardia, M. J. Inhalation a significant exposure route for chlorinated organophosphate flame retardants. *Chemosphere* **2016**, *150*, 499–504.
- (18) Phillips, A. L.; Hammel, S. C.; Hoffman, K.; Lorenzo, A. M.; Chen, A.; Webster, T. F.; Stapleton, H. M. Children's residential exposure to organophosphate ester flame retardants and plasticizers: Investigating exposure pathways in the TESIE study. *Environ. Int.* **2018**, *116*, 176–185.
- (19) Li, J.; Zhao, L.; Letcher, R. J.; Zhang, Y.; Jian, K.; Zhang, J.; Su, G. A review on organophosphate ester (OPE) flame retardants and plasticizers in foodstuffs: Levels, distribution, human dietary exposure, and future directions. *Environ. Int.* **2019**, *127*, 35–51.
- (20) Li, J.; He, J.; Li, Y.; Liu, Y.; Li, W.; Wu, N.; Zhang, L.; Zhang, Y.; Niu, Z. Assessing the threats of organophosphate esters (flame retardants and plasticizers) to drinking water safety based on USEPA oral reference dose (RfD) and oral cancer slope factor (SFO). *Water Res.* **2019**, *154*, 84–93.
- (21) Kim, U.-J.; Kannan, K. Occurrence and distribution of organophosphate flame retardants/plasticizers in surface waters, tap water, and rainwater: Implications for human exposure. *Environ. Sci. Technol.* **2018**, *52*, S625–S633.
- (22) Li, W.; Wang, Y.; Asimakopoulos, A. G.; Covaci, A.; Gevao, B.; Johnson-Restrepo, B.; Kumosani, T. A.; Malarvannan, G.; Moon, H.-B.; Nakata, H.; Sinha, R. K.; Tran, T. M.; Kannan, K. Organophosphate esters in indoor dust from 12 countries: Concentrations, composition profiles, and human exposure. *Environ. Int.* **2019**, *133*, 105178.
- (23) Hoffman, K.; Butt, C. M.; Chen, A.; Limkakeng, A. T., Jr.; Stapleton, H. M. High exposure to organophosphate flame retardants in infants: Associations with baby products. *Environ. Sci. Technol.* **2015**, *49*, 14554–9.
- (24) Hammel, S. C.; Hoffman, K.; Lorenzo, A. M.; Chen, A.; Phillips, A. L.; Butt, C. M.; Sosa, J. A.; Webster, T. F.; Stapleton, H. M. Associations between flame retardant applications in furniture foam, house dust levels, and residents' serum levels. *Environ. Int.* **2017**, *107*, 181–189.
- (25) Stubbings, W. A.; Guo, J.; Simon, K.; Romanak, K.; Bowerman, W.; Venier, M. Flame retardant metabolites in addled bald eagle eggs from the Great Lakes region. *Environ. Sci. Technol. Lett.* **2018**, *5*, 354–359.
- (26) Ta, N.; Li, C.; Fang, Y.; Liu, H.; Lin, B.; Jin, H.; Tian, L.; Zhang, H.; Zhang, W.; Xi, Z. Toxicity of TDCPP and TCEP on PC12 cell: changes in CAMKII, GAP43, tubulin and NF-H gene and protein levels. *Toxicol. Lett.* **2014**, *227*, 164–71.
- (27) Meeker, J. D.; Stapleton, H. M. House dust concentrations of organophosphate flame retardants in relation to hormone levels and semen quality parameters. *Environ. Health Perspect.* **2010**, *118*, 318–323.
- (28) Castorina, R.; Bradman, A.; Stapleton, H. M.; Butt, C.; Avery, D.; Harley, K. G.; Gunier, R. B.; Holland, N.; Eskenazi, B. Current-use flame retardants: Maternal exposure and neurodevelopment in children of the CHAMACOS cohort. *Chemosphere* **2017**, *189*, 574–580.
- (29) Doherty, B. T.; Hammel, S. C.; Daniels, J. L.; Stapleton, H. M.; Hoffman, K. Organophosphate esters: Are these flame retardants and plasticizers affecting children's health? *Curr. Environ. Health Rep.* **2019**, *6*, 201–213.
- (30) Dishaw, L. V.; Hunter, D. L.; Padnos, B.; Padilla, S.; Stapleton, H. M. Developmental exposure to organophosphate flame retardants elicits overt toxicity and alters behavior in early life stage zebrafish (*Danio rerio*). *Toxicol. Sci.* **2014**, *142*, 445–454.
- (31) Farhat, A.; Crump, D.; Chiu, S.; Williams, K. L.; Letcher, R. J.; Gauthier, L. T.; Kennedy, S. W. Ovo effects of two organophosphate flame retardants—TCPP and TDCPP—on pipping success, develop-

ment, mRNA expression, and thyroid hormone levels in chicken embryos. *Toxicol. Sci.* **2013**, *134*, 92–102.

(32) Crump, D.; Chiu, S.; Kennedy, S. W. Effects of tris(1,3-dichloro-2-propyl) phosphate and tris(1-chloropropyl) phosphate on cytotoxicity and mRNA expression in primary cultures of avian hepatocytes and neuronal cells. *Toxicol. Sci.* **2012**, *126*, 140–148.

(33) Auletta, C. S.; Weiner, M. L.; Richter, W. R. A dietary toxicity/oncogenicity study of tributyl phosphate in the rat. *Toxicology* **1998**, *128*, 125–134.

(34) National Toxicology Program. Toxicology and carcinogenesis studies of tris(2-ethylhexyl)phosphate in F344/N rats and B6C3F1 mice (gavage studies). 1984. https://ntp.niehs.nih.gov/publications/reports/tr/200s/tr274/index.html?utm_source=direct&utm_medium=prod&utm_campaign=ntpgoilinks&utm_term=tr274abs (accessed 2020-11-11).

(35) World Health Organization. Flame retardants: tris(2-butoxyethyl) phosphate, tris(2-ethylhexyl) phosphate, tetrakis(hydroxymethyl) phosphonium salts. 2000. <https://www.who.int/ipcs/publications/ehc/en/EHC218.pdf> (accessed 2020-11-11).

(36) Wang, X.; Liu, Q.; Zhong, W.; Yang, L.; Yang, J.; Covaci, A.; Zhu, L. Estimating renal and hepatic clearance rates of organophosphate esters in humans: Impacts of intrinsic metabolism and binding affinity with plasma proteins. *Environ. Int.* **2020**, *134*, 105321.

(37) Guo, J.; Simon, K.; Romanak, K.; Bowerman, W.; Venier, M. Accumulation of flame retardants in paired eggs and plasma of bald eagles. *Environ. Pollut.* **2018**, *237*, 499–507.

(38) Cequier, E.; Marce, R. M.; Becher, G.; Thomsen, C. Comparing human exposure to emerging and legacy flame retardants from the indoor environment and diet with concentrations measured in serum. *Environ. Int.* **2015**, *74*, 54–9.

(39) Hammel, S. C.; Stapleton, H. M.; Eichner, B.; Hoffman, K. Reconsidering an appropriate urinary biomarker for flame retardant tris(1-chloro-2-propyl) phosphate (TCIPP) exposure in children. *Environ. Sci. Technol. Lett.* **2020**, DOI: 10.1021/acs.estlett.0c00794.

(40) He, C.; Wang, X.; Tang, S.; Thai, P.; Li, Z.; Baduel, C.; Mueller, J. F. Concentrations of organophosphate esters and their specific metabolites in food in southeast queensland, Australia: Is dietary exposure an important pathway of organophosphate esters and their metabolites? *Environ. Sci. Technol.* **2018**, *52*, 12765–12773.

(41) Tan, H.; Yang, L.; Yu, Y.; Guan, Q.; Liu, X.; Li, L.; Chen, D. Co-existence of organophosphate di- and tri-esters in house dust from South China and midwestern United States: Implications for human exposure. *Environ. Sci. Technol.* **2019**, *53*, 4784–4793.

(42) Hou, R.; Xu, Y.; Wang, Z. Review of OPFRs in animals and humans: Absorption, bioaccumulation, metabolism, and internal exposure research. *Chemosphere* **2016**, *153*, 78–90.

(43) Meeker, J. D.; Cooper, E. M.; Stapleton, H. M.; Hauser, R. Exploratory analysis of urinary metabolites of phosphorus-containing flame retardants in relation to markers of male reproductive health. *Endocr. Disruptors* **2013**, *1*, No. e26306.

(44) Hill, K. L.; Hamers, T.; Kamstra, J. H.; Willmore, W. G.; Letcher, R. J. Organophosphate triesters and selected metabolites enhance binding of thyroxine to human transthyretin in vitro. *Toxicol. Lett.* **2018**, *285*, 87–93.

(45) Su, G.; Letcher, R. J.; Crump, D.; Gooden, D. M.; Stapleton, H. M. In vitro metabolism of the flame retardant triphenyl phosphate in chicken embryonic hepatocytes and the importance of the hydroxylation pathway. *Environ. Sci. Technol. Lett.* **2015**, *2*, 100–104.

(46) Wang, D.; Zhu, W.; Chen, L.; Yan, J.; Teng, M.; Zhou, Z. Neonatal triphenyl phosphate and its metabolite diphenyl phosphate exposure induce sex- and dose-dependent metabolic disruptions in adult mice. *Environ. Pollut.* **2018**, *237*, 10–17.

(47) Doherty, B. T.; Hoffman, K.; Keil, A. P.; Engel, S. M.; Stapleton, H. M.; Goldman, B. D.; Olshan, A. F.; Daniels, J. L. Prenatal exposure to organophosphate esters and behavioral development in young children in the pregnancy, infection, and nutrition study. *Neurotoxicology* **2019**, *73*, 150–160.

(48) Hoffman, K.; Stapleton, H. M.; Lorenzo, A.; Butt, C. M.; Adair, L.; Herring, A. H.; Daniels, J. L. Prenatal exposure to organo-

phosphates and associations with birthweight and gestational length. *Environ. Int.* **2018**, *116*, 248–254.

(49) LaKind, J. S.; Lehmann, G. M.; Davis, M. H.; Hines, E. P.; Marchitti, S. A.; Alcalá, C.; Lorber, M. Infant dietary exposures to environmental chemicals and infant/child health: A critical assessment of the literature. *Environ. Health Perspect.* **2018**, *126*, 096002.

(50) Mondal, D.; Weldon, R. H.; Armstrong, B. G.; Gibson, L. J.; Lopez-Espinosa, M. J.; Shin, H. M.; Fletcher, T. Breastfeeding: A potential excretion route for mothers and implications for infant exposure to perfluoroalkyl acids. *Environ. Health Perspect.* **2014**, *122*, 187–92.

(51) Lehmann, G. M.; LaKind, J. S.; Davis, M. H.; Hines, E. P.; Marchitti, S. A.; Alcalá, C.; Lorber, M. Environmental chemicals in breast milk and formula: Exposure and risk assessment implications. *Environ. Health Perspect.* **2018**, *126*, 096001.

(52) Kim, J. W.; Isobe, T.; Muto, M.; Tue, N. M.; Katsura, K.; Malarvannan, G.; Sudaryanto, A.; Chang, K. H.; Prudente, M.; Viet, P. H.; Takahashi, S.; Tanabe, S. Organophosphorus flame retardants (PFRs) in human breast milk from several Asian countries. *Chemosphere* **2014**, *116*, 91–7.

(53) Sundkvist, A. M.; Olofsson, U.; Haglund, P. Organophosphorus flame retardants and plasticizers in marine and fresh water biota and in human milk. *J. Environ. Monit.* **2010**, *12*, 943–51.

(54) Beser, M. I.; Pardo, O.; Beltran, J.; Yusa, V. Determination of 21 perfluoroalkyl substances and organophosphorus compounds in breast milk by liquid chromatography coupled to orbitrap high-resolution mass spectrometry. *Anal. Chim. Acta* **2019**, *1049*, 123–132.

(55) Ma, J.; Zhu, H.; Kannan, K. Organophosphorus flame retardants and plasticizers in breast milk from the United States. *Environ. Sci. Technol. Lett.* **2019**, *6*, 525–531.

(56) Hoffman, K.; Gearhart-Serna, L.; Lorber, M.; Webster, T. F.; Stapleton, H. M. Estimated tris(1,3-dichloro-2-propyl) phosphate exposure levels for U.S. infants suggest potential health risks. *Environ. Sci. Technol. Lett.* **2017**, *4*, 334–338.

(57) Hammel, S. C.; Zhang, S.; Lorenzo, A. M.; Eichner, B.; Stapleton, H. M.; Hoffman, K. Young infants' exposure to organophosphate esters: Breast milk as a potential source of exposure. *Environ. Int.* **2020**, *143*, 106009.

(58) Zheng, G.; Miller, P.; von Hippel, F. A.; Buck, C. L.; Carpenter, D. O.; Salamova, A. Legacy and emerging semi-volatile organic compounds in sentinel fish from an arctic formerly used defense site in Alaska. *Environ. Pollut.* **2020**, *259*, 113872.

(59) She, J.; Holden, A.; Sharp, M.; Tanner, M.; Williams-Derry, C.; Hooper, K. Polybrominated diphenyl ethers (PBDEs) and polychlorinated biphenyls (PCBs) in breast milk from the Pacific Northwest. *Chemosphere* **2007**, *67*, S307–S317.

(60) Marchitti, S. A.; Fenton, S. E.; Mendola, P.; Kenneke, J. F.; Hines, E. P. Polybrominated diphenyl ethers in human milk and serum from the U.S. EPA MAMA study: Modeled predictions of infant exposure and considerations for risk assessment. *Environ. Health Perspect.* **2017**, *125*, 706–713.

(61) Tao, L.; Kannan, K.; Wong, C. M.; Arcaro, K. F.; Butenhoff, J. L. Perfluorinated compounds in human milk from Massachusetts. *Environ. Sci. Technol.* **2008**, *42*, 3096–3101.

(62) Zhu, H.; Kannan, K. Occurrence of melamine and its derivatives in breast milk from the United States and its implications for exposure in infants. *Environ. Sci. Technol.* **2019**, *53*, 7859–7865.

(63) Hou, M.; Shi, Y.; Jin, Q.; Cai, Y. Organophosphate esters and their metabolites in paired human whole blood, serum, and urine as biomarkers of exposure. *Environ. Int.* **2020**, *139*, 105698.

(64) Dodson, R. E.; Perovich, L. J.; Covaci, A.; Van den Eede, N.; Ionas, A. C.; Dittu, A. C.; Brody, J. G.; Rudel, R. A. After the PBDE phase-out: A broad suite of flame retardants in repeat house dust samples from California. *Environ. Sci. Technol.* **2012**, *46*, 13056–13066.

(65) He, C.; Toms, L.-M. L.; Thai, P.; Van den Eede, N.; Wang, X.; Li, Y.; Baduel, C.; Harden, F. A.; Heffernan, A. L.; Hobson, P.; Covaci, A.; Mueller, J. F. Urinary metabolites of organophosphate

esters: Concentrations and age trends in Australian children. *Environ. Int.* **2018**, *111*, 124–130.

(66) Hou, R.; Huang, C.; Rao, K.; Xu, Y.; Wang, Z. Characterized in vitro metabolism kinetics of alkyl organophosphate esters in fish liver and intestinal microsomes. *Environ. Sci. Technol.* **2018**, *52*, 3202–3210.

(67) Greaves, A. K.; Su, G.; Letcher, R. J. Environmentally relevant organophosphate triesters in herring gulls: In vitro biotransformation and kinetics and diester metabolite formation using a hepatic microsomal assay. *Toxicol. Appl. Pharmacol.* **2016**, *308*, 59–65.

(68) Sasaki, K.; Suzuki, T.; Takeda, M.; Uchiyama, M. Metabolism of phosphoric acid triesters by rat liver homogenate. *Bull. Environ. Contam. Toxicol.* **1984**, *33*, 281–288.

(69) Van den Eede, N.; Maho, W.; Erratico, C.; Neels, H.; Covaci, A. First insights in the metabolism of phosphate flame retardants and plasticizers using human liver fractions. *Toxicol. Lett.* **2013**, *223*, 9–15.

(70) Kojima, H.; Takeuchi, S.; Itoh, T.; Iida, M.; Kobayashi, S.; Yoshida, T. In vitro endocrine disruption potential of organophosphate flame retardants via human nuclear receptors. *Toxicology* **2013**, *314*, 76–83.

(71) European Chemical Agency. Brief profile: Tributyl phosphate. 2016. <https://echa.europa.eu/brief-profile/-/briefprofile/100.004.365> (accessed 2020-11-11).

(72) Agency for Toxic Substances and Disease Registry. Toxicological profile for phosphate ester flame retardants. 2012. <https://www.atsdr.cdc.gov/ToxProfiles/tp202.pdf> (accessed 2020-11-11).

(73) PubChem. Compound summary: Tributyl phosphate. 2020. <https://pubchem.ncbi.nlm.nih.gov/compound/Tributyl-phosphate> (accessed 2020-11-11).

(74) Van den Eede, N.; Tomy, G.; Tao, F.; Halldorson, T.; Harrad, S.; Neels, H.; Covaci, A. Kinetics of tris (1-chloro-2-propyl) phosphate (TCIPP) metabolism in human liver microsomes and serum. *Chemosphere* **2016**, *144*, 1299–1305.

(75) Völkel, W.; Fuchs, V.; Wöckner, M.; Fromme, H. Toxicokinetic of tris(2-butoxyethyl) phosphate (TBOEP) in humans following single oral administration. *Arch. Toxicol.* **2018**, *92*, 651–660.

(76) Van den Eede, N.; Erratico, C.; Exarchou, V.; Maho, W.; Neels, H.; Covaci, A. In vitro biotransformation of tris(2-butoxyethyl) phosphate (TBOEP) in human liver and serum. *Toxicol. Appl. Pharmacol.* **2015**, *284*, 246–253.

(77) *Exposure factors handbook*; U.S. Environmental Protection Agency: Washington, DC, 2011.

(78) He, C.; Wang, X.; Thai, P.; Baduel, C.; Gallen, C.; Banks, A.; Bainton, P.; English, K.; Mueller, J. F. Organophosphate and brominated flame retardants in Australian indoor environments: Levels, sources, and preliminary assessment of human exposure. *Environ. Pollut.* **2018**, *235*, 670–679.

(79) Kang, H.; Lee, J.; Lee, J. P.; Choi, K. Urinary metabolites of organophosphate esters (OPEs) are associated with chronic kidney disease in the general US population, NHANES 2013–2014. *Environ. Int.* **2019**, *131*, 105034.